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Xin Yang, Songjing Duan, Mei Li, Dongxu Li, Rongcong Yang, Shihui Zhang, Taimin Xu, Wen Li, Hongmin Zhou, Changlin Zhao

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Research Article

A new genus and three new species of Lycoperdaceae (Agaricales) from Southern China revealed by molecular phylogeny and taxonomy

Xin Yang[1,2], Songjing Duan[2], Mei Li[2], Dongxu Li[2], Rongcong Yang[3], Shihui Zhang[3], Taimin Xu[4], Wen Li[1,2], Hongmin Zhou[1,2], Changlin Zhao[1,2]

1 *Yunnan Provincial Key Laboratory for Conservation and Utilization of In-forest Resource, the Key Laboratory of Forest Resources Conservation and Utilization in the Southwest Mountains of China, Ministry of Education, Southwest Forestry University, Kunming 650224, China*
2 *College of Forestry, Southwest Forestry University, Kunming 650224, China*
3 *Yunnan Tongbiguan Provincial Nature Reserve, Mangshi, 678400, China*
4 *College of Agriculture and Life Sciences, Zhaotong University, Zhaotong 657000, China*
Corresponding author: Changlin Zhao (fungi@swfu.edu.cn; fungichanglinz@163.com)

Abstract

Lycoperdaceae (Agaricales, Agaricomycetes) is a taxonomically significant fungal family based on the globally distributed and morphologically defined puffball structures. In the present study, one new genus, *Lycoperdia*, and three new species, viz. *Calvatia phlebioides*, *Lycoperdia tomentosa*, and *Morganella minima*, collected from southern China are proposed based on a combination of morphological characteristics and molecular evidence. *Calvatia phlebioides* is characterized by broadly obpyriform to turbinate basidiomes and globose to subglobose basidiospores with a short pedicel. *Lycoperdia tomentosa* is characterized by pyriform basidiomes, tomentose exoperidium, and globose to subglobose basidiospores with a long pedicel. *Morganella minima* is characterized by tiny basidiomes (5–8 mm in diameter, 3–7 mm in height), globose to subglobose basidiospores with distinct spines, and a short pedicel (<0.5 µm). Sequences of the internal transcribed spacers (ITS), nuclear large subunit ribosomal RNA (nLSU), and RNA polymerase second largest subunit (*rpb2*) of the nuclear ribosomal DNA (rDNA) markers of the studied samples were generated, and the phylogenetic analyses were performed with maximum likelihood and Bayesian inference methods. The results showed that our collections were clustered within the family Lycoperdaceae. The phylogenetic tree inferred from the ITS+nLSU sequences revealed that all four new taxa were clustered into the family Lycoperdaceae, in which *Lycoperdia tomentosa* formed a monophyletic lineage. The phylogenetic tree inferred from the ITS+nLSU+*rpb2* sequences revealed that one new taxon, *Calvatia phlebioides*, was clustered into the genus *Calvatia*, sister to *C. longisetulosa*. The phylogenetic tree inferred from the ITS+nLSU sequences revealed that *Morganella minima* was clustered into the genus *Morganella*, forming a monophyletic lineage. Full morphological descriptions, illustrations, and phylogenetic analysis results for the new genus and three new species are provided.

Key words: Classification, molecular systematics, new taxa, taxonomy, puffballs

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Introduction

The global One Health approach seeks to better understand and improve the interconnectedness of people, animals, plants, and their environment, and such insight will engender a more sustainable future for life on earth, and fungi are essential to this quest because they affect earth's ecosystems in a myriad of beneficial and detrimental ways (Zhao et al. 2024; Case et al. 2025). Fungi are among the most diverse groups of organisms on this planet and play a core role in ecosystem processes and functioning (Wei and Dai 2004; Cui et al. 2018; Hyde 2022; Dong et al. 2024a; Zhou et al. 2025). Currently, 19 phyla of fungi are accepted: Aphelidiomycota, Ascomycota, Basidiobolomycota, Basidiomycota, Blastocladiomycota, Calcarisporiellomycota, Chytridiomycota, Entomophthoromycota, Entorrhizomycota, Glomeromycota, Kickxellomycota, Monoblepharomycota, Mortierellomycota, Mucoromycota, Neocallimastigomycota, Olpidiomycota, Rozellomycota, Sanchytriomycota, and Zoopagomycota (Zhao et al. 2023b; Wijayawardene et al. 2024). Fungi secrete a spectacular array of bioactive chemical compounds and enzymes; these have crucial roles in the biosphere, from digesting organic matter and recycling nutrients from dead plant and animal tissues to mediating intimate and mutually beneficial associations with the roots of almost all land plants (Wu et al. 2019; Case et al. 2025), in which most Basidiomycota species act as decomposers and mutualists of plants and animals, which play fundamental ecological roles such as driving carbon cycling in forest soils (Tedersoo et al. 2014; Cui et al. 2019; Yuan et al. 2023b).

Lycoperdaceae F. Berchtold & J. Presl is commonly known as puffballs, and species in the family are characterized by the globose to subglobose basidiomes and mature gleba, releasing a powdery mass of spores passively through the apical opening of the endoperidium (Cunningham 1926b; Kreisel 1962; Krüger et al. 2001). These species are widely distributed in temperate, arid, and tropical climates, found on the forest floor, rotting wood, or open meadows (Pegler et al. 1995). Some species within this family have culinary and medicinal values, such as *Calvatia craniiformis* (Schwein.) Fr. ex De Toni and *C. cyathiformis* (Bosc) Morgan are used as food in the UK, while *C. gigantea* (Batsch) Lloyd and *C. lilacina* (Mont. & Berk.) Henn are used in traditional medicine for their hemostatic properties and to treat various ailments (Coetzee and van Wyk 2009).

Lycoperdaceae was established by F. Berchtold and J. Presl in 1820, originally encompassed 18 genera and 150 species, and initially this family was considered synonymous with Agaricaceae (Kirk et al. 2008); later, the molecular phylogenetic analyses revealed this family, Lycoperdaceae, to be a monophyletic gasteroid lineage distinct from Agaricaceae (He et al. 2019). Subsequent studies identified four main subclades within Lycoperdaceae as the genera *Bovista* Pers., *Calvatia* Fr., *Disciseda* Czern., and *Lycoperdon* Pers. *sensu lato* (Larsson and Jeppson 2008). In the recent taxonomic system of Basidiomycota, Lycoperdaceae was treated as a distinct family comprising the genera *Apioperdon* (Kreisel & D. Krüger) Vizzini, *Gastropila* Homrich & J.E. Wright, *Bovista*, *Bryoperdon*, *Calvatia*, *Calbovista* Morse ex M.T. Seidl., *Lycoperdon* Pers., and *Morganella* Zeller (He et al. 2019). However, certain puffball-like genera were still accommodated within Agaricaceae, such as *Abstoma*, *Acutocapillitium* P. Ponce de León, *Arachnion* Schwein., *Calvatiopsis* Hollós, *Disciseda* Czern, *Glyptoderma* R. Heim & Perr. – Bertr., *Japonogaster* Kobayasi, and *Lycoperdopsis*

Henn. (He et al. 2019). Conversely, a recent compendium of generic names for agarics and Agaricales, Kalichman et al. (2020), proposed the concept of Agaricaceae s.l., in which this group includes five families: Lycoperdaceae, Agaricaceae s. str., Coprinaceae, Lepiotaceae, and Tulostomataceae. Lycoperdaceae has more recently been acknowledged as an independent family (He et al. 2019; Kalichman et al. 2020), even though historically it was treated as synonymous with Agaricaceae (Kirk et al. 2008). Both Index Fungorum (http://www.indexfungorum.org; 23 March 2025) and the MycoBank database (http://www.MycoBank.org; 23 March 2025) have registered 1,453 specific and infraspecific names in Lycoperdaceae, but the actual number of species has been estimated to be around 430 (He et al. 2024; Hyde et al. 2024).

Historically, the genera of Lycoperdaceae were classified by morphological characteristics such as the presence of capillitium or paracapillitium, the type of capillitium (*Lycoperdon*-type, *Bovista*-type, *Calvatia*-type, intermediate *Bovista-Lycoperdon* type, and *Mycenastrum*-type), the apical opening type of endoperidium, and the presence or absence of the pseudostipe (Kreisel 1969; Pegler et al. 1995; Larsson and Jeppson 2008).

Lycoperdon is the type genus of this family, Lycoperdaceae, and Larsson and Jeppson (2008) recommended a wide-sense concept of *Lycoperdon* based on molecular phylogenetics, in which this family incorporates some traditional genera such as *Bovistella*, *Morganella*, and *Vascellum*. However, some recent studies showed the taxonomic arrangement of five subgenera (*Bovistella*, *Lycoperdon*, *Morganella*, *Utraria* Quél., and *Vascellum*) did not fit well with some taxa (Bates et al. 2009; Gube and Piepenbring 2009; Jeppson and Larsson 2010; Alfredo et al. 2017).

The delineation of genera within the family Lycoperdaceae has undergone significant changes based on the integration of morphological characteristics (Kreisel 1969; Hibbett et al. 1997; Krüger et al. 2001; Moncalvo et al. 2002; Krüger and Kreisel 2003; Bates 2004; Vizzini and Ercole 2017; Krakhmalnyi et al. 2023). With the contributions of numerous prominent mycologists, the taxonomic system of Lycoperdaceae was converging toward a comprehensive understanding of its natural divergence history. Based on ITS and nLSU sequences, the first molecular phylogenetic analysis of Lycoperdaceae was conducted (Larsson and Jeppson 2008), which includes *Lycoperdon*, *Bovista*, *Calvatia*, and *Disciseda* (Krüger et al. 2001; Larsson and Jeppson 2008; Bates et al. 2009; Larsson et al. 2009; Alfredo et al. 2017). A comprehensive phylogeny of the family Lycoperdaceae was reconstructed using sequences from ITS+nLSU+*rpb2*+TEF1-α genetic markers, in which divergence times were estimated for taxa, serving as an additional criterion for taxonomic classification. Combined with morphological analyses, the updated phylogenetic framework supported the division of Lycoperdaceae into 19 genera with divergence times spanning 26.7–75.5 million years ago (Li et al. 2024).

During investigations on puffball fungi in China, we collected many puffball fungi specimens. The objectives of the present study are to explore new fungal taxa in Lycoperdaceae and to reveal the phylogenetic relationships of puffball fungi. Based on the morphological and molecular phylogenetic study, we discovered one new genus, *Lycoperdia*, and three new species, *Calvatia phlebioides*, *Lycoperdia tomentosa*, and *Morganella minima*, on the basis of ITS, nLSU, and *rpb2* sequences.

Materials and methods

Sample collection and examination

The fresh basidiomes of fungi growing on the ground were collected from Qingyuan in Guangdong Province, Dehong, and Zhaotong in Yunnan Province, P.R. China. The samples were photographed using a Xiaomi 12 *in situ*, and fresh macroscopic details, such as the color of the basidiomes, the type of exoperidium, and the shape of the basidiomes, were recorded. All the photographs were focus-stacked and merged using Helicon Focus Pro 7.7.5 software. Specimens were dried in an electric food dehydrator at 40 °C (Hu et al. 2022; Zhao et al. 2023a; Dong et al. 2024a, b) and then sealed and stored in an envelope bag and deposited in the herbarium of the Southwest Forestry University (SWFC), Kunming, Yunnan Province, P.R. China.

Morphology

The macromorphological descriptions were based on field notes and photos captured in the field and laboratory and followed the color terminology of Petersen (1996). Micromorphological data were obtained from the dried specimens following observation under a light microscope (Dong et al. 2024a; Yang et al. 2025). Drawings were made using a fungus plotter (Zhao et al. 2023a). The measurements and drawings were made from slide preparations stained with Cotton Blue (0.1 mg aniline blue dissolved in 60 g pure lactic acid) and 5% potassium hydroxide. Spore size data, excluding 5% of the measurements from each end of the range, are shown in parentheses. The following abbreviations were used: KOH = 5% potassium hydroxide water solution, CB+ = cyanophilous, CB = cotton clue, CB– = acyanophilous, Q = variation in the L/W ratios between the specimens studied and n = a/b (number of spores (a) measured from a given number (b) of specimens), Q_m represented the average Q of basidiospores measured ± standard deviation.

DNA extraction, PCR amplification, sequencing, and phylogenetic analyses

The CTAB rapid plant genome extraction kit—DN14 (Aidlab Biotechnologies Co., Ltd., Beijing, China)—was used to obtain genomic DNA from the dried fungal specimens according to the manufacturer's instructions (Dong et al. 2024a; Yuan et al. 2024; He et al. 2025; Zhang et al. 2025). The extracted DNA was maintained at –20 °C for long-term storage. Three molecular markers were investigated, i.e., internal transcribed spacer (ITS), nuclear large subunit ribosomal RNA (nLSU), and RNA polymerase II subunit 2 (*rpb2*) gene, and the primers and conditions are shown in Table 1. The PCR products were purified and sequenced at Kunming Tsingke Biological Technology Limited Company (Yunnan Province, China). All newly generated sequences were deposited in NCBI GenBank (https://www.ncbi.nlm.nih.gov/genbank/) (Table 2).

Table 1. Loci, primers, PCR amplification procedures, and references used in this study.

Name	Abbreviation	Name	Direction	Sequence (5'-3')	PCR amplification procedures	References
Internal transcribed spacer region of the rDNA	ITS	ITS5	Forward	GGAAGTAAAAGTCGTAACAAGG	94 °C 2 min; 35 cycles of 94 °C 60 s, 55 °C 60 s, 72 °C 2 min; 72 °C 10 min.	White et al. (1990)
		ITS4	Reverse	TCCTCCGCTTATTGATATGC		
Nuclear large subunit ribosomal	nLSU	LR0R	Forward	ACCCGCTGAACTTAAGC	94 °C 2 min; 35 cycles of 94 °C 30 s, 48 °C 1 min, 72 °C 1.5 min; 72 °C 10 min.	Vilgalys and Hester (1990)
		LR7	Reverse	TACTACCACCAAGATCT		
RNA polymerase second largest subunit	*rpb2*	bRPB2-6F	Forward	TGGGGYATGGTNTGYCCYGC	94 °C 2 min; 9 cycles of 94 °C 45 s, 60 °C 45 s, 72 °C 1.5 min; 36 cycles of 94 °C 45 s, 53 °C 1 min, 72 °C 1.5 min; 72 °C 10 min.	Liu et al. (1999)
		bRPB2-7.1R	Reverse	CCCATRGCTTGYTTRCCCAT		

Table 2. A list of species, specimens, and GenBank accession numbers of sequences used in this study. [New species are shown in bold; * indicates type material; - indicates data unavailable].

Species Name	Locality	Sample No.	GenBank Accession No.			References
			ITS	nLSU	*rpb2*	
Abstoma indicum	UZ-04-19	India	MN231720	–	–	Altaf et al. (2022)
Abstoma purpureum	KM162954	England	GQ981488	–	–	Li et al. (2024)
Apioperdon pyriforme	QL20170019	China	PP175742	PP175746	–	Li et al. (2024)
Apioperdon pyriforme	ZRL20182005	China	PP175743	PP175747	–	Li et al. (2024)
Bovista cretacea	ANMH11622	Iceland	DQ112611	–	–	Larsson and Jeppson (2008)
Bovista cretacea	MJ5207	Norway	DQ112610	–	–	Larsson and Jeppson (2008)
Bovista cretacea	ANMH11622	Iceland	DQ112611	–	–	Larsson and Jeppson (2008)
Bovista litangensis	HMAS 258800	China	OR792635	OR831301	–	Li et al. (2024)
Bovista litangensis	HMAS 258801	China	OR792636	OR831302	–	Li et al. (2024)
Bovista nyalamensis	HMAS 258836	China	OR792637	OR831304	–	Li et al. (2024)
Bovista nyalamensis	HMAS 258837	China	OR792638	OR831306	–	Li et al. (2024)
Bovista plumbea	NYGD01	Pakistan	JX183694	–	–	Yousaf et al. (2013)
Bovista plumbea	MJ4856	Sweden	DQ112613	–	–	Larsson and Jeppson (2008)
Bovistella emodensis	HMAS 287485	China	PP175744	PP175752	–	Li et al. (2024)
Bovistella emodensis	HMAS 287486	China	PP175745	PP175753	–	Li et al. (2024)
Bryoperdon acuminatum	TO HG191016	Italy	KY581201	KY581199	–	Vizzini and Ercole (2017)
Bryoperdon acuminatum	TO HG201016	Italy	KY581202	KY581200	–	Vizzini and Ercole (2017)
Calvatia bicolor	LMG756-58	USA	EU833651			Bates et al. (2009)
Calvatia candida	MJ3514	Hungary	DQ112624	–	–	Larsson and Jeppson (2008)
Calvatia craniiformis	Steinke001017	USA	DQ112625	–	–	Larsson and Jeppson (2008)
Calvatia cyathiformis	Strain JTT10	USA	MF686508	–	–	Senthilarasu et al. (2018)
Calvatia cyathiformis	MP12	Canada	KY706183	–	–	Senthilarasu et al. (2018)
Calvatia fenzlii	Strain Jz01	China	FJ772413	–	–	Senthilarasu et al. (2018)
Calvatia fragilis	AH 24114	Argentina	AJ486959	–	–	Senthilarasu et al. (2018)
Calvatia fragilis	AH 25227	Pakistan	AJ486958	–	–	Senthilarasu et al. (2018)
Calvatia gigantea	MJ3566	Sweden	DQ112623	–	–	Larsson and Jeppson (2008)
Calvatia gigantea	HMAS 258889	China	OR792628	OR831299	OR853762	Li et al. (2024)
Calvatia holothurioides	LE 287408	Japan	JQ734547	–	–	Rebriev (2013)
Calvatia holothurioides	KA11-0287	South Korea	KJ909662	–	–	Kim et al. (2016)
Calvatia longisetulosa	HMAS 258802	Thailand	OR792617	OR831229	OR853757	Li et al. (2024)
Calvatia longisetulosa	HMAS 258803	Thailand	OR792618	OR831230	OR853758	Li et al. (2024)
Calvatia nodulata	BAFC 4549	Argentina	KY366490	–	–	Alfredo et al. (2014)

Species Name	Locality	Sample No.	GenBank Accession No.			References
			ITS	nLSU	*rpb2*	
Calvatia nodulata	UFRN Fungos 1691	Brazil	KP751206	–	–	Alfredo et al. (2014)
Calvatia pachydermica	AN014692	USA	EU833653	–	–	Bates et al. (2009)
Calvatia phlebioides	**CL Zhao 33216***	**China**	**PV345681**	**PV345675**	**PV341017**	Present study
Calvatia phlebioides	**CL Zhao 33366**	**China**	**PV345682**	**PV345676**	**PV341018**	Present study
Calvatia rubroflava	TFB11269	Argentina	KY559335	–	–	Senthilarasu et al. (2018)
Calvatia shennongjiaensis	HMAS 258804	China	OR792621	OR831294	OR853761	Li et al. (2024)
Calvatia shennongjiaensis	HMAS 258806	China	OR792623	OR831291	OR853759	Li et al. (2024)
Calvatia shennongjiaensis	HMAS 258808	China	OR792625	OR831292	OR853760	Li et al. (2024)
Calvatia subbooniana	HMAS 258809	China	OR792631	OR831296	–	Li et al. (2024)
Calvatia subbooniana	HMAS 258810	China	OR792630	OR831297	–	Li et al. (2024)
Disciseda bovista	MJ5078	Sweden	DQ112627	–	–	Larsson and Jeppson (2008)
Disciseda candida	STB304	USA	EU833654	–	–	Bates et al. (2009)
Disciseda cervina	FK17016	Argentina	MN338568	–	–	Li et al. (2024)
Fuscospina nigrescens	MJ5376	Sweden	DQ112577	–	–	Larsson and Jeppson (2008)
Fuscospina nigrescens	HMAS 258881	China	OR792702	OR831256	–	Li et al. (2024)
Fuscospina scabricapillitia	HMAS 258812 T	China	OR792678	PP175748	–	Li et al. (2024)
Globaria aestivalis	MJ1122	Sweden	DQ112620	–	–	Larsson and Jeppson (2008)
Globaria gyirongensis	HMAS 258813	China	OR792660	OR831288	–	Li et al. (2024)
Globaria gyirongensis	HMAS 258814	China	OR792661	OR831287	–	Li et al. (2024)
Globaria jingningensis	HMAS 258816	China	OR792645	OR831272	–	Li et al. (2024)
Globaria jingningensis	HMAS 258817	China	OR792642	OR831271	–	Li et al. (2024)
Globaria testacea	HMAS 258819	China	OR792656	OR831280	–	Li et al. (2024)
Holocotylon biconicum	HMAS 258778	China	OR792667	OR831266		Li et al. (2024)
Holocotylon dermoxanthum	MJ4856	Sweden	DQ112579	–	–	Larsson and Jeppson (2008)
Holocotylon rupicola	MJ4304	Norway	DQ112580	–	–	Jeppson et al. (2012)
Holocotylon rupicola	MJ7007	Sweden	JN572902	–	–	Jeppson et al. (2012)
Leptocaulis ericaeus	MJ5395	Sweden	DQ112605	–	–	Larsson and Jeppson (2008)
Leptocaulis ericaeus	KA13-1463	South Korea	KP340185	–	–	Kim et al. (2016)
Leptocaulus albiperidia	KA12-1210	South Korea	KP340182	–	–	Kim et al. (2016)
Leptocaulus albiperidia	KA121551 t	South Korea	KP340183	–	–	Kim et al. (2016)
Leptocaulus muscorum	MJ9017	Sweden	JN572905	–	–	Jeppson et al. (2012)
Leptocaulus sublongistipes	HMAS 258775	China	OR792741	OR831318	–	Li et al. (2024)
Leptocaulus subumbrinus	HMAS 258885	China	OR792673	OR831315	–	Li et al. (2024)
Leptocaulus subumbrinus	HMAS 258886	China	OR792675	OR831317	–	Li et al. (2024)
Lycoperdia tomentosa	**CL Zhao 37502***	**China**	**PV345683**	**PV345677**	–	**Present study**
Lycoperdia tomentosa	**CL Zhao 45073**	**China**	**PV345684**	**PV345678**	–	**Present study**
Lycoperdiscus lividus	MJ4005	Sweden	DQ112600	–	–	Larsson and Jeppson (2008)
Lycoperdiscus lividus	Dobremez 19740514	Nepal	DQ112599	–	–	Larsson and Jeppson (2008)
Lycoperdiscus tianzhuensis	HMAS 258767	China	OR792725	OR831324	–	Li et al. (2024)
Lycoperdiscus tianzhuensis	HMAS 258766	China	OR792725	OR831324	–	Li et al. (2024)
Lycoperdon norvegicum	MJ5453	Sweden	DQ112631	–	–	Larsson and Jeppson (2008)
Lycoperdon perlatum	HMAS 258865	China	OR792758	OR831262	–	Li et al. (2024)
Lycoperdon perlatum	HMAS 258866	China	OR792763	OR831257	–	Li et al. (2024)
Lycoperdon subperlatum	HMAS 258873	China	OR792751	OR831261	–	Li et al. (2024)
Lycoperdon subperlatum	HMAS 258875	China	OR792750	OR831260	–	Li et al. (2024)
Morganella albostipitata	INPA239563	Brazil	KU958363	KU958364	–	Alfredo et al. (2017)
Morganella albostipitata	UFRN-Fungos2249	Brazil	KU958361	KU958362	–	Alfredo et al. (2017)
Morganella albostipitata	UFRN-Fungos2569	Brazil	KU958357	KU958358	–	Alfredo et al. (2017)
Morganella albostipitata	UFRN-Fungos2572	Brazil	KU958359	KU958360	–	Alfredo et al. (2017)
Morganella fuliginea	UFRN-Fungos2582	Brazil	KU958339	KU958340	–	Alfredo et al. (2017)

Species Name	Locality	Sample No.	GenBank Accession No.			References
			ITS	nLSU	*rpb2*	
Morganella fuliginea	UFRN-Fungos2586	Brazil	KU958343	KU958344	–	Alfredo et al. (2017)
Morganella fuliginea	UFRN-Fungos2579	Brazil	KU958349	KU958350	–	Alfredo et al. (2017)
Morganella minima	**CL Zhao 40537***	**China**	**PV345685**	**PV345679**	–	**Present study**
Morganella minima	**CL Zhao 45072**	**China**	**PV345686**	**PV345680**	–	**Present study**
Morganella nuda	UFRN-Fungos2568	Brazil	KU958313	KU958314	–	Alfredo et al. (2017)
Morganella nuda	UFRN-Fungos1766	Brazil	KU958315	KU958316	–	Alfredo et al. (2017)
Morganella nuda	UFRN-Fungos 2565	Brazil	KU958311	KU958312	–	Alfredo et al. (2017)
Morganella nuda	ICN 154541	Brazil	KU958317	KU958318	–	Alfredo et al. (2017)
Morganella nuda	UFRN-Fungos 1765	Brazil	KU958319	KU958320	–	Alfredo et al. (2017)
Morganella oblongata	UFRN-Fungos2570	Brazil	KU958355	KU958356	–	Alfredo et al. (2017)
Morganella purpurascens	MEL 2382736	Australia	KP012918	–	–	Alfredo et al. (2017)
Morganella sosinii	YR2013	Russia	KC591769	–	–	Alfredo et al. (2017)
Morganella subincarnata	REG106	Germany	AJ237626	–	–	Alfredo et al. (2017)
Morganella subincarnata	TNS Kasuya B286	Japan	KF551244	–	–	Alfredo et al. (2017)
Morganella tricolor	HMAS 287487	China	PP175741	PP175750	–	Li et al. (2024)
Mycenastrum corium	MJ5467	Sweden	DQ112628	–	–	Larsson and Jeppson (2008)
Pseudoperdon medogense	HMAS 258784	China	OR792745	OR831254	–	Li et al. (2024)
Pseudoperdon medogense	HMAS 258785	China	OR792746	OR831255	–	Li et al. (2024)
Pseudoperdon subcretaceum	MJ9032	Sweden	JN572908	–	–	Jeppson et al. (2012)
Sinoperdon caudatum	RGC920818	Sweden	DQ112633	–	–	Jeppson and Larsson (2010)
Sinoperdon gyirongense	HMAS 258787	China	OR792686	OR831248	–	Li et al. (2024)
Sinoperdon gyirongense	HMAS 258788	China	OR792699	OR831243	–	Li et al. (2024)
Utraria excipuliformis	HMAS 258850	China	OR792705	OR831326	–	Li et al. (2024)
Utraria excipuliformis	HMAS 258851	China	OR792703	OR831328	–	Li et al. (2024)
Vascellum curtisii	HMAS 258878	China	OR792665	OR831258	–	Li et al. (2024)
Vascellum curtisii	HMAS 258879	China	OR792666	OR831259	–	Li et al. (2024)
Vascellum intermedium	STB091	USA	EU833667	–	–	Bates et al. 2009
Vascellum pratense	MJ5858	Czechia	DQ112556	–	–	Larsson and Jeppson (2008)
Vascellum pratense	HMAS 258880	China	OR792664	PP175749	–	Li et al. (2024)

Sequences generated for this study were aligned with additional sequences downloaded from GenBank. Sequences were aligned in MAFFT 7 (https://mafft.cbrc.jp/alignment/server/), adjusting the direction of nucleotide sequences according to the first sequence (accurate enough for most cases) and selecting the G-INS-i iterative refinement method (Katoh et al. 2019). The alignment was adjusted manually using AliView version 1.27 (Larsson 2014). The dataset was aligned first, and then the sequences of ITS+nLSU+*rpb2* were combined with Mesquite v. 3.51. The combined ITS+nLSU+*rpb2* sequences and ITS+nLSU datasets were used to infer the positions of the new species and related species. The sequence of *Mycenastrum corium* (Guers.) Desv. was retrieved from GenBank and used as outgroup taxa in the ITS+nLSU analysis (Fig. 1) in the family Lycoperdaceae (Li et al. 2024); *Disciseda candida* (Schwein.) Lloyd was selected as the outgroup taxon for the ITS+nLSU+ *rpb2* analysis (Fig. 2) in the genus *Calvatia* (Li et al. 2024); *Bovista cretacea* T.C.E. Fr. was selected as the outgroup taxon for the ITS+nLSU analysis (Fig. 3) in the genus *Morganella* (Li et al. 2024).

Maximum likelihood (ML) and Bayesian inference (BI) analyses were applied to the combined two datasets following a previous study (Zhao and Wu 2017; Xu et al. 2025). Maximum likelihood (ML) analysis was performed using the CIPRES Science Gateway (Miller et al. 2012) based on the dataset using the RAxML-HPC BlackBox tool, with RAxML-HPC BlackBox halted after bootstrapping automatically

0.25 with maximum hours and obtaining the best tree using ML search. Other parameters in ML analysis used default settings, and statistical support values were obtained using nonparametric bootstrapping with 1,000 replicates.

The best evolutionary model of each alignment was estimated using jModelTest (Guindon and Gascuel 2003; Posada 2008) under the Akaike information criterion. MrModeltest 2.3 (Nylander 2004) was used to determine the best-fit evolution model for the dataset for Bayesian inference (BI). Bayesian inference was performed with MrBayes 3.1.2 with a general time reversible (GTR+I+G) model of DNA substitution and a gamma distribution rate variation across sites (Ronquist et al. 2012). Branches were considered significantly supported if they received a maximum likelihood bootstrap value (BS) of ≥ 70% or Bayesian posterior probabilities (BPP) of ≥ 0.95.

Results

Phylogenetic analyses

The combined ITS+nLSU dataset (Fig. 1) included sequences from 86 fungal specimens representing 54 species. The best model for the ITS+nLSU dataset estimated and applied in the Bayesian analysis was GTR+I+G (lset nst = 6; rates = invgamma; prset statefreqpr = dirichlet (1, 1, 1, 1). The Bayesian and ML analyses resulted in a similar topology to that of the MP analysis, with an average standard deviation of split frequencies = 0.009598 (BI), and the effective sample size (ESS) across the two runs is double the average ESS (avg. ESS) = 558. Branches that received bootstrap support for ML and BI ≥ 70% and 0.95 were considered significantly supported, respectively. The results of BLAST queries in NCBI, based on ITS+nLSU separately, showed the sequences producing significant alignment descriptions.

The combined ITS+nLSU+*rpb2* dataset (Fig. 2) included sequences from 26 fungal specimens representing 16 species. The best model for the ITS+nLSU+*rpb2* dataset estimated and applied in the Bayesian analysis was GTR+I+G (lset nst = 6; rates = invgamma; prset statefreqpr = dirichlet (1, 1, 1, 1). The Bayesian and ML analyses resulted in a similar topology to that of the MP analysis, with an average standard deviation of split frequencies = 0.006367 (BI), and the effective sample size (ESS) across the two runs is double the average ESS (avg. ESS) = 1,441. Branches that received bootstrap support for ML and BI ≥ 70% and 0.95 were considered significantly supported, respectively. The results of BLAST queries in NCBI, based on ITS+nLSU+*rpb2* separately, showed the sequences producing significant alignment descriptions.

The combined ITS+nLSU dataset (Fig. 3) included sequences from 18 fungal specimens representing 10 species. The best model for the ITS+nLSU dataset estimated and applied in the Bayesian analysis was GTR+I+G (lset nst = 6; rates = invgamma; prset statefreqpr = dirichlet (1, 1, 1, 1). The Bayesian and ML analyses resulted in a similar topology to that of the MP analysis with an average standard deviation of split frequencies = 0.008915 (BI), and the effective sample size (ESS) across the two runs is double the average ESS (avg. ESS) = 1243.5. Branches that received bootstrap support for ML and BI ≥ 70% and 0.95 were considered significantly supported, respectively. The results of BLAST queries in NCBI, based on ITS+nLSU separately, showed the sequences producing significant alignment descriptions.

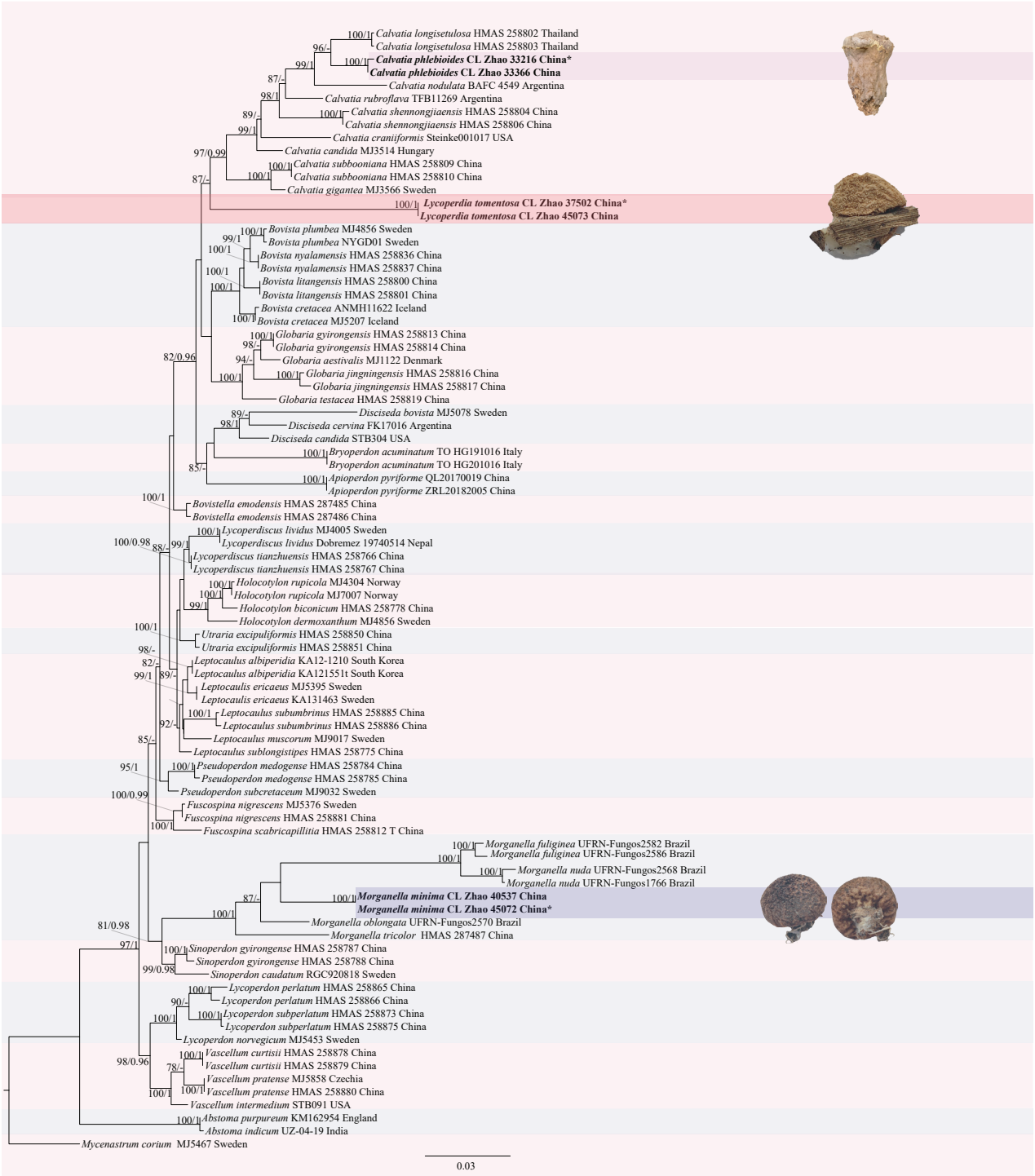

Figure 1. Maximum likelihood strict consensus tree illustrating the phylogeny of one new genus and three new species and related species in Lycoperdaceae based on ITS+nLSU sequences. Branches are labeled with maximum likelihood bootstrap values ≥ 70% and Bayesian posterior probabilities ≥ 0.95, respectively. New species are in bold, * type material.

In the ITS BLAST results of *Calvatia phlebioides*, the top ten taxa were *C. holothurioides* Rebriev and *C. candida* (Rostk.) Hollós (maximum record descriptions: max score 1096; total score 1096; query cover 98%; E value 0; ident 94.63%). In nLSU BLAST results, the top ten taxa were *Bovista nigrescens* Pers., *Lycoperdon perlatum*, *Calvatia candida*, *C. craniiformis*, and *C. caatingaensis* R.L. Oliveira, R.J. Ferreira, B.D.B. Silva, M.P. Martín & Baseia (Maximum record

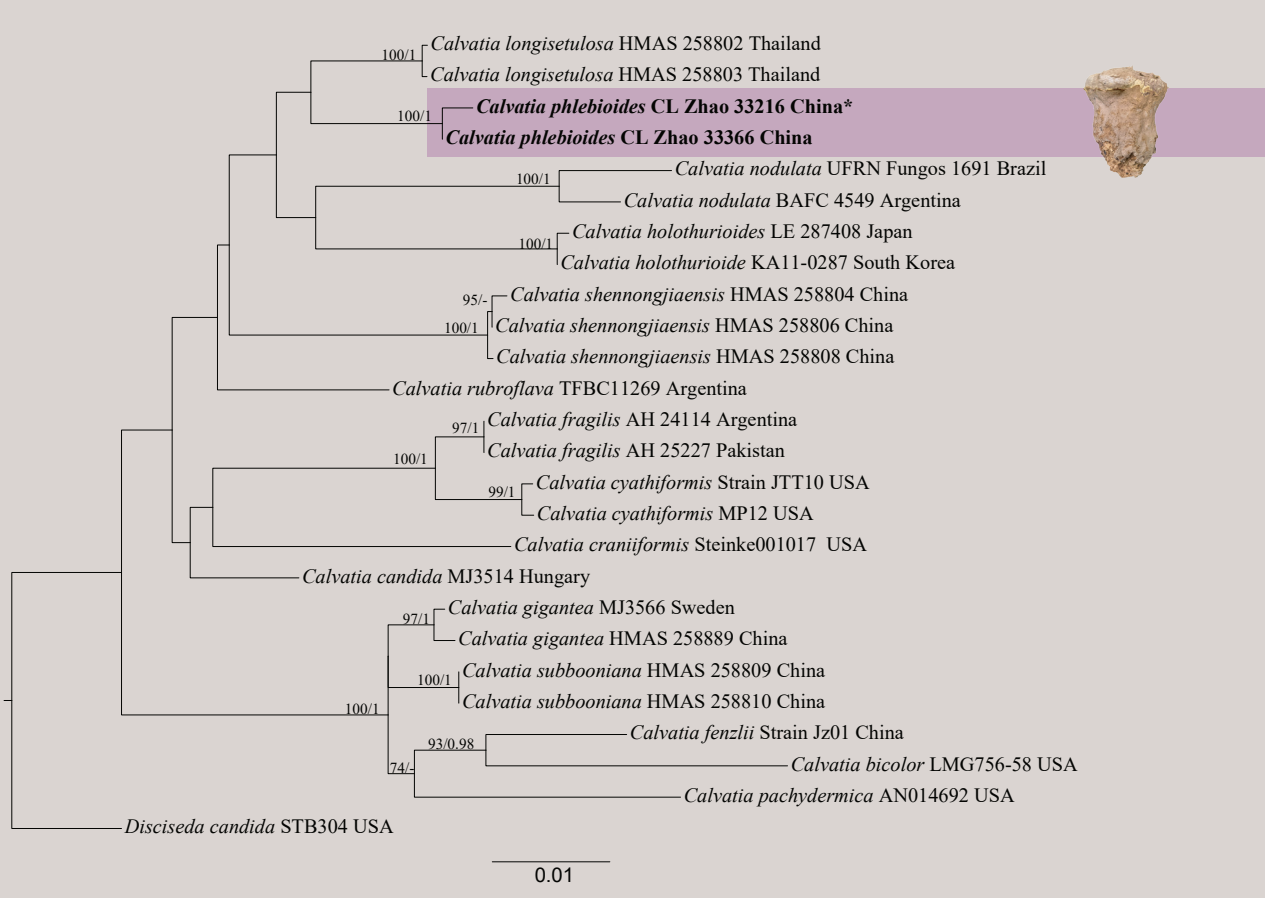

Figure 2. Maximum likelihood strict consensus tree illustrating the phylogeny of *Calvatia phlebioides* and related species in *Calvatia* based on ITS+nLSU+*rpb2* sequences. Branches are labeled with maximum likelihood bootstrap values ≥ 70% and Bayesian posterior probabilities ≥ 0.95, respectively. New species accessions are in bold, * type material.

descriptions: Max score 1648; Total score 1648; Query cover 100%; E value 0.0; Ident 97.81%). In *rpb2* BLAST results, the top ten taxa were *Calvatia longisetulosa* R.L. Zhao & J.X. Li, *C. shennongjiaensis* R.L. Zhao & J.X. Li, *C. rubroflava* (Cragin) Lloyd, *Globaria muscicola* R.L. Zhao & J.X. Li, *Bovistella emodensis* R.L. Zhao & J.X. Li, and *Bovista plumbea* Pers. (Maximum record descriptions: Max score 966; Total score 966; Query cover 88%; E value 0.0; Ident 91.91%).

In the ITS BLAST results of *Lycoperdia tomentosa*, the top ten taxa were *Morganella puiggarii* (Speg.) Kreisel & Dring (maximum record descriptions: max score 771; total score 771; query cover 93%; E value 0; ident 89.82%). In nLSU BLAST results, the top ten taxa were *Bovista nigrescens* Pers. and *Lycoperdon perlatum* (maximum record descriptions: max score 1517; total score 1517; query cover 98%; E value 0.0; ident 94.76%).

In the ITS BLAST results of *Morganella minima*, the top ten taxa were *Morganella subincarnata* (Peck) Kreisel & Dring, *Lycoperdon purpurascens* Berk. & M.A. Curtis, and *M. fuliginea* (Berk. & M.A. Curtis) Kreisel & Dring (maximum record descriptions: max score 1026; total score 1026; query cover 93%; E value 0; ident 94.01%). In nLSU BLAST results, the top ten taxa were *Lycoperdon perlatum* and *Bovista nigrescens* (maximum record descriptions: max score 1690; total score 1690; query cover 99%; E value 0.0; ident 98.64%).

The topology based on ITS+nLSU sequences (Fig. 1) showed that all four new taxa were clustered into the family Lycoperdaceae, in which *Lycoperdia tomentosa*

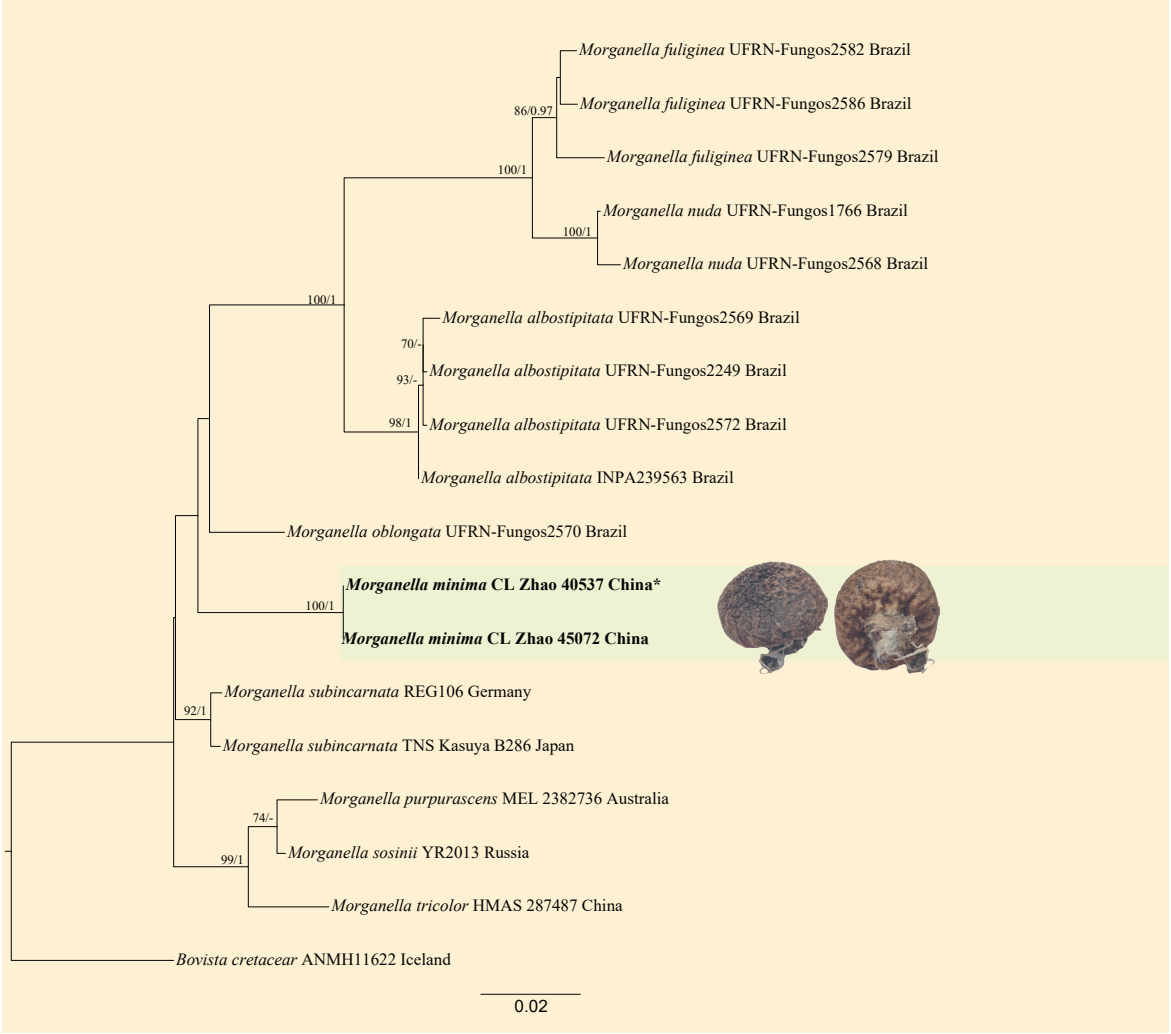

Figure 3. Maximum likelihood strict consensus tree illustrating the phylogeny of *Morganella minima* and related species in *Morganella* based on ITS+nLSU sequences. Branches are labeled with maximum likelihood bootstrap values ≥ 70% and Bayesian posterior probabilities ≥ 0.95, respectively. New species accessions are in bold, * type material.

formed a monophyletic lineage. The topology based on ITS+nLSU+*rpb2* sequences (Fig. 2) showed that one new taxon, *Calvatia phlebioides*, was clustered into the genus *Calvatia*, which was sister to *C. longisetulosa*. The topology based on ITS+nLSU sequences (Fig. 3) showed that *Morganella minima* was clustered into the genus *Morganella*, forming a monophyletic lineage.

Taxonomy

***Calvatia phlebioides* X. Yang & C.L. Zhao, sp. nov.**
MycoBank No: 858401
Figs 4, 5

Holotype. China • Yunnan Province, Zhaotong, Yiliang County, Wumengshan National Nature Reserve, 27°47'19.69"N, 104°15'4.78"E, elev. 2300 m, on the ground, 19 September 2023, CLZhao 33216 (SWFC).

Etymology. *phlebioides* (Lat.) refers to basidiomes that have a phlebioid surface of the stem base.

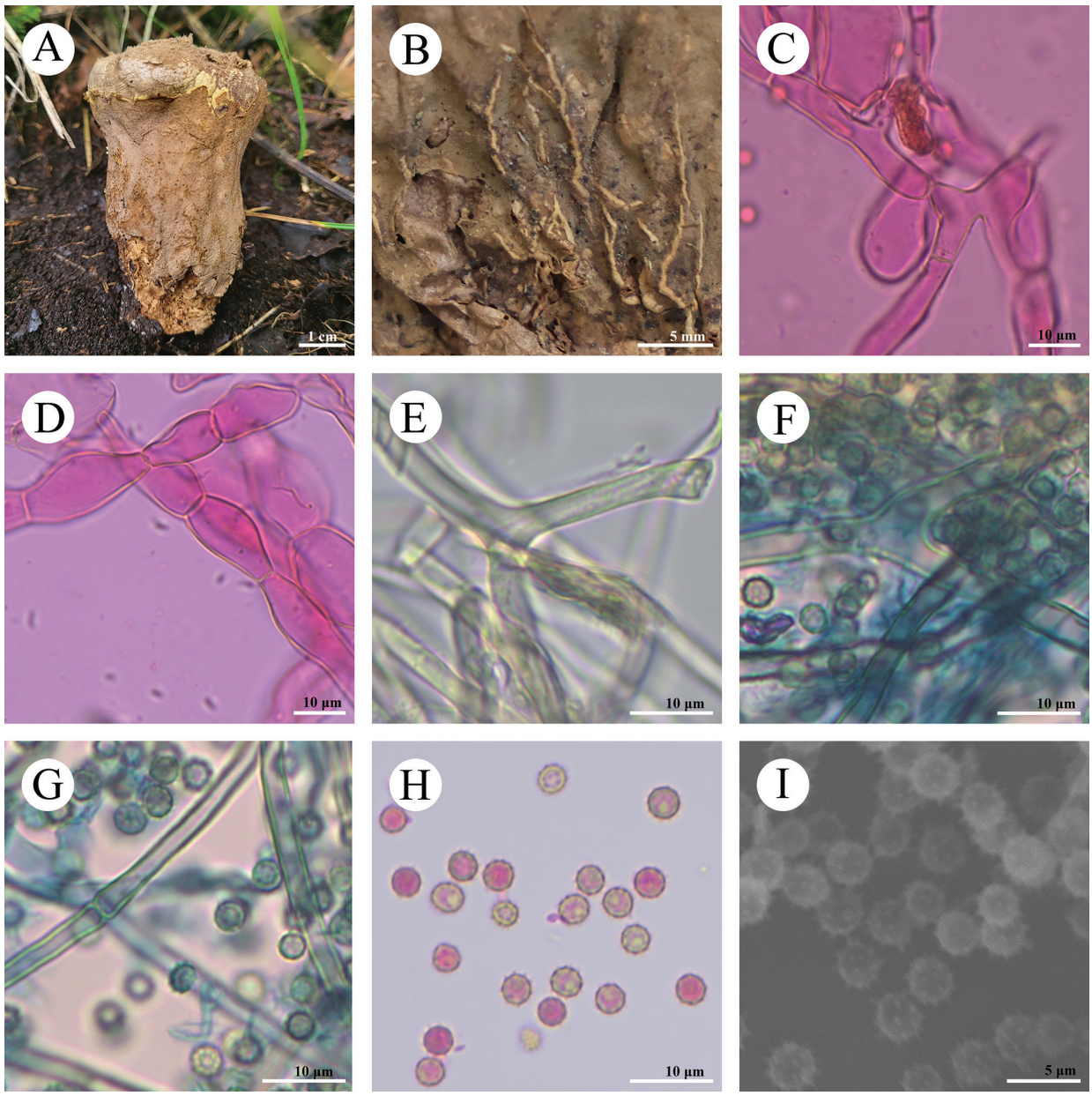

Figure 4. *Calvatia phlebioides* (CLZhao 33216) **A, B** basidiomes of *C. phlebioides* **C, D** exoperidial elements **E** endoperidial hyphae **F, G** capillitial **H** basidiospores **I** basidiospores by SEM.

Description. Fruiting body: ***Basidiomes*** broadly obpyriform to turbinate, 30–50 mm in diameter, 45–65 mm in height, slightly tapering at the base, and developing a round base. ***Peridium*** layered: ***Exoperidium*** brownish to dark brownish when mature and grayish-brown when dry, breaking up into thin, irregular-shaped, flaky patches, and the upper part completely detached, the gleba fully exposed. ***Endoperidium*** fragile, bluish-gray when fresh, ash-gray to bluish-gray when dry. ***Gleba*** pale brown to olive-yellow when mature, cottony.

Hyphal structure: ***Exoperidium*** composed of chains of colorless hyphae, ellipsoid, oblong, pyriform, occasionally globose to subglobose, sphaecysts, thin-walled, smooth, colorless in 5% KOH, (16–)20–33(–35) × 10–15 µm. ***Endoperidium*** composed of undulate, colorless hyphae, thick-walled, occasionally branched, 2.5–3.5 µm in diameter. ***Capillitium*** of *Calvatia*-type, rare branched, septate, thick-walled, 2.5–3.6 µm in diameter. *Paracapillitium* absent.

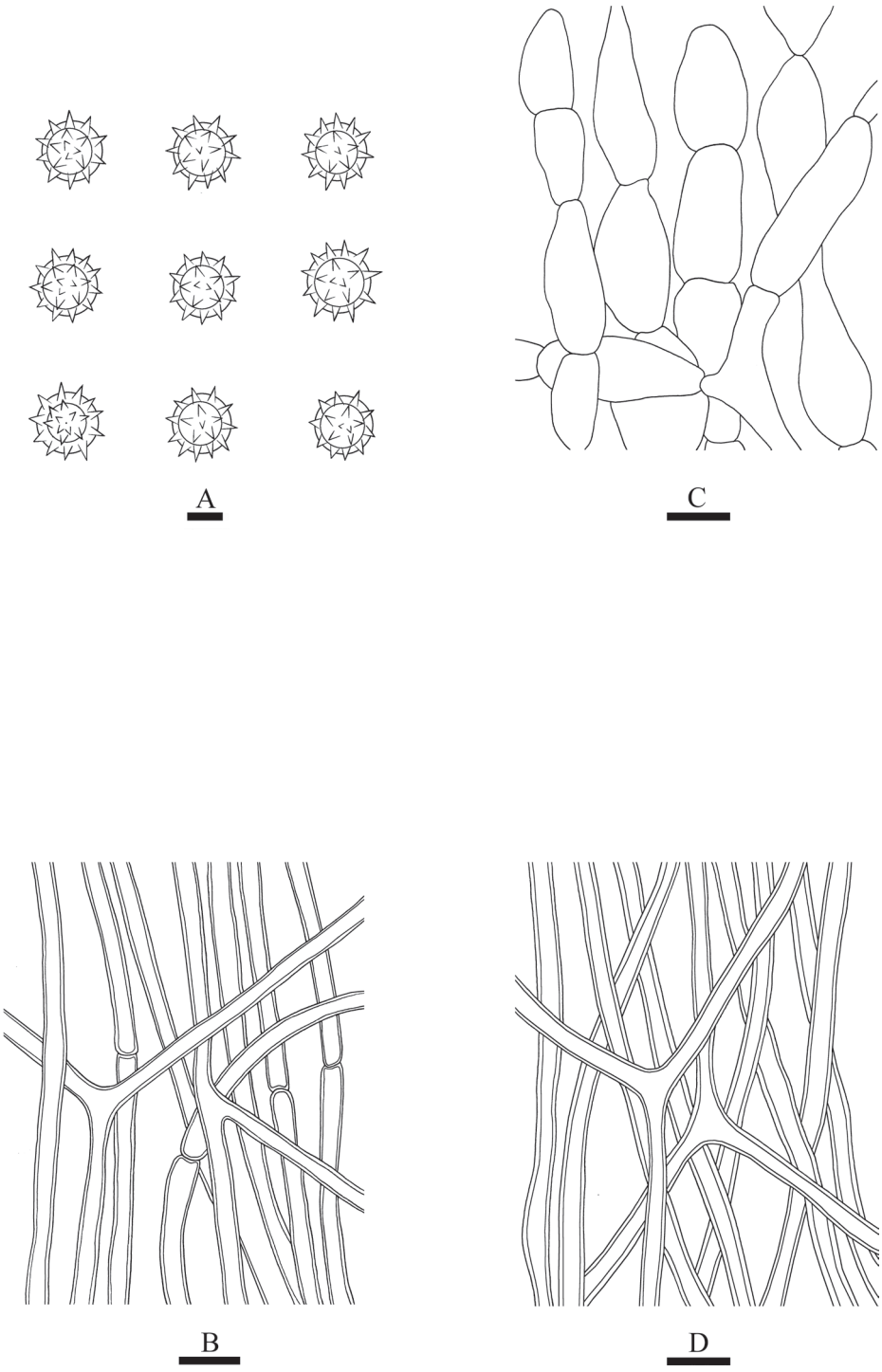

Figure 5. *Calvatia phlebioides* **(**CLZhao 33216) **A** basidiospores **B** capillitial **C** exoperidial elements **D** endoperidial hyphae. Scale bars: 1 μm (**A**); 10 μm (**B–D**).

Basidiospores: Globose to subglobose, 2.8–3.5(–3.8) × 2.7–3.3(–3.7) μm, honey-yellow, thick-walled, CB–, spinose (less than 1 μm), with a short pedicel (less than 0.5 μm), Q = 1.01–1.02, Q_m = 1.02 ± 0.02, n = 60/2.

Additional specimen examined (paratype). China • Yunnan Province, Zhaotong, Yiliang County, Wumengshan National Nature Reserve, 27°47'19.69"N, 104°15'4.78"E, elev. 2300 m, on the ground, 20 September 2023, CLZhao 33366 (SWFC).

***Lycoperdia* X. Yang & C.L. Zhao, gen. nov.**
MycoBank No: 858402

Type species. *Lycoperdia tomentosa* X. Yang & C.L. Zhao

Etymology. *Lycoperdia* (Lat.) refers to the new genus resembling *Lycoperdon* in basidiome morphology.

Description. *Basidiomes* pyriform and subglobose to globose when dry. *Exoperidium* with densely arranged tomentose structures. *Endoperidium* papery and fragile. *Gleba* with cottony texture when dry. *Exoperidium* made up of chains of inflated cells, ellipsoid, oblong, pyriform, thin-walled to slightly thick-walled, smooth, colorless hyphae in 5% KOH. *Endoperidium* made up of colorless hyphae, thick-walled, occasionally branched, no septa. *Capillitium* of *Lycoperdon*-type, branched, thick-walled, no septa. *Paracapillitium* composed of chains of colorless, inflated cells, branched, no septa. *Basidiospores* globose to subglobose, with distinct spines, with a short pedicel.

Notes. In our phylogenetic analyses (Fig. 1), *Lycoperdia* is identified as a monophyletic group, typified by *L. tomentosa*. The new genus *Lycoperdia* falls within the family Lycoperdaceae (Agaricales) and is closely related to *Calvatia*. However, *Calvatia* is distinguished from *Lycoperdia* by its dehiscence of peridium, which occurs by irregular fragmentation, and *Calvatia*-type capillitium (Larsson and Jeppson 2008; Li et al. 2024).

***Lycoperdia tomentosa* X. Yang & C.L. Zhao, sp. nov.**
MycoBank No: 858403
Figs 6, 7

Holotype. China • Yunnan Province, Dehong, Yingjiang County, Tongbiguan Provincial Nature Reserve, 24°28'18.86"N, 97°40'5.60"E, elev. 1000 m, on rotten wood, 3 July 2024, CLZhao 37502 (SWFC).

Etymology. *tomentosa* (Lat.) refers to the tomentose exoperidium surface.

Description. Fruiting body: *Basidiomes* pyriform and subglobose to globose when dry, 5–8 mm in diameter, 6–10 mm in height. ***Peridium*** layered: ***Exoperidium*** sulphur-yellow when fresh, buff-yellow to orange-yellow when dry, with densely arranged tomentose structures, not easily falling off. ***Endoperidium*** fragile, white when fresh, off-white when dry. ***Gleba*** white, powdery, or fibrous.

Hyphal structure: *Exoperidium* composed of chains of inflated cells, ellipsoid to oblong, thin-walled to slightly thick-walled, colorless in 5% KOH, 25–40 × 15–25 µm. ***Endoperidium*** made up of colorless hyphae, thick-walled, occasionally branched, no septa, 2.5–4 µm in diameter. ***Capillitium*** of ***Lycoperdon***-type, branched, thick-walled, no septa, 3.5–5 µm in diameter. ***Paracapillitium*** composed of chains of colorless, inflated cells, 2.5–5.5 µm width.

Basidiospores: Globose to subglobose, (3.3–)3.5–4.8(–5.3) × (3–)3.2–4.5(–4.8) µm, CB+, ash-gray, thick-walled, with distinct spines, with a long pedicel (1.0–6.5 µm), Q = 1.06–1.07, Q_m = 1.06 ± 0.09, n = 60/2.

Additional specimen examined (paratype). China • Yunnan Province, Dehong, Yingjiang County, Tongbiguan Provincial Nature Reserve, 24°28'18.86"N, 97°40'5.60"E, elev. 1000 m, on rotten wood, 18 January 2025, CLZhao 45073 (SWFC).

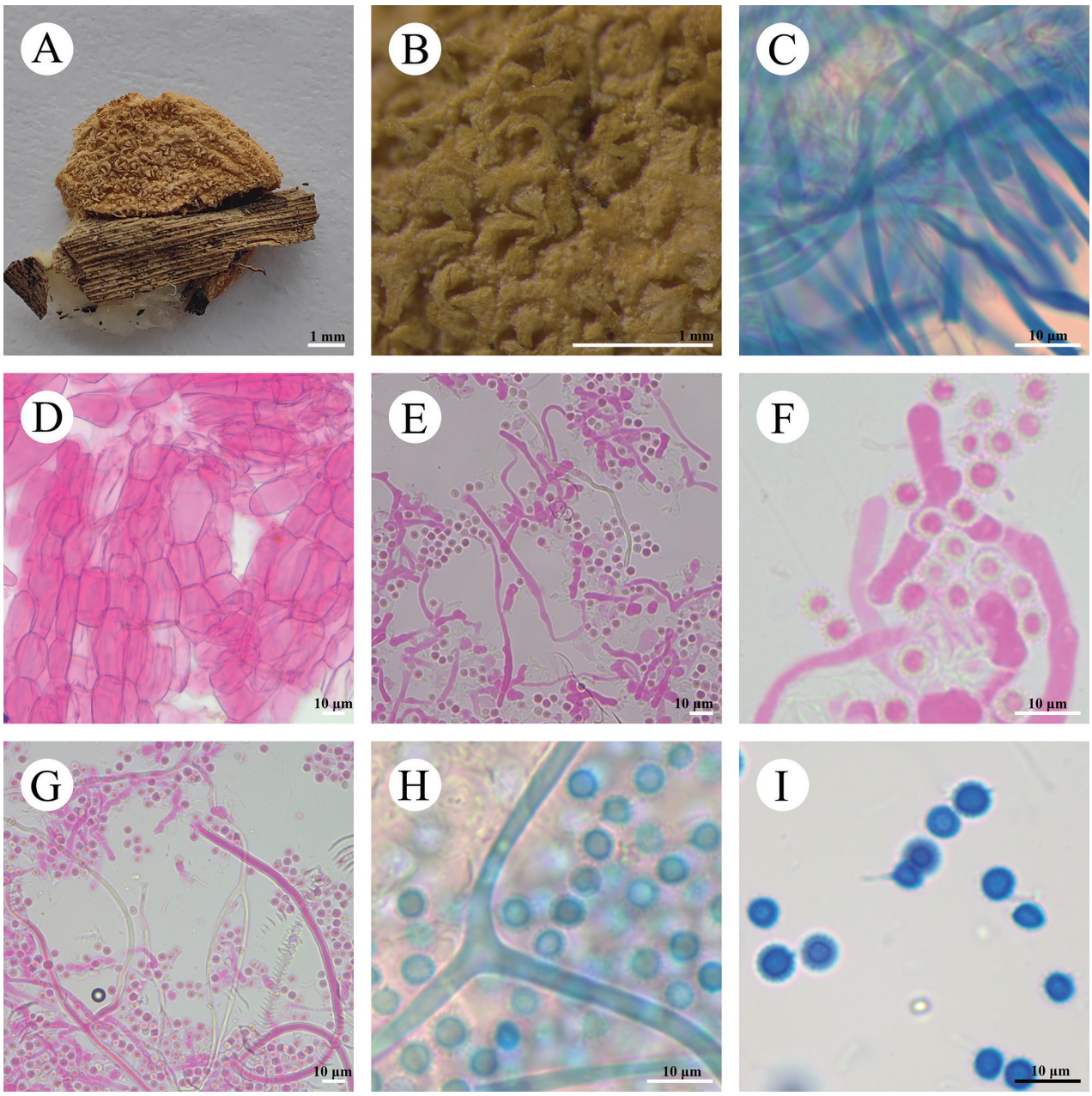

Figure 6. *Lycoperdia tomentosa* (CLZhao 37502) **A, B** basidiomes of *L. tomentosa* **C** endoperidial hyphae **D** exoperidial elements **E, F** paracapillitium **G, H** capillitial **I** basidiospores.

Morganella minima **X. Yang & C.L. Zhao, sp. nov.**
MycoBank No: 858404
Figs 8, 9

Holotype. China • Guangdong Province, Qingyuan, Yingde City, Shimentai National Nature Reserve, 24°25'45.17"N, 113°14'58.4"E, elev. 1100 m, on the ground, 15 September 2024, CLZhao 40537 (SWFC).

Etymology. *minima* (Lat.) refers to the type species having tiny basidiomes.

Description. Fruiting body: ***Basidiomes*** subglobose, 5–8 mm in diameter, 3–7 mm in height. ***Peridium*** layered: ***Exoperidium*** slightly brown when fresh, grayish-brown to fuscous when dry, hymenial surface with coarse granular, not easily falling off, densely grouped on the top of the basidioma;

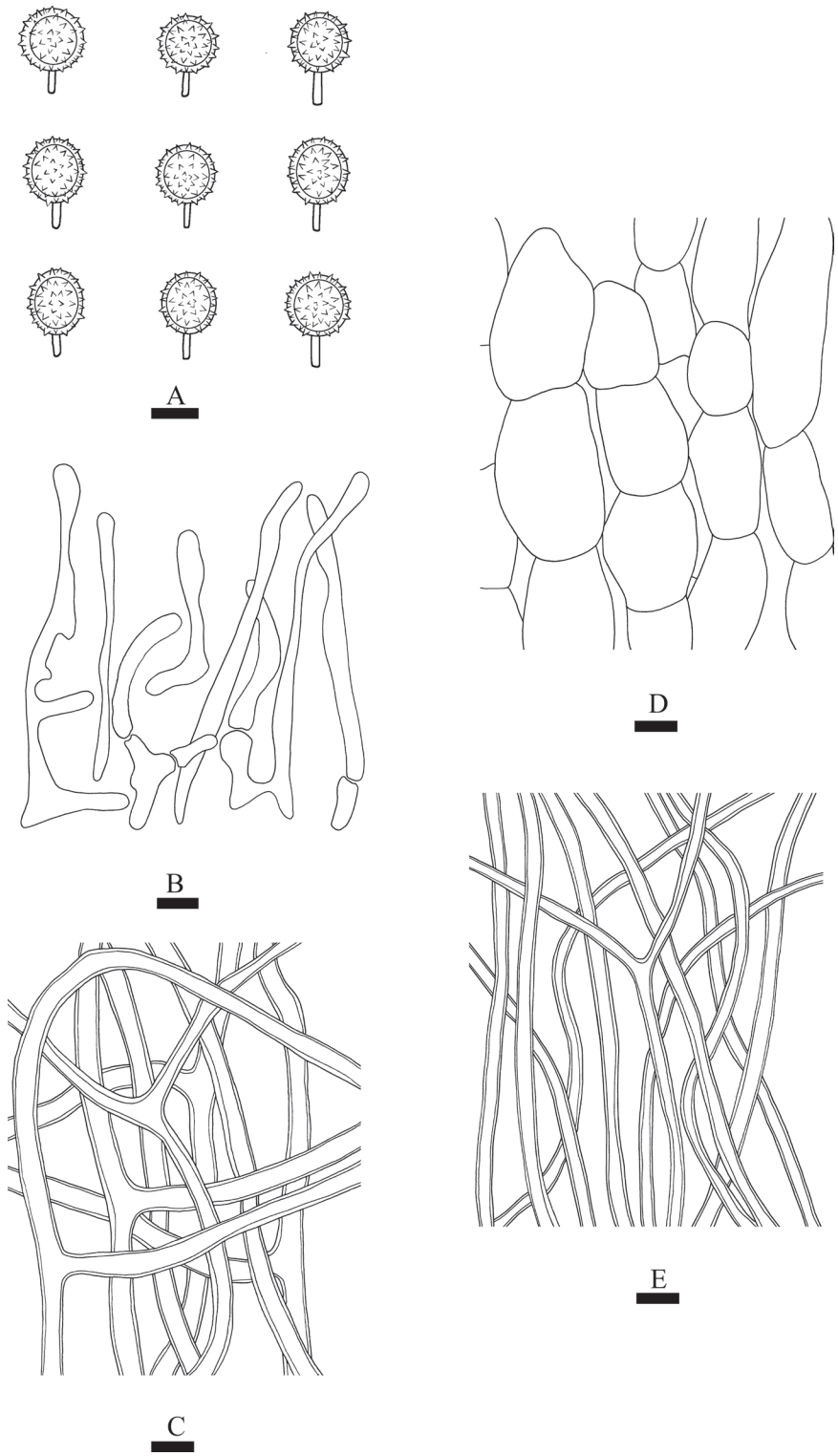

Figure 7. *Lycoperdia tomentosa* **(**CLZhao 37502**) A** basidiospores **B** paracapillitium **C** capillitial **D** exoperidial elements **E** endoperidial hyphae. Scale bars: 1 μm (**A**); 10 μm (**B–E**).

Endoperidium fragile, cream to slightly brown when fresh, slightly brown upon drying. ***Gleba*** fuscous, powdery.

Hyphal structure: *Exoperidium* composed of irregular inflated cells, globose, subglobose, or pyriform, thin-walled, colorless in 5% KOH, 15–50 (–90) × (10)

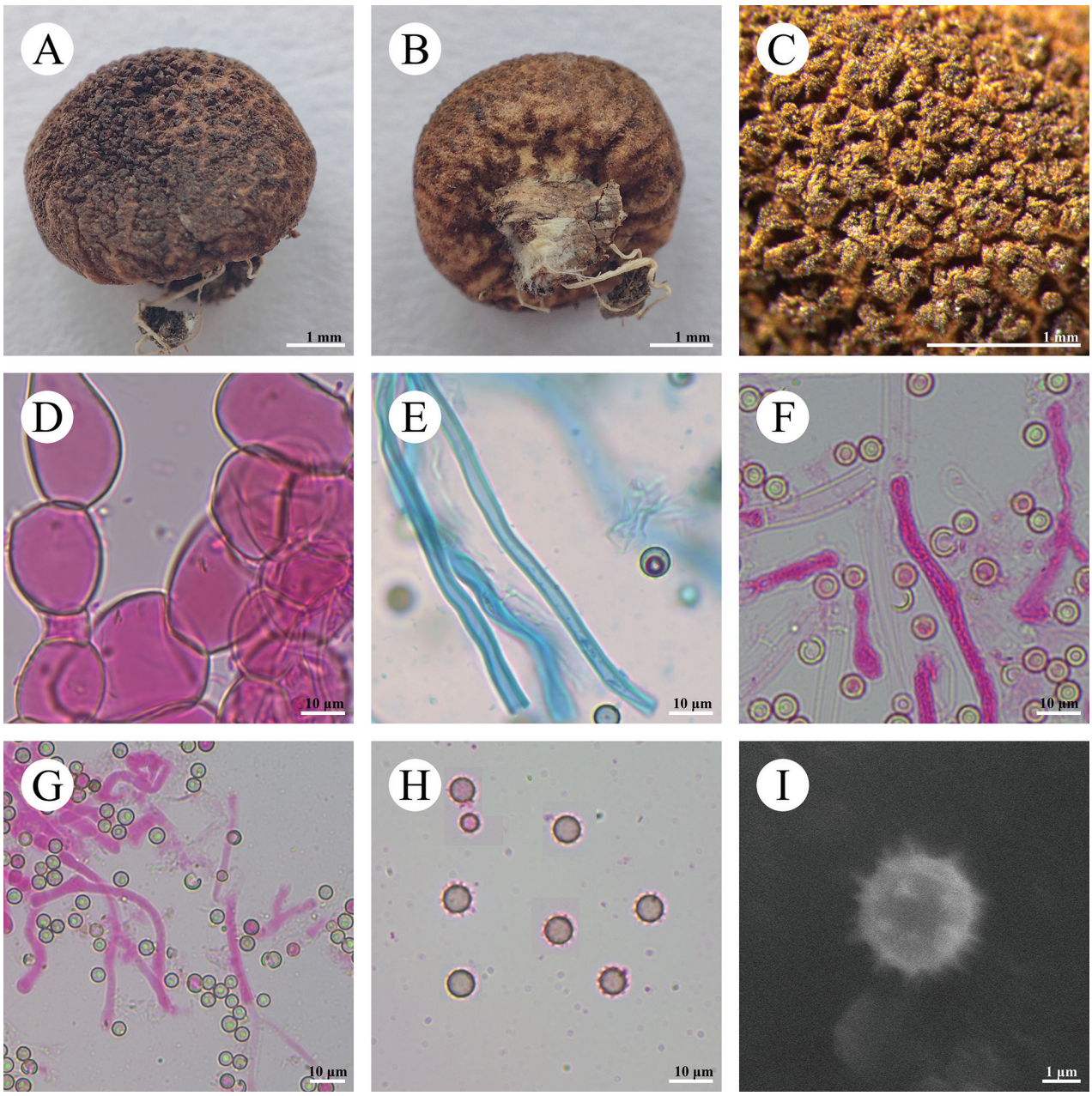

Figure 8. *Morganella minima* (CLZhao 40537) **A–C** basidiomes of *C. phlebioides* **D** exoperidial elements **E** endoperidial hyphae **F, G** paracapillitium **H** basidiospores **I** basidiospores by SEM.

15–32 (–40) µm. ***Endoperidium*** made up of intertwined colorless hyphae, thick-walled, slightly branched, 3–5(–7.5) µm in diameter. ***Capillitium*** absent. ***Paracapillitium*** interwoven and branched, thick-walled, 4–7 µm in diameter, septate.

Basidiospores: Globose to subglobose, 4.7–5.5(–6.3) × 4.6–5.5(–6.2) µm in diameter, brownish, CB–, thin-walled, ornamented with distinct spines (less than 1 µm), with a short pedicel (less than 0.5 µm), Q = 1.02–1.03, Q_m = 1.02 ± 0.02, n = 60/2.

Additional specimen examined (paratype). China • Guangdong Province, Qingyuan, Yingde City, Shimentai National Nature Reserve, 24°25'45.17"N, 113°14'58.4"E, elev. 1100 m, on the ground, 17 January 2025, CLZhao 45072 (SWFC).

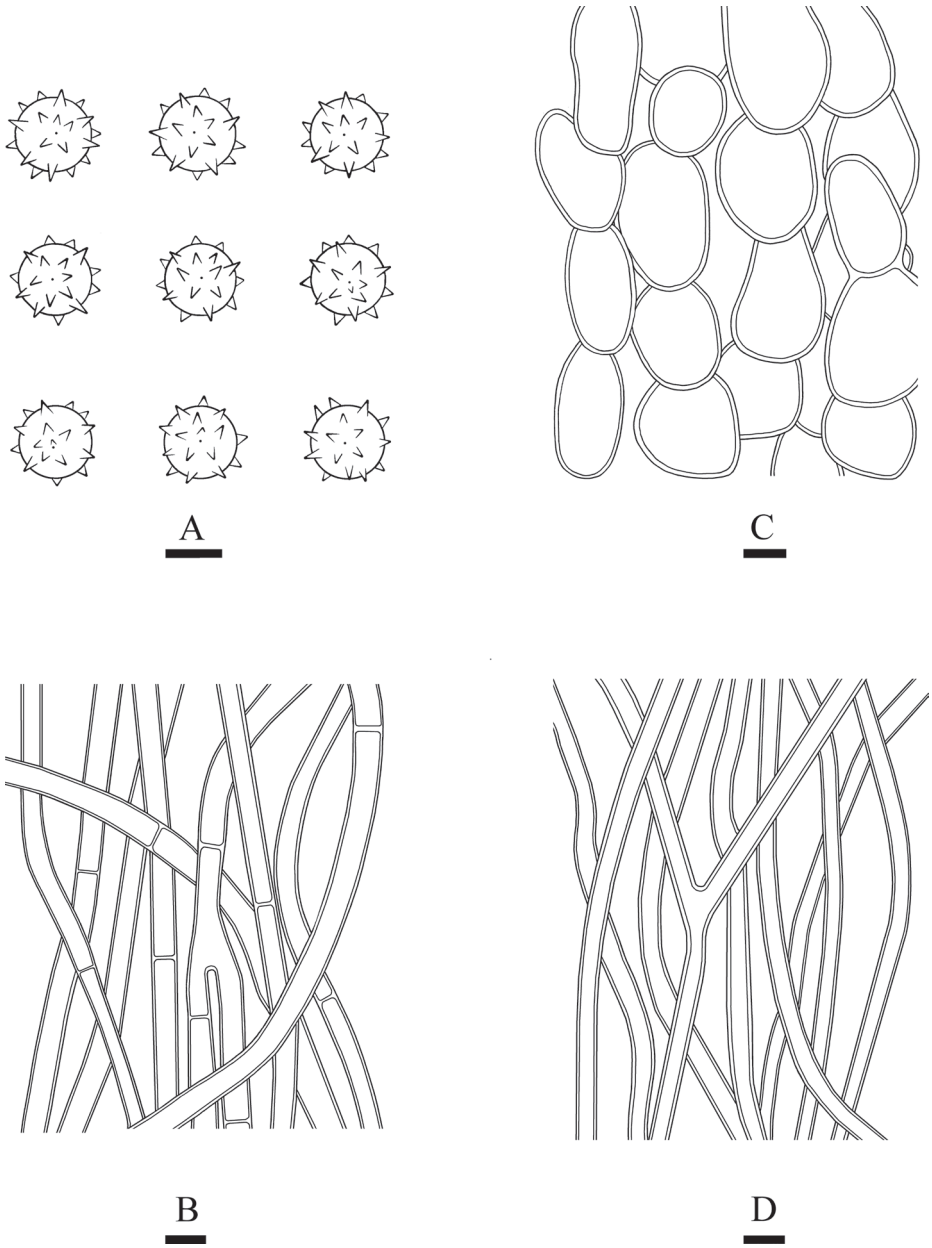

Figure 9. *Morganella minima* (CLZhao 40537) **A** basidiospores **B** paracapillitium **C** exoperidial elements **D** endoperidial hyphae. Scale bars: 1 μm (**A**); 10 μm (**B–D**).

Discussion

In the family Lycoperdaceae, several morphological characteristics act as distinguishing features for its genera, which include the existence and type of capillitium, the presence or lack of paracapillitium, the existence and structure of subgleba, the opening modes of the endoperidium, and the anatomy of the peridium (Larsson and Jeppson 2008; Bates et al. 2009; Larsson et al. 2009; Jeppson and Larsson 2010; Kim et al. 2016; Krakhmalnyi et al. 2023; Li et al. 2024). In the present study, one new genus, *Lycoperdia*, and three new species, *Calvatia phlebioides*, *Lycoperdia tomentosa*, and *Morganella minima*, are described based on phylogenetic analyses and morphological characteristics.

Phylogenetically, the phylogram based on the combined ITS+nLSU sequences (Fig. 1) showed that one new genus, *Lycoperdia*, formed a mono-

phyletic clade, and the new species, *Lycoperdia tomentosa*, was assigned to the genus *Lycoperdia* within the family Lycoperdaceae. However, morphologically, *Lycoperdon* differs from *Lycoperdia* in its peridium dehiscing by a definite apical stoma (Cunningham 1926b).

Based on the combined ITS+nLSU+*rpb2* sequences (Fig. 2), the phylogram showed that *Calvatia phlebioides* was assigned to the genus *Calvatia*, in which *C. phlebioides* was retrieved as a sister to *C. longisetulosa*. However, morphologically the species *C. longisetulosa* differs from *C. phlebioides* by having the exoperidium densely covered with robust spines and longer pedicels present (0.5–1 μm; Li et al. 2024).

Phylogenetically, the phylogram based on the combined ITS+nLSU sequences (Fig. 3) showed that *Morganella minima* was assigned to the genus *Morganella* and formed a monophyletic lineage with *M. oblongata* (Accioly, Baseia & M.P. Martín) R.L. Zhao & J.X. Li. However, morphologically, *M. oblongata* differs from *M. minima* by having larger basidiomes (10 mm in height, 15.5 mm in diameter; Alfredo et al. 2017).

Morphologically, *Lycoperdia* resemble the other fourteen genera in the family Lycoperdaceae: *Abstoma*, *Apioperdon*, *Bovista*, *Bovistella* Morgan, *Bryoperdon*, *Calbovista* Morse ex M.T. Seidl, *Calvatia*, *Disciseda*, *Fuscospina* R.L. Zhao & J.X. Li, *Gastropila*, *Lycoperdon*, *Morganella*, *Pseudoperdon* R.L. Zhao & J.X. Li, and *Sinoperdon* R.L. Zhao & J.X. Li. A morphological comparison between the new genus *Lycoperdia* and the other fourteen genera is presented in Table 3.

Calvatia phlebioides resembles the other ten species in the genus *Calvatia*: *Calvatia bicolor* (Lév.) Kreisel, *C. candida*, *C. craniiformis*, *C. cyathiformis*, *C. fragilis* (Quél.) Morgan, *C. holothurioides* Rebriev, *C. longisetulosa*, *C. nodulata* Alfredo & Baseia, *C. shennongjiaensis*, and *C. subbooniana*. Table 4 presents a morphological comparison between *Calvatia phlebioides* and the other ten species.

Morganella minima resembles the other ten species in the genus *Morganella*, *M. afra* Kreisel & Dring, *M. austromontana* C.R. Alves, Cortez & R.M. Silveira, *M. benjaminii* (Rick) Cortez, Calonge & Baseia, *M. fimbriata* Rebriev, *M. mengsongensis* (L. Ye, P.E. Mortimer, & Karunarathna) R.L. Zhao & J.X. Li, *M. nuda* Alfredo & Baseia, *M. oblongata*, *M. rimosa* Baseia & Alfredo, *M. sosinii* Rebriev & Bulakh, *M. tricolor* R.L. Zhao & J.X. Li. Table 5 presents a morphological comparison between the new species, *Morganella minima*, and the other ten species.

Macrofungi are an important part of forest ecosystems, which are mainly composed of most members of Basidiomycota, in which they possess important economic values and ecological functions (Chen et al. 2015; Wu et al. 2022a, 2022b; Dai et al. 2021; Deng and Zhao 2023; Guan et al. 2023; Wang et al. 2023, 2024; Yuan et al. 2023a; Dong et al. 2024a, b, 2025; Luo et al. 2024; Zhao et al. 2024). The family Lycoperdaceae is an extensively studied group of Basidiomycota (Larsson and Jeppson 2008; Krakhmalnyi et al. 2023). However, the diversity of puffball fungi in China is still not well known, especially in the subtropical and tropical areas. Puffball fungi have huge potential as food and medicines, especially in Asia, and their prospects (Dai and Yang 2008; Dai et al. 2010; Cui et al. 2023). In the present study, three new species are introduced from China, further enriching our knowledge of the macrofungal diversity. We anticipate that more undescribed Lycoperdaceae taxa will be discovered throughout China after extensive collection combined with morphological and molecular analyses.

Table 3. A morphological comparison between the new genus *Lycoperdia* and fourteen other genera in the family Lycoperdaceae.

Species name	*Basidiomes*	Exoperidium	Endoperidium	Gleba	*Capillitium*	*Paracapillitium*	Basidiospores	References
Abstoma	Subglobose, stoma absent.	Fragile; breaking away irregularly.	Fragile; dehisces by irregular rupture.	Firm at maturity.	Calvatia-type, occasionally branched, smooth.	–	Apedicellate, globose.	Cunningham (1926a)
Apioperdon	Obovoid to obpyriform.	–	–	–	Lycoperdon-type.	–	Smooth to minutely ornamented.	Kreisel (1962)
Arachnion	Small-sized;	–	Disintegrates at maturity	Develops minute peridioles resembling sand grains	Absent or poorly developed.	–	Exhibit reticulate ornamentation.	Cortez et al. (2010)
Bovista	–	Persistent spines	–		Bovista-type or Bovista-Lycoperdon type.	–	Ellipsoid, pedicellate	Li et al. (2024)
Bovistella	Medium-sized; distinct pseudo-diaphragm	–	–	–	Fragile; more or less abundant pits of irregular outline.	–	Subglobose to slightly ellipsoid	Larsson and Jeppson (2008)
Bryoperdon	Small, ovoid, with mycelial cords;	–	–	–	Lycoperdon-type.	–	Smooth to minutely pustulose-verrucose	Vizzini and Ercole (2017)
Calbovista	Medium to large; top-shaped	Coriaceous, fall away from top downward at maturity	–	Fragile, dark umber at maturit	Abundant discrete, ochraceous yellow, antler-like.	–	–	Morse (1935)
Calvatia	Globose to pyriform, turbinate.	Dehiscence of peridium occurring by irregular fragmentation.	–	Pulverulent to cottony.	Calvatia-type.	–	Smooth to verrucose and echinate.	Li et al. (2024)
Disciseda	Globose to globose-depressed.	Covered by a sand case.	–	–	Calvatia-type; commonly wavy.	–	Smooth to verrucose, and shortly pedicellate.	Li et al. (2024)
Fuscospina	Lycoperdoid; dark brown appearance	Adorned with conical and curved warts.	Sessile; 6–20 mm in diam; obvious oral margin ring.	–	Lycoperdon-type capillitium with pits.	–	covered with rounded warts.	Li et al. (2024)
Gastropila	Almost globose, dehiscing in an irregularly radiate-stellate manner.	Fragile.	Corky-spongy.	–	Smooth threads, sparsely branched, not easily broken, much entangled.	–	Smooth.	Homrich and Wright (1973)
Lycoperdon	–	Echinate.	Smooth.	–	–	–	Globose to subglobose, usually ornamented with minute warts.	Li et al. (2024)
Lycoperdia	Pyriform, and subglobose to globose when dry.	Densely arranged tomentose structures.	Papery and fragile.	Cottony texture when dry.	Lycoperdon-type, branched, thick-walled, no septa.	Composed of chains of colorless, inflated cells, branched, no septa.	Globose to subglobose, with distinct spines, with a short pedicel.	Present study
Morganella	Depressed, globose to pyriform.	–	–	Consists of abundant paracapillitium.	Absent.	Abundant.	Asperulate to echinulate.	Li et al. (2024)
Pseudoperdon	Globose to subglobose	–	–	–	Lycoperdon-type, with abundant large irregular pores	–	–	Li et al. (2024)
Sinoperdon	Lycoperdoid.	Echinate exoperidium covred with conical spines.	–	–	Lycoperdon-type, thick walls.	Abundant.	Verrucose; long pedicels	Li et al. (2024)

Table 4. A morphological comparison between the new species *Calvatia phlebioides* and ten other species in the genus *Calvatia*.

Species name	Basidiomes	Exoperidium	Endoperidium	Gleba	*Capillitium*	Basidiospores	References
Calvatia bicolor	35–68 mm in height, 41–103 mm in diameter; subglobose to globose.	Thin; membranous and fragile; brownish orange to light brown.	Thin; smooth; reddish blond to brownish orange; dehiscing by irregular rupture of whole peridium.	Lanose; persistent; yellowish-brown at maturity.	2.5–6 µm in diameter; smooth and thick-walled; dichotomously branched; septate, without pores.	4.2–5 µm in diameter; globose; strongly echinate (<1.5 µm) long; shortly pedicellate (<1 µm).	Cortez et al. (2012)
Calvatia candida	15–50 mm in height, 20–60 mm in diameter; subglobose, depressed globose to turbinate.	Thin and papery; glabrous to minutely floccose; grayish orange to orange.	Thin and fragile; felted; grayish orange.	Cottony, rather firm; yellow.	3.2–4.8 µm in diameter; thick-walled; straight to subundulate, with abundant pores.	4.0–4.8 × 4.0–4.8 µm; globose to subglobose; echinate; shortly pedicellate (<1.6 µm).	Bates et al. (2009)
Calvatia craniiformis	60–120 mm in height, 70–150 mm in diameter; obovoid, broadly obpyriform to turbinate.	Thin and papery; glabrous to minutely floccose or sparsely encrusted with particles of dirt; white to off-white at first, becoming dark yellow to yellowish brown.	Thin and fragile; glabrous or felted.	Cottony and firm; white to off-white and solid at first, becoming olive yellow, olive-brown to light brown.	4.8 µm in diameter; thick-walled; straight to subundulate, glabrous with abundant pores, septate.	3.2–4.0 × 3.2–4.0 µm; globose to subglobose; smooth to asperulate.	Bates et al. (2009)
Calvatia cyathiformis	80–130 mm in height, 70–130 mm in diameter; obovoid, subglobose, depressed globose, broadly obpyriform to turbinate.	Thin and papery; glabrous to minutely floccose or sparsely encrusted with particles of dirt; white to off-white at first, becoming light brown to brown or remaining white, finally turning violet-brown to dark magenta.	Thin and fragile; felted; violet-brown to dull violet or purplish gray.	Cottony, rather firm; white to off-white and solid at first, becoming violet-brown to dark magenta or grayish magenta.	3.2–6.4 µm in diameter; thick-walled; straight to subundulate, glabrous with abundant pores, septate.	5.6–8.0 × 5.6–8.0 µm; globose; strongly and densely verrucose (<1.0 µm).	Bates et al. (2009)
Calvatia fragilis	35–60 mm in height, 40–70 mm in diameter; subglobose, depressed globose to turbinate.	Thin and papery; glabrous to appressed floccose; white to off-white at first, becoming pale yellow or remaining white, finally turning violetbrown as the peridia begin to disintegrate.	Thin and fragile; felted; gray to violet-brown.	Cottony, rather firm; white to off-white and solid at first, becoming grayish magenta to dull violet.	3.2–4 µm in diameter; thick-walled; straight to subundulate, glabrous with abundant pores, septate.	6.4–7.2 × 6.4–7.2 µm; globose; verrucose (<0.8 µm).	Bates et al. (2009)
Calvatia holothurioides	40–50 mm in height, 30–55 mm in diameter; pyriform, turbinate to broadly excipuliform.	Thin, fragile, and tomentose; yellow-orange to fulvous.	olive-brown.	cottony, fulvous.	2–4 µm in diameter; thin-walled; branch, septate.	3.4–4.5 × 2.3–3.1 µm; globose; spines (0.4–0.6 µm high); without Pedicels.	Rebriev (2013)
Calvatia longisetulosa	20–35 mm in height, 24–40 mm in diameter; pyriform, tdepressed globose.	densely with robust spines, coniform; yellowish-white, yellowish-brown, dark brown to reddish-brown in upper part.	Papery and fragile; whitish.	pulverulent, cottony; initially whitish, grayish brown, light-brown.	1.4–3.6 µm in diameter; thick-walled; straight, occasionally branched and septa.	3.3–3.7 µm in diameter; globose; distinctly ornamented echinate; short pedicels present (0.5–1 µm).	Li et al. (2024)
Calvatia nodulata	29–56 mm in height, 24–55 mm in diameter; pyriform to turbinate.	granulose to pilose, not persistent, brown in base becoming olive brown in apex.	papery, surface smooth to wrinkly, plicate; light brown to brown.	initially cottony becoming powdery olive brown at maturity.	2–4 µm in diameter; thick-walled; occasionally branched and septate, rare circular pits.	3.1–4.7 × 2.9–4.6 µm in diameter; subovoid to ampulliformis, punctate, pedicels (0.5–3.7 µm).	Alfredo et al. (2014)

Species name	Basidiomes	Exoperidium	Endoperidium	Gleba	*Capillitium*	Basidiospores	References
Calvatia phlebioides	45–65 mm in height, 30–50 mm in diameter; broadly obpyriform to turbinate	Brownish to dark brownish when mature, grayish-brown when dry.	Fragile; bluish-gray when fresh, ash-gray to bluish-gray when dry.	Cottony; pale brown to olive-yellow when mature.	2.5–3.6 µm in diameter; rare branched, septate, thick-walled.	2.8–3.5 × 2.7–3.3 µm in diameter; spinose (< 1 µm), with a short pedicel (< 0.5 µm).	Present study
Calvatia shennongjiaensis	30 mm in height, 35 mm in diameter; broadly obpyriform to turbinate.	initially whitish, ochraceous to dark brownish when mature.	papery and fragile; pale yellow.	Cottony; initially whitish, pale brown to olive-yellow when mature.	straight, rare branched, septate; fragile.	3.2–4.0 µm in diameter; globose to subglobose; spinose (<2 µm); short pedicel (<3 µm).	Li et al. (2024)
Calvatia subbooniana	30–50 mm in height, 30–80 mm in diameter; globose, depressed globose, subovate.	Thin; reenish-olive or sometimes light reddish-brown.	Papery; whitish.	Pulverulent, yellowish-white to dark olive-brown when mature.	2.4–7.2 µm in diameter; thick-walled; occasionally branched and septate, constricted at the septa.	5.2–6.6 × 3.7–5.1 µm in diameter; ovoid, oblong; smooth; short pedicel (<0.5 µm).	Li et al. (2024)

Table 5. A morphological comparison between the new species *Morganella minima* and the other ten species in the genus *Morganella*.

Species name	***Basidiomes***	Exoperidium	Endoperidium	Gleba	***Paracapillitium***	Basidiospores	References
Morganella afra	10 mm in diameter; depressed globose to pyriform.	Minutely granular; fuscous above, becoming lighter below.	Thin; smooth to very minutely areolate; fawn.	Becoming grayish as the spores are shed.	–	3.5–4.5 µm in diameter; globose; minutely asperulate to short-spined.	Kreisel and Dring (1967)
Morganella austromontana	10 mm in height, 8–20 mm in diameter; subglobose to pyriform.	Spiny, formed of short spines (<1 mm high); light brown when fresh.	Papery; smooth to areolate; white when fresh; yellow gray after dried.	Cottony; brownish gray.	7–10 µm in diameter; infated; with numerous small pores.	4–5 × 3–4 µm in diameter; ovoid to subglobose; spiny under SEM (<2 µm).	Alves et al. (2017)
Morganella benjaminii	5–10 mm in diameter; globose, sessile; grayish orange.	Composed of minute spines.	Smooth; fragile.	–	2–4 µm in diameter; thin-walled; more or less branched, smooth, septate.	2.5–3.5 µm in diameter; globose; verrucose, with a pedicel (<1.5 µm).	Cortez et al. (2017)
Morganella fimbriata	10–15 mm in height, 11–16 mm in diameter; turbinate to pyriform.	Dark-brown to cream-brown slender spines (0.5–1.5 mm).	Light grayish to cream-brown.	Cream-brown.	5–7 µm in diameter; rarely bifurcated.	2.9–3.4 µm in diameter; globose; verruculose under the SEM, with a pedicel (5–7 µm).	Rebriev (2023)
Morganella mengsongensis	11–22 mm in height, 10–23 mm in diameter; globose or depressed globose.	Minute conical tubercles; grayish white to dark gray.	Thin, papery, smooth to wrinkled with age; white to dark brown.	Cottony and white when young, powdery and light brown when mature.	2.7–3.2 µm in diameter; septate, occasionally branched.	2.8–3.7 µm in diameter; subglobose or globose; echinate, spine (<0.5 µm).	Ye et al. (2022)
Morganella minima	3–7 mm in height, 5–8 mm in diameter; subglobose.	Hymenial surface with coarse granular; slightly brown when fresh, grayish-brown to fuscous when dry	Fragile; cream to slightly brown when fresh, slightly brown upon drying	Fuscous; powdery	4–7 µm in diameter; thick-walled; branched.	4.7–5.5 × 4.6–5.5 µm in diameter; globose to subglobose; ornamented with distinct spines (< 1 µm), with a short pedicel (< 0.5 µm).	Present study
Morganella nuda	depressed globose to pyriform.	Ornamentation granulose.	Surface smooth; orange when young to yellowish brown.	Powdery; brown.	–	5.5–7.5 µm in diameter; globose; aculeate in SEM (0.5–1 µm).	Alfredo et al. (2017)
Morganella oblongata	10 mm in height, 15.5 mm in diameter; epigeous, depressed globose to subglobose, slightly umbonate.	Formed by a tomentum organized in tufts sphaerocysts; light yellow to light orange.	Smooth; cream.	Cottony; furfuraceous; grayish orange to brownish orange.	3.2–6.3 µm in diameter; branched.	5.4–6.6 µm in diameter; globose; aculeate (<1 µm).	Alfredo et al. (2017)

Species name	*Basidiomes*	Exoperidium	Endoperidium	Gleba	*Paracapillitium*	Basidiospores	References
Morganella rimosa	45–55 mm in height, 50–60 mm in diameter; globose to subglobose.	rimose, granulose; pale yellow.	Smooth; pale yellow.	Reddish yellow.	1.5–2 µm in diameter; septate and branched.	2–3 µm in diameter; globose, subglobose to ovoid; verrucose to equinulate, apiculate.	Alfredo et al. (2012)
Morganella sosinii	7–15 mm in height, 7–20 mm in diameter; subglobose, depressed-globose.	Consists of granules and echinulate; dark-brown to blackish.	Papery thin; brown, to dark-brown.	Olive brown.	4–6 µm in diameter; septate and unbranched.	2–3 µm in diameter; globose, subglobose to ovoid; verrucose to equinulate, apiculate.	Rebriev and Bulakh (2015)
Morganella tricolor	10–13 mm in diameter; subglobose, depressed-globose.	Surface with coarse granular; yellowish to brownish at the upperpart, deep brownish at apical part.	Smooth, fragile; off-white.	Powdery or fibrous; brown.	3.1–5.7 µm in diameter; straight without branches; septa abundant.	3.2–3.6 µm in diameter; globose to subglobose; densely ornamented coarse verrucae, cylindrical or inverted pyramid.	Li et al. (2024)

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Additional information

Conflict of interest

The authors have declared that no competing interests exist.

Ethical statement

No ethical statement was reported.

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Author contributions

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Author ORCIDs

Xin Yang https://orcid.org/0009-0009-7755-0581
Songjing Duan https://orcid.org/0009-0005-3122-0318
Mei Li https://orcid.org/0009-0003-6281-4990
Dongxu Li https://orcid.org/0009-0008-3991-1844
Rongcong Yang https://orcid.org/0009-0006-0646-5265
Shihui Zhang https://orcid.org/0009-0008-3309-4214

Taimin Xu https://orcid.org/0000-0002-5230-4140
Wen Li https://orcid.org/0009-0005-1394-4945
Hongmin Zhou https://orcid.org/0000-0002-0724-5815
Changlin Zhao https://orcid.org/0000-0002-8668-1075

Data availability

All of the data that support the findings of this study are available in the main text.

References

Alfredo DS, Leite AG, Braga-Neto R, Baseia IG (2012) Two new *Morganella* species from the Brazilian Amazon rainforest. Mycosphere 3: 66–71. https://doi.org/10.5943/mycosphere/3/1/8

Alfredo DS, Rodrigues ACM, Baseia IG (2014) *Calvatia nodulata*, a new gasteroid fungus from Brazilian semiarid region. Journal of Mycology 2014: 1–7. https://doi.org/10.1155/2014/697602

Alfredo DS, Baseia IG, Accioly T, Silva BDB, Moura MP, Marinho P, Martín MP (2017) Revision of species previously reported from Brazil under *Morganella*. Mycological Progress 16: 965–985. https://doi.org/10.1007/s11557-017-1332-y

Altaf U, Singh U, Verma K, Sharma YP (2022) A new species of *Abstoma* (Agaricaceae – from Kashmir Himalaya, India. Nova Hedwigia 115: 89–98. https://doi.org/10.1127/nova_hedwigia/2022/0706

Alves CR, Cortez VG, Silveira RMB (2017) *Morganella austromontana* sp. nov. from the south Brazilian plateau. Mycotaxon 132: 281–287. https://doi.org/10.5248/132.281

Bates ST (2004) Arizona members of the Geastraceae and Lycoperdaceae (Basidiomycota, Fungi). Arizona State University Tempe.

Bates ST, Roberson RW, Desjardin DE (2009) Arizona gasteroid fungi I: Lycoperdaceae (Agaricales, Basidiomycota). Fungal Diversity 37: 153–207.

Case NT, Gurr SJ, Fisher MC, Blehert DS, Boone C, Casadevall A, Chowdhary A, Cuomo CA, Currie CR, Denning DW, Ene IV, Fritz-Laylin LK, Gerstein AC, Gow NAR, Gusa A, Iliev ID, James TY, Jin H, Kahmann R, Klein BS, Kronstad JW, Ost KS, Peay KG, Shapiro RS, Sheppard DC, Shlezinger N, Stajich JE, Stukenbrock EH, Taylor JW, Wright GD, Cowen LE, Heitman J, Segre JA (2025) Fungal impacts on Earth's ecosystems. Nature 638: 49–57. https://doi.org/10.1038/s41586-024-08419-4

Chen JJ, Cui BK, Zhou LW, Korhonen K, Dai YC (2015) Phylogeny, divergence time estimation, and biogeography of the genus *Heterobasidion* (Basidiomycota, Russulales). Fungal Diversity 71: 185–200. https://doi.org/10.1007/s13225-014-0317-2

Coetzee JC, van Wyk AE (2009) The genus *Calvatia* ('Gasteromycetes', Lycoperdaceae): A review of its ethnomycology and biotechnological potential. African Journal of Biotechnology 8: 6007–6015. https://doi.org/10.5897/AJB09.360

Cortez VG, Baseia IG, Silveira RMB (2010) Gasteroid mycobiota of Rio Grande do Sul, Brazil: *Arachnion* and *Disciseda* (Lycoperdaceae). Acta Biológica Paranaense 39: 19–27. https://doi.org/10.5380/abpr.v39i0.20238

Cortez VG, Baseia IG, Silveira RMB (2012) Gasteroid mycobiota of Rio Grande do Sul, Brazil: *Calvatia*, *Gastropila* and *Langermannia* (Lycoperdaceae). Kew Bulletin 67: 471–482. https://doi.org/10.1007/s12225-012-9364-6

Cortez VG, Calonge FD, Baseia IG (2017) Rick's species revision 2: *Lycoperdon benjaminii* recombined in *Morganella*. Mycotaxon 102: 425–429.

Cui YY, Cai Q, Tang LP, Liu JW, Yang ZL (2018) The family Amanitaceae: Molecular phylogeny, higher-rank taxonomy and the species in China. Fungal Diversity 91: 5–230. https://doi.org/10.1007/s13225-018-0405-9

Cui BK, Li HJ, Ji X, Zhou JL, Song J, Si J, Yang ZL, Dai YC (2019) Species diversity, taxonomy and phylogeny of Polyporaceae (Basidiomycota) in China. Fungal Diversity 97: 137–392. https://doi.org/10.1007/s13225-019-00427-4

Cui BK, Pan XH, Pan F, Sun YF, Xing JH, Dai YC (2023) Species diversity and resources of *Ganoderma* in China. Junwu Xuebao 42: 170–178.

Cunningham GH (1926a) Development of *Lycoperdon depressum* (fungi). New Zealand Journal of Science and Technology 8: 228–232.

Cunningham GH (1926b) Lycoperdaceae of New Zealand. Transactions and Proceedings of the New Zealand Institute 57: 187–217.

Dai YC, Yang ZL (2008) A revised checklist of medicinal fungi in China. Junwu Xuebao 27: 801–824.

Dai YC, Zhou LW, Yang ZL, Wen HA, Bao T, Li TH (2010) A revised checklist of edible fungi in China. Junwu Xuebao 29: 1–21.

Dai YC, Yang ZL, Cui BK, Wu G, Yuan HS, Zhou LW, He SH, Ge ZW, Wu F, Wei YL, Yuan Y, Si J (2021) Diversity and systematics of the important macrofungi in Chinese forests. Junwu Xuebao 40: 770–805. https://doi.org/10.13346/j.mycosystema.210036

Deng YL, Zhao CL (2023) The molecular phylogeny and morphology revealed a new wood-rotting fungus *Vararia yunnanensis* (Peniophoraceae, Russulales) in Yunnan Province, China. Phytotaxa 583: 39–49. https://doi.org/10.11646/phytotaxa.583.1.4

Dong JH, Li Q, Yuan Q, Luo YX, Zhang XC, Dai YF, Zhou Q, Liu XF, Deng YL, Zhou HM, Muhammad A, Zhao CL (2024a) Species diversity, taxonomy, molecular systematics and divergence time of wood-inhabiting fungi in Yunnan-Guizhou Plateau, Asia. Mycosphere 15: 1110–1293. https://doi.org/10.5943/mycosphere/15/1/10

Dong JH, Zhu YG, Qian CB, Zhao CL (2024b) Taxonomy and phylogeny of Auriculariales (Agaricomycetes, Basidiomycota) with descriptions of four new species from southwestern China. MycoKeys 108: 115–146. https://doi.org/10.3897/mycokeys.108.128659

Dong JH, Deng QH, Chen M, Chen DX, Zhou CQ, Zhao CL (2025) Molecular phylogeny and morphology reveal four new wood-inhabiting fungi of *Asterostroma* and *Radulomyces* (Basidiomycota) from Southwestern China. MycoKeys 112: 35–58. https://doi.org/10.3897/mycokeys.112.137098

Guan QX, Huang J, Huang J, Zhao CL (2023) Five new species of Schizoporaceae (Basidiomycota, Hymenochaetales) from East Asia. MycoKeys 96: 25–56. https://doi.org/10.3897/mycokeys.96.99327

Gube M, Piepenbring M (2009) Preliminary annotated checklist of Gasteromycetes in Panama. Nova Hedwigia 89: 519–543. https://doi.org/10.1127/0029-5035/2009/0089-0519

Guindon S, Gascuel O (2003) A simple, fast and accurate algorithm to estimate large phylogenies by maximum likelihood. Systematic Biology 52: 696–704. https://doi.org/10.1080/10635150390235520

He MQ, Zhao RL, Hyde KD, Begerow D, Kemler M, Yurkov A, McKenzie EHC, Raspé O, Kakishima M, Sánchez-Ramírezll S, Vellinga EC, Halling R, Papp V, Zmitrovich IV, Buyck B, Ertz D, Wijayawardene NN, Cui BK, Schoutteten N, Liu XZ, Li TH, Yao YJ, Zhu XY, Liu AQ, Li GJ, Zhang MZ, Ling ZL, Cao B, Antonín V, Boekhout T, Silva BDB, Crop ED, Decock C, Dima B, Dutta AK, Fell JW, Geml J, Ghobad-Nejhad M, Giachini AJ, Gibertoni TB, Gorjón SP, Haelewaters D, He SH, Hodkinson BP, Horak E, Hoshino T, Justo A, Lim

YW, Menolli N, Mešić A, Moncalvo JM, Mueller GM, Nagy LG, Nilsson LK, Noordeloos M, Nuytinck J, Orihara T, Ratchadawan C, Rajchenberg M, Silva-Filho AGS, Sulzbacher MA, Tkalčec Z, Valenzuela R, Verbeken A, Vizzini A, Wartchow F, Wei TZ, Weiß M, Zhao CL, Kirk PM (2019) Notes, outline and divergence times of Basidiomycota. Fungal Diversity 99: 105–367. https://doi.org/10.1007/s13225-019-00435-4

He MQ, Cao B, Liu F, Boekhout T, Denchev TT, Schoutteten N, Denchev CM, Kemler M, Gorjón SP, Begerow D, Valenzuela R, Davoodian N, Niskanen T, Vizzini A, Redhead SA, Ramírez-Cruz V, Papp V, Dudka VA, Dutta AK, García-Sandoval R, Liu XZ, Kijpornyongpan T, Savchenko A, Tedersoo L, Theelen B, Trierveiler-Pereira L, Wu F, Zamora JC, Zeng XY, Zhou LW, Liu SL, Ghobad-Nejhad M, Giachini AJ, Li GJ, Kakishima M, Olariaga I, Haelewaters D, Sulistyo B, Sugiyama J, Svantesson S, Yurkov A, Alvarado P, Antonín V, da Silva AF, Druzhinina I, Gibertoni TB, Guzmán-Dávalos L, Justo A, Karunarathna SC, Galappaththi MCA, Toome-Heller M, Hosoya T, Liimatainen K, Márquez R, Mešić A, Moncalvo JM, Nagy LG, Varga T, Orihara T, Raymundo T, Salcedo I, Silva-Filho AGS, Tkalčec Z, Wartchow F, Zhao CL, Bau T, Cabarroi-Hernández1 M, Cortés-Pérez A, Decock C, Lange RD, Weiss M, Menolli Jr N, Nilsson RH, Fan YG, Verbeken A, Gaforov Y, Meiras-Ottoni A, Mendes-Alvarenga RL, Zeng NK, Wu Q, Hyde KD, Kirk PM, Zhao RL (2024) Phylogenomics, divergence times and notes of orders in Basidiomycota. Fungal Diversity 99: 105–367. https://doi.org/10.1007/s13225-024-00535-w

He SY, Wang L, Shen KZ, Zhou HM (2025) Morphological characteristics and phylogenetic analyses revealed four new species (Basidiomycota) in the Yunnan-Guizhou Plateau, China. MycoKeys 113: 237–262. https://doi.org/10.3897/mycokeys.113.140932

Hibbett DS, Pine EM, Langer E, Langer G, Donoghue M (1997) Evolution of gilled mushrooms and puffballs inferred from ribosomal DNA sequences. Proceedings of the National Academy of Sciences of the United States of America 94: 12002–12006. https://doi.org/10.1073/pnas.94.22.12002

Homrich MH, Wright JE (1973) South American Gasteromycetes. The genera Gastropila, Lanopila and Mycenastrum. Mycologia 65: 779–794. https://doi.org/10.1080/00275514.1973.12019493

Hu Y, Karunarathna SC, Li H, Galappaththi MC, Zhao CL, Kakumyan P, Mortimer PE (2022) The impact of drying temperature on basidiospore size. Diversity 14: 239. https://doi.org/10.3390/d14040239

Hyde KD (2022) The numbers of fungi. Fungal Diversity 114: 1. https://doi.org/10.1007/s13225-022-00507-y

Hyde KD, Noorabadi MT, Thiyagaraja V, He MQ, Johnston PR, Wijesinghe SN, Armand A, Biketova AY, Chethana KWT, Erdoğdu M, Ge ZW, Groenewald JZ, Hongsanan S, Kušan I, Leontyev DV, Li DW, Lin CG, Liu NG, Maharachchikumbura SSN, Matočec N, May TW, McKenzie EHC, Mešić A, Perera RH, Phukhamsakda C, Piątek M, Samarakoon MC, Selcuk F, Senanayake IC, Tanney JB, Tian Q, Vizzini A, Wanasinghe DN, Wannasawang N, Wijayawardene NN, Zhao RL, Abdel-Wahab MA, Abdollahzadeh J, Abeywickrama PD, Abhinav, Absalan S, Acharya K, Afshari N, Afshan NS, Afzalinia S, Ahmadpour SA, Akulov O, Alizadeh A, Alizadeh M, Al-Sadi AM, Alves A, Alves VCS, Alves-Silva G, Antonín V, Aouali S, Aptroot A, Apurillo CCS, Arias RM, Asgari, B, Asghari R, Assis DMA, Assyov B, Atienza V, Aumentado HDR, Avasthi S, Azevedo E, Bakhshi M, Bao DF, Baral HO, Barata M, Barbosa KD, Barbosa RN, Barbosa FR, Baroncelli R, Barreto GG, Baschien C, Bennett RM, Bera I, Bezerra JDP, Bhunjun CS, Bianchinotti MV, Błaszkowski J, Boekhout T, Bonito GM, Boonmee S, Boonyuen N, Bortnikov FM, Bregant C, Bundhun D, Burgaud G, Buyck B, Caeiro MF, Cabarroi-Hernández M, Cai M Feng, Cai L, Calabon MS, Calaça FJS, Callalli M, Câmara MPS, Cano-Lira J, Cao

B, Carlavilla JR, Carvalho A, Carvalho TG, Castañeda-Ruiz RF, Catania MDV, Cazabonne J, Cedeño-Sanchez M, Chaharmiri-Dokhaharani S, Chaiwan N, Chakraborty N, Cheewankoon R, Chen C, Chen J, Chen Q, Chen YP, Chinaglia S, Coelho-Nascimento CC, Coleine C, CostaRezende DH, Cortés-Pérez A, Crouch, JA, Crous PW, Cruz RHSF, Czachura P, Damm U, Darmostuk V, Daroodi Z, Das K, Das K, Davoodian N, Davydov EA, da Silva GA, da Silva IR, da Silva RMF, da Silva Santos AC, Dai DQ, Dai YC, de Groot Michiel D, De Kesel A, De Lange R, de Medeiros EV, de Souza CFA, de Souza FA, dela Cruz TEE, Decock C, Delgado G, Denchev CM, Denchev TT, Deng YL, Dentinger BTM, Devadatha B, Dianese JC, Dima B, Doilom M, Dissanayake AJ, Dissanayake DMLS, Dissanayake LS, Diniz AG, Dolatabadi S, Dong JH, Dong W, Dong ZY, Drechsler-Santos ER, Druzhinina IS, Du TY, Dubey MK, Dutta AK, Elliott TF, Elshahed MS, Egidi E, Eisvand P, Fan L, Fan X, Fan XL, Fedosova AG, Ferro LO, Fiuza PO, Flakus A, W. Fonseca EO, Fryar SC, Gabaldón T, Gajanayake AJ, Gannibal PB, Gao F, GarcíaSánchez D, García-Sandoval R, Garrido-Benavent I, Garzoli L, Gasca-Pineda J, Gautam AK, Gené J, Ghobad-Nejhad M, Ghosh A, Giachini AJ, Gibertoni TB, Gentekaki E, Gmoshinskiy VI, GóesNeto A, Gomdola D, Gorjón SP, Goto BT, Granados-Montero MM, Griffith GW, Groenewald M, Grossart H-P, Gu ZR, Gueidan C, Gunarathne A, Gunaseelan S, Guo SL, Gusmão LFP, Gutierrez AC, Guzmán-Dávalos L, Haelewaters D, Haituk H, Halling RE, He SC, Heredia G, HernándezRestrepo M, Hosoya T, Hoog SD, Horak E, Hou CL, Houbraken J, Htet ZH, Huang SK, Huang WJ, Hurdeal VG, Hustad VP, Inácio CA, Janik P, Jayalal RGU, Jayasiri SC, Jayawardena RS, Jeewon R, Jerônimo GH, Jin J, Jones EBG, Joshi Y, Jurjević Ž, Justo A, Kakishima M, Kaliyaperumal M, Kang GP, Kang JC, Karimi O, Karunarathna SC, Karpov SA, Kezo K, Khalid AN, Khan MK, Khuna S, Khyaju S, Kirchmair M, Klawonn I, Kraisitudomsook N, Kukwa M, Kularathnage ND, Kumar S, Lachance MA, Lado C, Latha KPD, Lee HB, Leonardi M, Lestari AS, Li C, Li H, Li J, Li Q, Li Y, Li YC, Li YX, Liao CF, Lima JLR, Lima JMS, Lima NB, Lin L, Linaldeddu BT, Linn MM, Liu F, Liu JK, Liu JW, Liu S, Liu SL, Liu XF, Liu XY, Longcore JE, Luangharn T, Luangsa-ard JJ, Lu L, Lu YZ, Lumbsch HT, Luo L, Luo M, Luo ZL, Ma J, Madagammana AD, Madhushan A, Madrid H, Magurno F, Magyar D, Mahadevakumar S, Malosso E, Malysh JM, Mamarabadi M, Manawasinghe IS, Manfrino RG, Manimohan P, Mao N, Mapook A, Marchese P, Marasinghe DS, Mardones M, Marin-Felix Y, Masigol H, Mehrabi M, MehrabiKoushki M, Meiras-Ottoni A de, Melo RFR, Mendes-Alvarenga RL, Mendieta S, Meng QF, Menkis A, Menolli Jr N, Mikšík M, Miller SL, Moncada B, Moncalvo JM, Monteiro JS, Monteiro M, Mora-Montes HM, Moroz EL, Moura JC, Muhammad U, Mukhopadhyay S, Nagy GL, Najam ul Sehar A, Najafiniya M, Nanayakkara CM, Naseer A, Nascimento ECR, Nascimento SS, Neuhauser S, Neves MA, Niazi AR, Nie Yong, Nilsson RH, Nogueira PTS, Novozhilov YK, Noordeloos M, Norphanphoun C, Nuñez Otaño N, O'Donnell RP, Oehl F, Oliveira JA, Oliveira Junior I, Oliveira NVL, Oliveira PHF, Orihara T, Oset M, Pang KL, Papp V, Pathirana LS, Peintner U, Pem D, Pereira OL, Pérez-Moreno J, Pérez-Ortega S, Péter G, Pires-Zottarelli CLA, Phonemany M, Phongeun S, Pošta A, Prazeres JFSA, Quan Y, Quandt CA, Queiroz MB, Radek R, Rahnama K, Raj KNA, Rajeshkumar KC, Rajwar Soumyadeep, Ralaiveloarisoa AB, Rämä T, Ramírez-Cruz V, Rambold G, Rathnayaka AR, Raza M, Ren GC, Rinaldi AC, Rivas-Ferreiro M, Robledo GL, Ronikier A, Rossi W, Rusevska K, Ryberg M, Safi A, Salimi F, Salvador-Montoya CA, Samant B, Samaradiwakara NP, Sánchez-Castro I, Sandoval-Denis M, Santiago ALCMA, Santos ACDS, Santos LA dos, Sarma VV, Sarwar S. Savchenko A, Savchenko K, Saxena RK, Schoutteten N, Selbmann L, Ševčíková H, Sharma A, Shen HW, Shen YM, Shu YX, Silva HF, Silva-Filho AGS, Silva VSH, Simmons DR, Singh R, Sir EB, Sohrabi M, Souza FA, Souza-Motta CM, Sriindrasutdhi V,

Sruthi OP, Stadler M, Stemler J, Stephenson SL, Stoyneva-Gaertner MP, Strassert JFH, Stryjak-Bogacka M, Su H, Sun YR, Svantesson S, Sysouphanthong P, Takamatsu S, Tan TH, Tanaka K, Tang C, Tang X, Taylor JE, Taylor PWJ, Tennakoon DS, Thakshila SAD, Thambugala KM, Thamodini GK, Thilanga D, Thines M, Tiago PV, Tian XG, Tian WH, Tibpromma S, Tkalčec Z, Tokarev YS, Tomšovský M, Torruella G, Tsurykau A, Udayanga D, Ulukapı M, Untereiner WA, Usman M, Uzunov BA, Vadthanarat S, Valenzuela R, Van den Wyngaert S, Van Vooren N, Velez P, Verma RK, Vieira LC Vieira WAS, Vinzelj JM, Tang AMC, Walker A, Walker AK,Wang QM, Wang Y, Wang XY, Wang ZY, Wannathes N, Wartchow F, Weerakoon G, Wei DP, Wei X, White JF, Wijesundara DSA, Wisitrassameewong K, Worobiec G, Wu HX, Wu N, Xiong YR, Xu B, Xu JP, Xu R, Xu RF, Xu RJ, Yadav S, Yakovchenko LS, Yang HD, Yang X, Yang YH, Yang Y, Yang YY, Yoshioka R, Youssef Noha H, Yu FM, Yu ZF, Yuan LL, Yuan Q, Zabin DA, Zamora JC, Zapata CV, Zare R, Zeng M, Zeng XY, Zhang JF, Zhang JY, Zhang S, Zhang XC, Zhao CL, Zhao H, Zhao Q, Zhao H, Zhao HJ, Zhou HM, Zhu XY, Zmitrovich IV, Zucconi L, Zvyagina E (2024) The 2024 Outline of fungi and fungus-like taxa. Mycosphere 15: 5146–6239. https://doi.org/10.5943/mycosphere/15/1/25

Jeppson M, Larsson E (2010) *Lycoperdon subcretaceum*, a puffball new to Europe found in Norway. Agarica: Mykologisk Tidsskrift 29: 87–92.

Jeppson M, Larsson E, Martín MP (2012) *Lycoperdon rupicola* and *L. subumbrinum*: Two new puffballs from Europe. Mycological Progress 11: 887–897. https://doi.org/10.1007/s11557-011-0804-8

Kalichman J, Kirk PM, Matheny PB (2020) A compendium of generic names of agarics and Agaricales. Taxon 69: 425–447. https://doi.org/10.1002/tax.12240

Katoh K, Rozewicki J, Yamada KD (2019) MAFFT online service: Multiple sequence alignment, interactive sequence choice and visualization. Briefings in Bioinformatics 20(4): 1160–1166. https://doi.org/10.1093/bib/bbx108

Kim CS, Jo JW, Kwag YN, Sung GH, Han JG, Shrestha B, Oh SO, Kim SY, Shin CH, Han SK (2016) Two new *Lycoperdon* species collected from Korea: *L. albiperidium* and *L. subperlatum* spp. nov. Phytotaxa 260: 101–115. https://doi.org/10.11646/phytotaxa.260.2.1

Kirk P, Cannon P, Minter D, Stalpers J (2008) Dictionary of the fungi. CABI, Wallingford, 335 pp.

Krakhmalnyi M, Isikhuemhen OS, Jeppson M, Wasser SP, Nevo E (2023) Species diversity of Lycoperdaceae (Agaricales) in Israel, with some insights into the phylogenetic structure of the family. Journal of Fungi (Basel, Switzerland) 9: 1038. https://doi.org/10.3390/jof9101038

Kreisel H (1962) Die Lycoperdaceae der Deutschen Demokratischen Republik. Feddes Repertorium 64: 89–201.

Kreisel H (1969) Gasteromyzeten aus Nepal. Khumbu Himal 6: 25–36.

Kreisel H, Dring DM (1967) An emendation of the genus *Morganella* Zeller (*Lycoperdaceae*). Feddes Repertorium 74: 109–122. https://doi.org/10.1002/fedr.19670740105

Krüger D, Kreisel H (2003) Proposing *Morganella* subgen. *Apioperdon* subgen. nov. for the puffball *Lycoperdon pyriforme*. Mycotaxon 86: 169–177.

Krüger D, Binder M, Fischer M, Kreisel H (2001) The Lycoperdales. A molecular approach to the systematics of some gasteroid mushrooms. Mycologia 93: 947–957. https://doi.org/10.1080/00275514.2001.12063228

Larsson A (2014) AliView: A fast and lightweight alignment viewer and editor for large data sets. Bioinformatics (Oxford, England) 30: 3276–3278. https://doi.org/10.1093/bioinformatics/btu531

Larsson E, Jeppson M (2008) Phylogenetic relationships among species and genera of Lycoperdaceae based on ITS and LSU sequence data from north European taxa. Mycological Research 112(1): 4–22. https://doi.org/10.1016/j.mycres.2007.10.018

Larsson E, Jeppson M, Larsson KH (2009) Taxomomy, ecology and phylogenetic relationships of *Bovista pusilla* and *B. limosa* in North Europe. Mycological Progress 8: 289–299. https://doi.org/10.1007/s11557-009-0599-z

Li JX, Cao B, Phurbu D, He MQ, Zhu XY, Parra LA, Zhao RL (2024) The revision of the taxonomic system of Lycoperdaceae. Mycosphere 15: 4919–5016. https://doi.org/10.5943/mycosphere/15/1/22

Liu YJ, Whelen S, Hall BD (1999) Phylogenetic relationships among ascomycetes: Evidence from an RNA polymerase II subunit. Molecular Biology and Evolution 16: 1799–1808. https://doi.org/10.1093/oxfordjournals.molbev.a026092

Luo KY, Su JQ, Zhao CL (2024) Morphological and molecular identification for four new wood-inhabiting species of *Trechispora* (Basidiomycota) from China. MycoKeys 105: 155–178. https://doi.org/10.3897/mycokeys.105.120438

Miller MA, Pfeiffer W, Schwartz T (2012) The CIPRES science gateway: Enabling high–impact science for phylogenetics researchers with limited resources. Association for Computing Machinery 39: 1–8. https://doi.org/10.1145/2335755.2335836

Moncalvo JM, Vilgalys R, Redhead SA, Johnson JE, James TY, Aime MC, Hofstetter V, Verduin SJW, Larsson E, Baroni TJ, Thorn RG, Jacobsson S, Clémenc H, Miller OK (2002) One hundred and seventeen clades of euagarics. Molecular Phylogenetics and Evolution 23: 357–400. https://doi.org/10.1016/S1055-7903(02)00027-1

Morse EE (1935) A new puffball. Mycologia 27: 96–101. https://doi.org/10.1080/00275514.1935.12017065

Nylander JAA (2004) *MrModeltest* v2. Program distributed by the author. Evolutionary Biology Centre, Uppsala University.ampignons de l'Équateur (Pugillus IV). Bulletin de l'Herbier Boissier 3: 53–74.

Pegler DN, Spooner BM, Læssøe T (1995) British puffballs, earthstars and stinkhorns: an account of the British gasteroid fungi, 255 pp.

Petersen JH (1996) Farvekort. The Danish Mycological Society's Colour-Chart. Forenin gen til Svampekundskabens Fremme, Greve, Germany, 6 pp.

Posada D (2008) jModelTest: Phylogenetic model averaging. Molecular Biology and Evolution 25: 1253–1256. https://doi.org/10.1093/molbev/msn083

Rebriev YA (2013) *Calvatia holothurioides* sp. nov., new species from Vietnam. Mikologiâ i Fitopatologiâ 47: 21–23.

Rebriev YA (2023) *Morganella fimbriata* (Lycoperdaceae, Agaricomycetes), a new species from Cameroon. Mycology and Phytopathology 57: 25–32. https://doi.org/10.31857/S0026364823010117

Rebriev YA, Bulakh EM (2015) *Morganella sosinii* sp. nov. (Agaricaceae) from the Russian far east. Mycology and phytopathology 49: 293–296.

Ronquist F, Teslenko M, van der Mark P, Ayres DL, Darling A, Hohna S, Larget B, Liu L, Suchard MA, Huelsenbeck JP (2012) Mrbayes 3.2: Efficient bayesian phylogenetic inference and model choice across a large model space. Systematic Biology 61(3): 539–542. https://doi.org/10.1093/sysbio/sys029

Senthilarasu G, Ravindran C, Anjali P (2018) Notes on indian species of *Calvatia* and *Langermannia* including *Calvatia natarajanii* sp. nov. Phytotaxa 362: 160. https://doi.org/10.11646/phytotaxa.362.2.3

Tedersoo L, Bahram M, Põlme S, Kõljalg U, Yorou NS, Wijesundera R, Ruiz LV, Vasco-Palacios AM, Thu PQ, Suija A, Smith ME, Sharp C, Saluveer E, Saitta A, Rosas M, Riit T,

Ratkowsky D, Pritsch K, Põldmaa K, Piepenbring M, Phosri C, Peterson M, Parts K, Pärtel K, Otsing E, Nouhra E, Njouonkou AL, Nilsson RH, Morgado LN, Mayor J, May TW, Majuakim L, Lodge DJ, Lee SS, Larsson KH, Kohout P, Hosaka K, Hiiesalu I, Henkel TW, Harend H, Guo LD, Greslebin A, Grelet G, Geml J, Gates G, Dunstan W, Dunk C, Drenkhan R, Dearnaley J, Kesel AD, Dang T, Chen X, Buegger F, Brearley FQ, Bonito G, Anslan S, Abell S, Abarenkov K (2014) Global diversity and geography of soil fungi. Science 346: 1256688. https://doi.org/10.1126/science.1256688

Vilgalys R, Hester M (1990) Rapid genetic identification and mapping of enzymatically amplified ribosomal DNA from several *Cryptococcus* species. Journal of Bacteriology 172(8): 4238–4246. https://doi.org/10.1128/jb.172.8.4238-4246.1990

Vizzini A, Ercole E (2017) Detecting the phylogenetic position of *Bovista acuminata* (Agaricales, Basidiomycota – by an ITS-LSU combined analysis: The new genus *Bryoperdon* and revisitation of *Lycoperdon* subgen. *Apioperdon*. Phytotaxa 299: 77–86. https://doi.org/10.11646/phytotaxa.299.1.5

Wang CG, Zhao H, Liu HG, Zeng GY, Yuan Y, Dai YC (2023) A multi-gene phylogeny clarifies species diversity, taxonomy, and divergence times of *Ceriporia* and other related genera in Irpicaceae (Polyporales, Basidiomycota). Mycosphere 14: 1665–1729. https://doi.org/10.5943/mycosphere/17/1/9

Wang CG, Dai YC, Kout J, Gates GM, Liu HG, Yuan Y, Vlasák J (2024) Multi-gene phylogeny and taxonomy of *Physisporinus* (Polyporales, Basidiomycota). Mycosphere : Journal of Fungal Biology 15: 1455–1521. https://doi.org/10.1080/21501203.2024.2379476

Wei YL, Dai YC (2004) The ecological function of wood-inhabiting fungi in forest ecosystem. Ying Yong Sheng Tai Xue Bao 15: 1935–1938.

White TJ, Bruns T, Lee S, Taylor J (1990) Amplification and direct sequencing of fungal ribosomal RNA genes for phylogenetics. In: Innis MA, Gelfand DH, Sninsky JJ, White TJ (Eds) PCR protocols: A guide to methods and ap-plications. Academic Press, San Diego, 315–322. https://doi.org/10.1016/B978-0-12-372180-8.50042-1

Wijayawardene NN, Hyde KD, Mikhailov KV, Péter G, Aptroot A, Pires-Zottarelli CLA, Goto BT, Tokarev YS, Haelewaters D, Karunarathna SC, Kirk PM, Santiago ALCMA, Saxena RK, Schoutteten N, Wimalasena MK, Aleoshin VV, Al-Hatmi AMS, Ariyawansa KGSU, Assunção AR, Bamunuarachchige TC, Baral HO, Bhat DJ, Blaszkowski J, Boekhout T, Boonyuen N, Brysch-Herzberg M, Cao B, Cazabonne J, Chen XM, Coleine C, Dai DQ, Daniel HM, Silva SBG, Souza FA, Dolatabadi S, Dubey MK, Dutta AK, Ediriweera A, Egidi E, Elshahed MS, Fan XL, Felix JRB, Galappaththi MCA, Groenewald M, Han LS, Huang B, Hurdeal VG, Ignatieva AN, Jerônimo GH, Jesus AL, Kondratyuk S, Kumla J, Kukwa M, Li QR, Lima JLR, Liu XY, Lu WH, Lumbsch HT, Madrid H, Magurno F, Marson G, McKenzie EHC, Menkis A, Mešić A, Nascimento ECR, Nassonova ES, Nie Y, Oliveira NVL, Ossowska EA, Pawtowska J, Peintner U, Pozdnyakov LR, Premarathne BM, Priyashantha AKH, Quandt CA, Queiroz MB, Rajeshkuma KC, Raza M, Roy N, Samarakoon MC, Santos AA, Santos LA, Schumm F, Selbmann L, Selçuk F, Simmons DR, Simakova AV, Smith MT, Sruthi OP, Suwannarach N, Tanaka K, Tibpromma S, Tomás EO, Ulukap M, Vooren NV, Wanasinghe DN, Weber E, Wu QZ, Yang EF, Yoshioka R, Youssef NH, Zandijk A, Zhang GQ, Zhang JY, Zhao H, Zhao RL, Zverkov OA, Thines M, Karpov SA (2024) Classes and phyla of the kingdom Fungi. Fungal Diversity 127: 1–165. https://doi.org/10.1007/s13225-024-00540-z

Wu F, Zhou LW, Yang ZL, Bau T, Li TH, Dai YC (2019) Resource diversity of Chinese macrofungi: Edible, medicinal and poisonous species. Fungal Diversity 98: 1–76. https://doi.org/10.1007/s13225-019-00432-7

Wu F, Zhou LW, Vlasák J, Dai Y (2022a) Global diversity and systematics of Hymenochaetaceae with poroid hymenophore. Fungal Diversity 113: 1–192. https://doi.org/10.1007/s13225-021-00496-4

Wu F, Man XW, Tohtirjap A, Dai YC (2022b) A comparison of polypore funga and species composition in forest ecosystems of China, North America, and Europe. Forest Ecosystems 9: 100051. https://doi.org/10.1016/j.fecs.2022.100051

Xu Y, Yang Y, Yang X, Chen DX, Zheng W, Shen KZ, Zhang SC, Zhao CL (2025) Molecular phylogeny and taxonomy reveal two new genera and five new species in Phanerochaetaceae (Polyporales) from Yunnan, Southwest China. MycoKeys 113: 263–294. https://doi.org/10.3897/mycokeys.113.140624

Yang X, Zhu YG, Duan SJ, Wu XX, Zhao CL (2025) Morphology and multigene phylogeny revealed four new species of *Geastrum* (Geastrales, Basidiomycota) from China. MycoKeys 113: 73–100. https://doi.org/10.3897/mycokeys.113.139672

Ye L, Karunarathna SC, Li HL, Wanasinghe DN, Kumla J, Xu JC, Galappaththi MCA, Mortimer PE (2022) A new tropical species of *Lycoperdon* subgenus *Morganella* (Agaricales, Basidiomycota) from Yunnan Province, China. Warasan Khana Witthayasat Maha Witthayalai Chiang Mai 49: 641–653. https://doi.org/10.12982/CMJS.2022.051

Yousaf N, Kreisel H, Khalid A (2013) *Bovista himalaica* sp. nov. (gasteroid fungi; Basidiomycetes – from Pakistan. Mycological Progress 12: 569–574. https://doi.org/10.1007/s11557-012-0864-4

Yuan Q, Luo KY, Zhang Y, Zhao CL (2023a) Morphological characteristics and phylogenetic analyses revealed three new wood-inhabiting fungi (Agaricomycetes, Basidiomycota) in Southern China. Phytotaxa 592: 179–195. https://doi.org/10.11646/phytotaxa.592.3.1

Yuan Y, Bian LS, Wu YD, Chen JJ, Wu F, Liu HG, Zeng GY, Dai YC (2023b) Species diversity of pathogenic wood-rotting fungi (Agaricomycetes, Basidiomycota) in China. Mycology 14: 204–226. https://doi.org/10.1080/21501203.2023.2238779

Yuan Q, Li YC, Dai YF, Wang KY, Wang YX, Zhao CL (2024) Morphological and molecular identification for four new wood-inhabiting species of *Lyomyces* (Basidiomycota) from China. MycoKeys 110: 67–92. https://doi.org/10.3897/mycokeys.110.133108

Zhang JL, Gu ZR, Zhou CQ, Zhou HM (2025) Molecular phylogeny and morphology reveal four new species in Hymenochaetales and one new species in Cantharellales from Southwestern China. MycoKeys 115: 87–135. https://doi.org/10.3897/mycokeys.115.142433

Zhao CL, Wu ZQ (2017) *Ceriporiopsis kunmingensis* sp. nov. (Polyporales, Basidiomycota) evidenced by morphological characters and phylogenetic analysis. Mycological Progress 16: 93–100. https://doi.org/10.1007/s11557-016-1259-8

Zhao CL, Qu MH, Huang RX, Karunarathna SC (2023a) Multi-gene phylogeny and taxonomy of the wood-rotting fungal genus *Phlebia* sensu lato (Polyporales, Basidiomycota). Journal of Fungi (Basel, Switzerland) 9: 320. https://doi.org/10.3390/jof9030320

Zhao H, Nie Y, Zong TK, Wang K, Wen SN, Lv ML, Cui YJ, Tohtirjap A, Chen JJ, Zhao CL, Wu F, Cui BK, Yuan Y, Dai YC, Liu XY (2023b) Species diversity, updated classification and divergence times of the phylum Mucoromycota. Fungal Diversity 123: 49–157. https://doi.org/10.1007/s13225-023-00525-4

Zhao H, Wu YD, Yang ZR, Liu HG, Wu F, Dai YC, Yuan Y (2024) Polypore funga and species diversity in tropical forest ecosystems of Africa, America and Asia, and a comparison with temperate and boreal regions of the Northern Hemisphere. Forest Ecosystems 11: 100200. https://doi.org/10.1016/j.fecs.2024.100200

Zhou HM, Dai YC, Bian LS, Liu HG, Vlasák J, Yuan Y (2025) Diversity, divergence time, and biogeography of the genus *Albatrellus* (Agaricomycetes, Russulales). Mycology, 738–776. https://doi.org/10.1080/21501203.2024.2386021