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## Morphology and multigene phylogeny revealed four new species of *Geastrum* (Geastrales, Basidiomycota) from China

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335-359

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# Morphology and multigene phylogeny revealed four new species of *Gastrum* (Gastrales, Basidiomycota) from China

Xin Yang<sup>1</sup>, Yonggao Zhu<sup>1</sup>, Songjing Duan<sup>1</sup>, Xingxing Wu<sup>2</sup>, Changlin Zhao<sup>1</sup>

1 College of Forestry, Southwest Forestry University, Kunming 650224, China

2 Yunnan Forestry Technological College, Kunming 650224, China

Corresponding author: Changlin Zhao (fungi@swfu.edu.cn, fungichanglinz@163.com)

## Abstract

In the present study, four new species, *Gastrum artocarpicola*, *G. fibulatum*, *G. sinense* and *G. trachelium* collected from China, are proposed based on a combination of morphological characteristics and molecular evidence. *Gastrum artocarpicola* is characterized by shallowly saccate to deep saccate exoperidium, bubble-shaped to flask shaped basidia, spherical basidiospores. *G. fibulatum* is characterized by shallowly saccate to deep saccate exoperidium, spherical basidiospores, generative hyphae with clamp connections in the mycelium layer. *G. sinense* has arched exoperidium, long stipe expanded basidiomata, and spherical basidiospores. *G. trachelium* has deep saccate exoperidium, flask-shaped basidia, and spherical basidiospores. Sequences of the internal transcribed spacers (ITS), large subunit (nrLSU), the largest subunit of ribosomal polymerase II (RPB1), and subunit 6 of ATP synthase (ATP6) of the nuclear ribosomal DNA (rDNA) markers of the studied samples were generated, and the phylogenetic analyses were performed with maximum likelihood, maximum parsimony and Bayesian inference methods. The results showed that our collection clustered within *Gastrum* but distinctly from the others. Full morphological descriptions, illustrations, and phylogenetic analyses results for the four new species are provided. In addition, *G. sanglinense* is treated as a synonym of *beijingense*.

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**Key words:** Classification, earthstar fungus, Geastraceae, molecular systematics, taxonomy

## Introduction

Fungi are among the most diverse groups of organisms on the earth and play an indispensable role in ecosystem processes and functioning (Wei and Dai 2004; Hyde 2022; Dong et al. 2024a). Currently, 19 phyla of fungi are accepted as Aphelidiomycota, Ascomycota, Basidiobolomycota, Basidiomycota, Blastocladiomycota, Calcarisporiellomycota, Chytridiomycota, Entomophthoromycota, Entorrhizomycota, Glomeromycota, Kickxellomycota, Monoblepharomycota, Mortierellomycota, Mucoromycota, Neocallimastigomycota, Olpidiomycota, Rozellomycota, Sanchytriomycota, and Zoopagomycota (Wijayawardene et al. 2024). Most Basidiomycota species act as decomposers and mutualists of plants and animals, and play fundamental ecological roles such as driving carbon cycling in forest soils, mediating the mineral nutrition of plants, and

alleviating carbon limitations of other soil organisms (Tedersoo et al. 2014; Yuan et al. 2023a). The genus *Gastrum* Pers., widely known as the earthstars, belongs to the family Geastraceae (Basidiomycota) (Sunhede 1989; Zamora et al. 2014). *Gastrum* species have been recorded on all continents except Antarctica, mostly in the forest humus layer, and occasionally on rotten wood, sand, or grassland, in which it is characterized by the exoperidium splitting into rays at maturity (León 1968).

*Gastrum* was first proposed by Persoon (1794) and typified by *G. coronatum* Pers. (Persoon 1794). The Gasteromycete means “stomach fungus” and these fungi produce their spores inside the fruiting body that are enclosed inside an outer covering called peridium, and the *Gastrum* is considered one of the most genera-diverse of gasteroid fungi (Chouhan and Panwar 2021; Zhang et al. 2023b). *Gastrum* is characterized by the stelliform basidiomata, exoperidium with three layers, sessile or stalked endoperidium, and sulcate, plicate, folded or fibrillose peristome, distinctly or indistinctly delimited, sometimes with mycosclereids (Sunhede 1989; Zamora et al. 2014). Both Index Fungorum (<http://www.indexfungorum.org>; 10 October 2024) and MycoBank database (<http://www.MycoBank.org>; 10 October 2024) have registered 388 specific and infraspecific names in *Gastrum*, but the actual number of species has been estimated to be around 130 (Finy et al. 2021; Wijayawardene et al. 2022; He et al. 2024). Of them, 36 species have been recorded from China (Wang and Bau 2023; Zhang et al. 2023b; Wu et al. 2024; Yang et al. 2024).

The classification studies on *Gastrum* were carried out in China, in which the morphological characteristics were as the core evidence for species taxonomy (Teng 1963; Tai 1979; Zhou et al. 2007). Phylogenetic relationships among European earthstars were inferred using sequence data from the nuclear ribosomal DNA internal transcribed region (ITS1, 5.8S, and ITS2), nuclear large subunit (nrLSU), and translation elongation factor 1-alpha (tef1- $\alpha$ ), and the results indicated that the phylogenetic analyses recovered 11 clades including thirty-one morphological species (Jeppson et al. 2013). They proposed a close phylogenetic relationship between *Myriostoma coliforme* (Dicks.) Corda and *Gastrum coronatum* (Jeppson et al. 2013). Later, on the basis of the phylogenetic analyses of four molecular markers as 5.8S nrDNA, nrLSU, the largest subunit of ribosomal polymerase II (RPB1), and subunit 6 of ATP synthase (ATP6), the research indicated that a new subdivision of the genus *Gastrum* was presented, and the phylogenetic and morphological boundaries among 3 genera *Gastrum*, *Myriostoma* Desv., and *Radiigera* Zeller (Geastraceae) were evaluated (Zamora et al. 2014). The genus *Myriostoma* represented a different phylogenetic lineage within the family Geastraceae which was confirmed as distinct from *Gastrum* (Zamora et al. 2014). Inferred from ITS, LSU nrDNA, RPB1, and ATP6, the phylogenetic reconstructions with 95 samples showed that 5 clades were considered as five subsections and a total of 27 lineages were proposed (Zamora et al. 2015). Multigene phylogenetic analyses involving sequences from ITS, nrLSU, RPB1, ATP6, and tef1- $\alpha$  revealed that *Gastrum* formed a distinct clade and had a close relationship with *G. granulosum* s.l. (Finy et al. 2021). Based on the morphological observation combined with phylogenetic analysis of ITS-nrLSU-RPB1-ATP6, the species *G. yanshanense* C.L. Hou et al. and *G. beijingense* C.L. Hou et al., were introduced from Yanshan Mountains, China

(Zhou et al. 2022). Based on morphological observation combined with phylogenetic analysis through ITS+nrLSU, 7 new *Gastrum* species were introduced from China (Wang and Bau 2023). In addition, phylogenetic analyses based on sequences of ITS, nrLSU, and ATP6 regions showed that *G. sanglinense* Y.Q. Wu & Shu R. Wang was sister to *G. yanshanense* and *G. rubellum* P.-A. Moreau & C. Lécuru (Wu et al. 2024). Phylogenetic analyses inferred from ITS+nrLSU dataset indicated that *G. yunnanense* Xin Yang & C.L. Zhao was nested within the *Gastrum*, which was forming a monophyletic lineage and then grouped with *G. velutinum* Morgan, and *G. javanicum* Lév. (Yang et al. 2024).

During investigations on earthstars fungi in China, we found four *Gastrum* species that could not be assigned to any described species. Based on the morphological characteristics and molecular phylogenetic analyses, these species are described as *G. artocarpicola*, *G. fibulatum*, *G. sinense* and *G. trachelium*.

## Materials and methods

### Sample collection and examination

The fresh basidiomata of fungi growing on the ground were collected from Guangzhou and Yangjiang in Guangdong Province, Nanjing in Jiangsu Province, Xiameng in Fujian Province, P.R. China. The samples were photographed using a Jianeng 80D camera (Tokyo, Japan) in situ and fresh macroscopic details, such as the color of the mycelial layer, the pseudoparenchymatous layer, the type of endoperideal body, and the shape of the peristome, were recorded. All the photographs were focus-stacked and merged using Helicon Focus Pro 7.7.5 software. Specimens were dried in an electric food dehydrator at 40 °C (Zhao et al. 2023; Dong et al. 2024a, b), and then sealed and stored in an envelope bag and deposited in the herbarium of the Southwest Forestry University (SWFC), Kunming, Yunnan Province, P.R. China.

### Morphology

The macromorphological descriptions were based on field notes and photos captured in the field and laboratory and followed the color terminology of Petersen (1996). Micromorphological data were obtained from the dried specimens following observation under a light microscope (Zhao and Wu 2017; Wang et al. 2023, 2024; Dong et al. 2024a). Drawings were made with the aid of a fungus plotter (Zhao et al. 2023). The measurements and drawings were made from slide preparations stained with Cotton Blue (0.1 mg aniline blue dissolved in 60 g pure lactic acid), Melzer's reagent (3 g potassium iodide, 1 g crystalline iodine, 44 g chloral hydrate, and 40 ml distilled water) and 5% potassium hydroxide. Spore size data, excluded 5% of the measurements from each end of the range, showing them in parentheses. The following abbreviations were used: KOH = 5% potassium hydroxide water solution, CB+ = cyanophilous, CB = cotton clue, CB- = acyanophilous, IKI = Melzer's reagent, IKI- = both inamyloid and index-trinoid. Q = variation in the L/W ratios between the specimens studied and n = a/b (number of spores (a) measured from a given number (b) of specimens).  $Q_m$  represented the average Q of basidiospores measured ± standard deviation.

## DNA extraction, PCR amplification, sequencing, and phylogenetic analyses

The CTAB rapid plant genome extraction kit-DN14 (Aidlab Biotechnologies Co., Ltd, Beijing, China) was used to obtain genomic DNA from the dried fungal specimens according to the manufacturer's instructions (Zhao and Wu 2017; Zhou et al. 2023; Dong et al. 2024a). Four molecular markers were investigated, i.e., ITS (including 5.8S) and 28S (nrLSU) nrDNA, the largest subunit of RNA polymerase II (RPB1), and subunit 6 of ATP synthase (ATP6). The nuclear ribosomal ITS region was amplified with ITS1F and ITS4 primer pair (White et al. 1990). The nuclear nrLSU region was amplified with the LR0R and LR7 primer pair (Vilgalys and Hester 1990; Rehner and Samuels 1994). The RPB1 region was initially amplified with RPB1-Af and RPB1-Cr (Matheny et al. 2002). For amplifying ATP6, ATP6-1 and ATP6-2 were used (Kretzer and Bruns 1999). The PCR procedure for ITS was as follows: initial denaturation at 94 °C for 3 min, followed by 35 cycles at 94 °C for 60 s, 56 °C for 60 s, and 72 °C for 1 min, and a final extension of 72 °C for 10 min (Yang et al. 2024). The PCR procedure for nrLSU was as follows: initial denaturation at 94 °C for 1 min, followed by 35 cycles at 94 °C for 30 s, 48 °C for 1 min, and 72 °C for 1.5 min, and a final extension of 72 °C for 10 min (Yang et al. 2024). The PCR procedure for RPB1 was 94 °C for 2 min, followed by 10 cycles at 94 °C for 40 s, 60 °C for 40 s and 72 °C for 2 min, then followed by 37 cycles at 94 °C for 45 s, 55 °C for 1.5 min and 72 °C for 2 min, and a final extension of 72 °C for 10 min (Dong et al. 2024a). The PCR procedure for ATP6 was as follows: initial denaturation at 94 °C for 3 min, followed by 35 cycles at 94 °C for 35 s, 55 °C for 50 s, and 72 °C for 45 min, and a final extension of 72 °C for 10 min (Kretzer and Bruns 1999). The PCR products were purified and sequenced at Kunming Tsingke Biological Technology Limited Company (Yunnan Province, China). All of the newly generated sequences were deposited in NCBI GenBank (<https://www.ncbi.nlm.nih.gov/genbank/>) (Table 1).

Sequences generated for this study were aligned, with additional sequences downloaded from GenBank. Sequences were aligned in MAFFT 7 (<https://mafft.cbrc.jp/alignment/server/>) adjusting the direction of nucleotide sequences according to the first sequence (accurate enough for most cases), and selecting the G-INS-i iterative refinement method (Katoh et al. 2019). The alignment was adjusted manually using AliView version 1.27 (Larsson 2014). The dataset was aligned first, and then the sequences of ITS+nrLSU+RPB1+ATP6 were combined with Mesquite version 3.51. The combined ITS+nrLSU+RPB1+ATP6 sequences were used to infer the position of four new species in the genus *Gastrum* and related species. Sequences of *Schenella pityophila* (Malençon & Riousset) Estrada & Lado retrieved from GenBank were used as an outgroup taxon (Zamora 2015; Wang and Bau 2023).

Maximum Parsimony (MP), Maximum Likelihood (ML) and Bayesian Inference (BI) analyses were applied to the combined three datasets following a previous study (Zhao and Wu 2017). MP analysis was performed in PAUP\* version 4.0b10 (Swofford 2002). All characters were equally weighted and gaps were treated as missing data. Trees were inferred using the heuristic search option with TBR branch swapping and 1,000 random sequence additions. Maxtrees were set to 5,000, branches of zero length were collapsed and all parsimonious trees were saved. Clade robustness was assessed using bootstrap (BT) analysis with 1,000

**Table 1.** List of species, specimens, and GenBank accession number of sequences used in this study. [New species is shown in bold; \* type material].

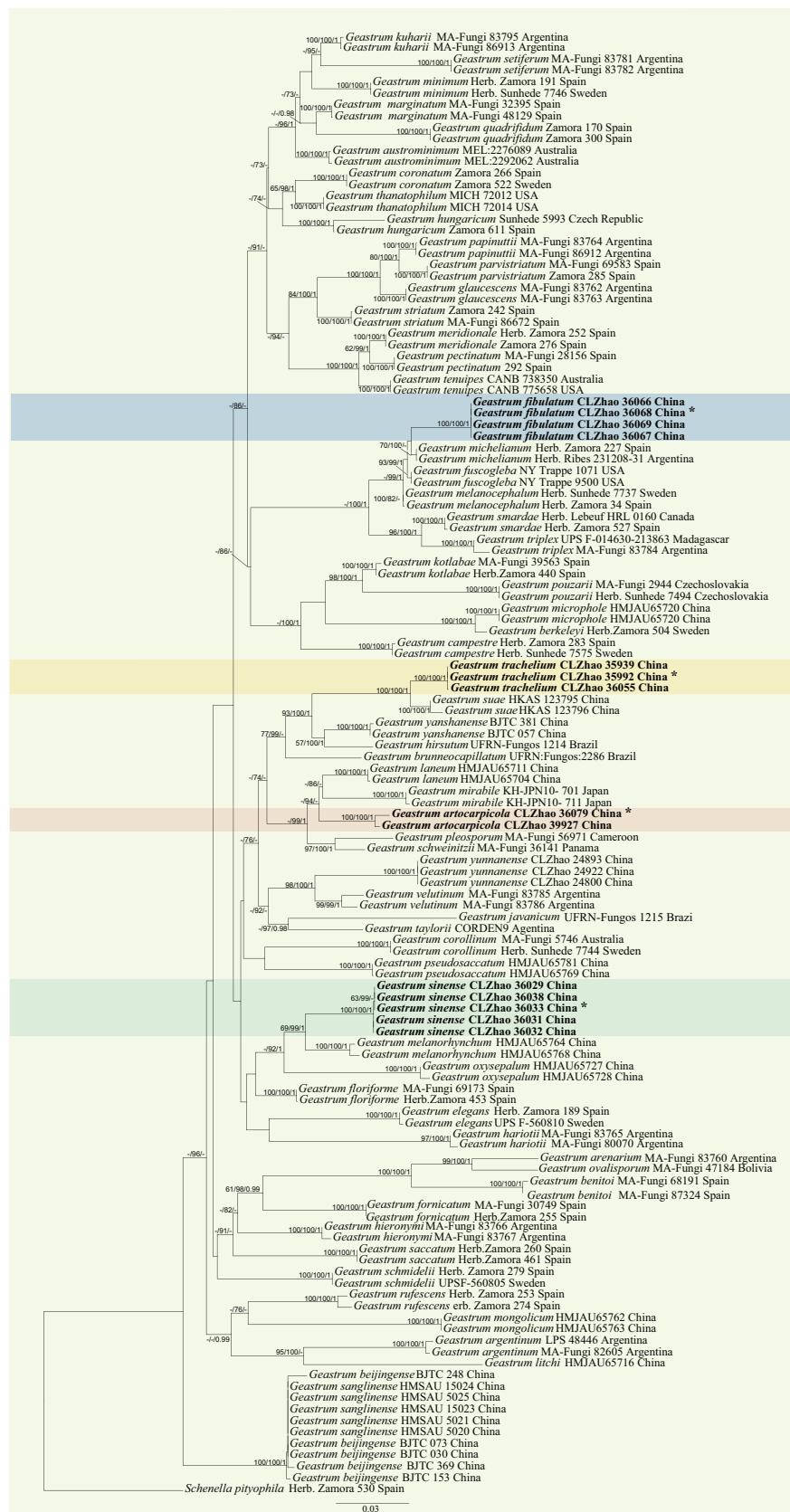
Species Name	Locality	Sample No.	GenBank Accession No.				References
			ITS	nrLSU	rpb1	atp6	
<i>Gastrum arenarium</i>	Argentina	MA-Fungi 83760	KF988351	KF988471	KF988606	KF988741	Zamora et al. (2014)
<i>Gastrum argentinum</i>	Argentina	LPS 48446	KF988352	KF988472	KF988607	KF988742	Zamora et al. (2014)
<i>Gastrum argentinum</i>	Argentina	MA-Fungi 82605	KF988353	KF988473	–	KF988743	Zamora et al. (2014)
<b><i>Gastrum artocarpicola</i></b>	<b>China</b>	<b>CLZhao 36079*</b>	<b>PQ484149</b>	<b>PQ481917</b>		<b>PQ634876</b>	<b>Present study</b>
<b><i>Gastrum artocarpicola</i></b>	<b>China</b>	<b>CLZhao 39927</b>	<b>PQ484150</b>	<b>PQ481918</b>		<b>PQ634877</b>	<b>Present study</b>
<i>Gastrum austrominimum</i>	Australia	MEL:2276089	KP687490	KP687451	KP687532	KP687573	Zamora et al. (2015)
<i>Gastrum austrominimum</i>	Australia	MEL:2292062	KP687491	KP687452	KP687533	KP687574	Zamora et al. (2015)
<i>Gastrum beijingense</i>	China	BJTC 248	MZ508872	MZ509376	MZ571167	MZ571178	Zhou et al. (2022)
<i>Gastrum beijingense</i>	China	BJTC 073	MZ508873	MZ509377	MZ571168	MZ571179	Zhou et al. (2022)
<i>Gastrum beijingense</i>	China	BJTC 369	–	MZ509375	MZ571166	MZ571177	Zhou et al. (2022)
<i>Gastrum beijingense</i>	China	BJTC 030	MZ508874	MZ509378	MZ571169	MZ571180	Zhou et al. (2022)
<i>Gastrum beijingense</i>	China	BJTC 1523	MZ508875	MZ509379	MZ571170	MZ571181	Zhou et al. (2022)
<i>Gastrum benitoi</i>	Spain	MA-Fungi 68191	KF988350	KF988469	KF988604	KF988739	Zamora et al. (2015)
<i>Gastrum benitoi</i>	Spain	MA-Fungi 87324	KP687494	KP687455	KP687536	KP687577	Zamora et al. (2015)
<i>Gastrum berkeleyi</i>	Sweden	Herb.Zamora 504	KF988356	KF988476	KF988611	KF988746	Zamora et al. (2015)
<i>Gastrum brunneocapillatum</i>	Brazil	UFRN:Fungos:2286	MH634996	MH635029	–	–	Accioly et al. (2019)
<i>Gastrum campestre</i>	Spain	Herb. Zamora 283	JN943167	JN939575	JN991286	KF988748	Zamora et al. (2014)
<i>Gastrum campestre</i>	Sweden	Herb. Sunhede 7575	KF988357	KF988479	KF988614	KF988749	Zamora et al. (2014)
<i>Gastrum corollinum</i>	Australia	MA-Fungi 5746	KF988359	KF988481	KF988616	KF988751	Zamora et al. (2014)
<i>Gastrum corollinum</i>	Sweden	Herb. Sunhede 7744	KF988360	KF988482	KF988617	KF988752	Zamora et al. (2014)
<i>Gastrum coronatum</i>	Spain	Zamora 266	KF988361	KF988483	KF988618	KF988753	Zamora et al. (2014)
<i>Gastrum coronatum</i>	Sweden	Zamora 522	KF988362	KF988484	KF988619	KF988754	Zamora et al. (2014)
<i>Gastrum elegans</i>	Spain	Herb. Zamora 189	KF988366	KF988488	KF988623	KF988758	Zamora et al. (2014)
<i>Gastrum elegans</i>	Sweden	UPS F-560810	KF988367	KF988489	KF988624	KF988759	Zamora et al. (2014)
<b><i>Gastrum fibulatum</i></b>	<b>China</b>	<b>CLZhao 36066</b>	<b>PQ484151</b>	<b>PQ481919</b>	<b>PQ811828</b>	–	<b>Present study</b>
<b><i>Gastrum fibulatum</i></b>	<b>China</b>	<b>CLZhao 36067</b>	<b>PQ484152</b>	<b>PQ481920</b>	<b>PQ811829</b>	–	<b>Present study</b>
<b><i>Gastrum fibulatum</i></b>	<b>China</b>	<b>CLZhao 36068*</b>	<b>PQ484153</b>	<b>PQ481921</b>	<b>PQ811830</b>	–	<b>Present study</b>
<b><i>Gastrum fibulatum</i></b>	<b>China</b>	<b>CLZhao 36069</b>	<b>PQ484154</b>	<b>PQ481922</b>	<b>PQ811831</b>	–	<b>Present study</b>
<i>Gastrum floriforme</i>	Spain	MA-Fungi 69173	KF988372	KF988494	KF988629	KF988764	Zamora et al. (2014)
<i>Gastrum floriforme</i>	Spain	Herb.Zamora 453	KF988373	KF988495	KF988630	KF988765	Zamora et al. (2014)
<i>Gastrum fornicatum</i>	Spain	MA-Fungi 30749	KF988375	KF988497	KF988632	KF988767	Zamora et al. (2014)
<i>Gastrum fornicatum</i>	Spain	Herb.Zamora 255	KF988374	KF988496	KF988631	KF988766	Zamora et al. (2014)
<i>Gastrum fuscogleba</i>	USA	NY Trappe 1071	KF988376	KF988498	KF988633	KF988768	Zamora et al. (2014)
<i>Gastrum fuscogleba</i>	USA	NY Trappe 9500	KF988377	KF988499	KF988634	KF988769	Zamora et al. (2014)
<i>Gastrum glaucescens</i>	Argentina	MA-Fungi 83762	KF988378	KF988500	KF988635	KF988770	Zamora et al. (2014)
<i>Gastrum glaucescens</i>	Argentina	MA-Fungi 83763	KF988379	KF988501	KF988636	KF988771	Zamora et al. (2014)
<i>Gastrum hariotii</i>	Argentina	MA-Fungi 83765	KF988381	KF988504	KF988639	KF988774	Zamora et al. (2014)
<i>Gastrum hariotii</i>	Dominican	MA-Fungi 80070	–	KF988503	KF988638	KF988773	Zamora et al. (2014)
<i>Gastrum hieronymi</i>	Argentina	MA-Fungi 83766	KF988384	KF988508	KF988643	KF988776	Zamora et al. (2014)
<i>Gastrum hieronymi</i>	Argentina	MA-Fungi 83767	KF988344	KF988509	KF988644	KF988777	Zamora et al. (2014)
<i>Gastrum hirsutum</i>	Brazil	UFRN-Fungos 1214	KJ127029	JQ683662	-	JQ683670	Accioly et al. (2019)
<i>Gastrum hungaricum</i>	Czech	Sunhede 5993	KP687500	KP687461	KP687542	KP687582	Zamora et al. (2015)
<i>Gastrum hungaricum</i>	Spain	Zamora 611	KP687501	KP687462	KP687543	KP687583	Zamora et al. (2015)
<i>Gastrum javanicum</i>	Brazil	UFRN-Fungos 1215	KJ127031	–	–	KJ127016	Cabral et al. (2017)
<i>Gastrum kotlabae</i>	Spain	MA-Fungi 39563	KF988385	KF988510	KF988645	KF988778	Zamora et al. (2014)
<i>Gastrum kotlabae</i>	Spain	Herb.Zamora 440	KF988386	KF988511	KF988646	KF988779	Zamora et al. (2014)
<i>Gastrum kuharii</i>	Argentina	MA-Fungi 83795	KF988463	KF988598	KF988733	KF988864	Zamora et al. (2014)
<i>Gastrum kuharii</i>	Argentina	MA:Fungi 86913	KP687502	KP687463	KP687544	KP687584	Zamora et al. (2014)
<i>Gastrum laneum</i>	China	HMJAU65711	OP964640	OP964638	–	–	Wang and Bau (2023)
<i>Gastrum laneum</i>	China	HMJAU65704	OP964641	OP964639	–	–	Wang and Bau (2023)

Species Name	Locality	Sample No.	GenBank Accession No.				References
			ITS	nrLSU	rpb1	atp6	
<i>Gastrum litchi</i>	China	HMJAU65716	QO360756	OP964619	—	—	Wang and Bau (2023)
<i>Gastrum marginatum</i>	Spain	MA-Fungi 32395	KP687505	KP687466	KP687547	KP687587	Zamora et al. (2015)
<i>Gastrum marginatum</i>	Spain	MA-Fungi 48129	KP687506	KP687467	KP687548	KP687588	Zamora et al. (2015)
<i>Gastrum melanocephalum</i>	Sweden	Herb. Sunhede 7737	KF988396	KF988523	KF988658	KF988790	Zamora et al. (2014)
<i>Gastrum melanocephalum</i>	Spain	Herb. Zamora 34	KF988395	KF988522	KF988657	KF988789	Zamora et al. (2014)
<i>Gastrum melanorhynchum</i>	China	HMJAU65764	OP964617	OP964614	—	—	Wang and Bau (2023)
<i>Gastrum melanorhynchum</i>	China	HMJAU65768	OP964618	OP964615	—	—	Wang and Bau (2023)
<i>Gastrum meridionale</i>	Spain	Herb. Zamora 252	KF988412	KF988540	KF988675	KF988808	Zamora et al. (2014)
<i>Gastrum meridionale</i>	Spain	Zamora 276	KP687513	KP687475	KP687556	KP687595	Zamora et al. (2015)
<i>Gastrum michelianum</i>	Spain	Herb. Zamora 227	KF988398	KF988525	KF988660	KF988792	Zamora et al. (2014)
<i>Gastrum michelianum</i>	Argentina	Herb. Ribes 231208-31	—	—	KF988661	KF988793	Zamora et al. (2014)
<i>Gastrum microphole</i>	China	HMJAU65720	OP964636	OP964643	—	—	Wang and Bau (2023)
<i>Gastrum microphole</i>	China	HMJAU65721	OP964637	OP964644	—	—	Wang and Bau (2023)
<i>Gastrum minimum</i>	Spain	Herb. Zamora 191	KF988400	KF988528	KF988663	KF988795	Wang and Bau (2023)
<i>Gastrum minimum</i>	Sweden	Herb. Sunhede 7746	KF988401	KF988529	KF988664	KF988796	Zamora et al. (2014)
<i>Gastrum mirabile</i>	Japan	KH-JPN10- 701	JN845107	JN845225	—	JN845349	Wu et al. (2024)
<i>Gastrum mirabile</i>	Japan	KH-JPN10- 711	JN845108	JN845226	—	JN845350	Wu et al. (2024)
<i>Gastrum mongolicum</i>	China	HMJAU65762	OP964647	OP964645	—	—	Wang and Bau (2023)
<i>Gastrum mongolicum</i>	China	HMJAU65763	OP964648	OP964646	—	—	Wang and Bau (2023)
<i>Gastrum ovalisporum</i>	Bolivia	MA-Fungi 47184	KF988411	KF988539	KF988674	KF988805	Zamora et al. (2015)
<i>Gastrum oxysepalum</i>	China	HMJAU65727	OP964632	OP964622	—	—	Wang and Bau (2023)
<i>Gastrum oxysepalum</i>	China	HMJAU65728	OP964633	OP964623	—	—	Wang and Bau (2023)
<i>Gastrum papinuttii</i>	Argentina	MA-Fungi 83764	KF988380	KF988502	KF988637	KF988772	Zamora et al. (2015)
<i>Gastrum papinuttii</i>	Argentina	MA-Fungi 86912	KP687515	KP687477	KP687558	KP687596	Zamora et al. (2015)
<i>Gastrum parvistriatum</i>	Spain	MA-Fungi 69583	JN943160	JN939560	JN991291	KF988806	Zamora et al. (2014)
<i>Gastrum parvistriatum</i>	Spain	Zamora 285	JN943161	JN939571	JN991282	KP687597	Zamora et al. (2015)
<i>Gastrum pectinatum</i>	Spain	MA-Fungi 28156	KP687516	KP687478	KP687559	KP687598	Zamora et al. (2015)
<i>Gastrum pectinatum</i>	Spain	Zamora 292	KP687521	KP687483	KP687564	KP687601	Zamora et al. (2015)
<i>Gastrum pleosporum</i>	Cameroon	MA-Fungi 56971	KF988416	KF988544	KF988679	KF988811	Zamora et al. (2014)
<i>Gastrum pouzarii</i>	Czechoslovakia	MA-Fungi 2944	KF988417	KF988545	KF988680	KF988812	Zamora et al. (2014)
<i>Gastrum pouzarii</i>	Czechoslovakia	Herb. Sunhede 7494	KF988418	KF988546	KF988681	KF988813	Zamora et al. (2014)
<i>Gastrum pseudosaccatum</i>	China	HMJAU65781	OP964625	OP964635	—	—	Wang and Bau (2023)
<i>Gastrum pseudosaccatum</i>	China	HMJAU65769	OP964628	OP964634	—	—	Wang and Bau (2023)
<i>Gastrum quadrifidum</i>	Spain	Zamora 170	KF988421	KF988549	KF988684	KF988816	Zamora et al. (2014)
<i>Gastrum quadrifidum</i>	Spain	Zamora 300	KP687524	KP687486	KP687567	KP687604	Zamora et al. (2015)
<i>Gastrum rufescens</i>	Spain	Herb. Zamora 253	KF988424	KF988552	KF988687	KF988819	Zamora et al. (2014)
<i>Gastrum rufescens</i>	Spain	Herb. Zamora 274	KF988425	KF988553	KF988688	KF988820	Zamora et al. (2014)
<i>Gastrum saccatum</i>	Spain	Herb.Zamora 260	KF988430	KF988560	KF988695	KF988827	Zamora et al. (2014)
<i>Gastrum saccatum</i>	Spain	Herb.Zamora 461	KF988431	KF988561	KF988696	KF988828	Zamora et al. (2014)
<i>Gastrum sanglinense</i>	China	HMSAU 15023	OP050118	OP050163	—	OP056323	Wu et al. (2024)
<i>Gastrum sanglinense</i>	China	HMSAU 15024	OP050119	OP050164	—	OP056324	Wu et al. (2024)
<i>Gastrum sanglinense</i>	China	HMSAU 15020	OP050116	OP050161	—	—	Wu et al. (2024)
<i>Gastrum sanglinense</i>	China	HMSAU 15021	OP050117	OP050162	—	—	Wu et al. (2024)
<i>Gastrum sanglinense</i>	China	HMSAU 15025	OP050120	OP050165	—	—	Wu et al. (2024)
<i>Gastrum schmidelii</i>	Spain	Herb. Zamora 279	KF988434	KF988564	KF988699	KF988831	Zamora et al. (2014)
<i>Gastrum schmidelii</i>	Sweden	UPSF-560805	KF988435	KF988565	KF988700	KF988832	Zamora et al. (2014)
<i>Gastrum schweinitzii</i>	Panama	MA-Fungi 36141	KF988438	KF988568	KF988703	KF988835	Zamora et al. (2014)
<i>Gastrum setiferum</i>	Argentina	MA-Fungi 83781	—	KF988571	KF988706	KF988837	Zamora et al. (2014)
<i>Gastrum setiferum</i>	Argentina	MA-Fungi 83782	—	KF988572	KF988707	KF988838	Zamora et al. (2014)
<i>Gastrum sinense</i>	China	CLZhao 36029	PQ484155	PQ481929	PQ822016	—	Present study
<i>Gastrum sinense</i>	China	CLZhao 36031	PQ484156	PQ481930	PQ822017	PQ645133	Present study
<i>Gastrum sinense</i>	China	CLZhao 36032	PQ484157	PQ481923	PQ822018	—	Present study
<i>Gastrum sinense</i>	China	CLZhao 36033*	PQ484158	PQ481924	PQ822019	—	Present study
<i>Gastrum sinense</i>	China	CLZhao 36038	PQ484159	PQ481925	PQ822020	PQ645134	Present study

Species Name	Locality	Sample No.	GenBank Accession No.				References
			ITS	nrLSU	rpb1	atp6	
<i>Geastrum smardae</i>	Canada	Herb. Lebeuf HRL 0160	KF988440	KF988573	KF988708	KF988839	Zamora et al. (2014)
<i>Geastrum smardae</i>	Spain	Herb. Zamora 527	KF988441	KF988574	KF988709	KF988840	Zamora et al. (2014)
<i>Geastrum striatum</i>	Spain	Zamora 242	JN943163	JN939559	JN991290	KP687606	Zamora et al. (2015)
<i>Geastrum striatum</i>	Spain	MA-Fungi 86672	KF988443	KF988577	KF988712	KF988843	Zamora et al. (2015)
<i>Geastrum suae</i>	China	HKAS 123795	ON529511	ON529515	–	–	Zhang et al. (2022)
<i>Geastrum suae</i>	China	HKAS 123796	ON529514	ON529518	–	–	Zhang et al. (2022)
<i>Geastrum taylorii</i>	Argentina	CORDEN9	JN845204	JN845329	–	JN845432	Kasuya et al. (2012)
<i>Geastrum tenuipes</i>	Australia	CANB 738350	KP687526	KP687488	KP687570	KP687609	Zamora et al. (2015)
<i>Geastrum tenuipes</i>	USA	CANB 775658	KP687527	KP687489	KP687571	KP687610	Zamora et al. (2014)
<i>Geastrum thanatophilum</i>	USA	MICH 72012	KF988364	KF988486	KF988621	KF988756	Zamora et al. (2014)
<i>Geastrum thanatophilum</i>	USA	MICH 72014	KF988365	KF988487	KF988622	KF988757	Zamora et al. (2014)
<i>Geastrum trachelium</i>	China	CLZhao 35939	PQ484160	PQ481926	PQ783782	–	Present study
<i>Geastrum trachelium</i>	China	CLZhao 36055	PQ484162	PQ481928	PQ783784	–	Present study
<i>Geastrum trachelium</i>	China	CLZhao 35992*	PQ484161	PQ481927	PQ783783	–	Present study
<i>Geastrum triplex</i>	Madagascar	UPS F-014630-213863	KF988444	KF988578	KF988713	KF988844	Zamora et al. (2014)
<i>Geastrum triplex</i>	Argentina	MA-Fungi 83784	KF988445	KF988579	KF988714	KF988845	Zamora et al. (2014)
<i>Geastrum velutinum</i>	Argentina	MA-Fungi 83785	KF988446	KF988581	KF988716	KF988847	Zamora et al. (2014)
<i>Geastrum velutinum</i>	Argentina	MA-Fungi 83786	KF988447	KF988582	KF988717	KF988848	Zamora et al. (2014)
<i>Geastrum yanshanense</i>	China	BJTC 381	MZ508878	MZ509383	MZ571175	MZ571184	Zhou et al. (2022)
<i>Geastrum yanshanense</i>	China	BJTC 057	MZ508879	MZ509384	MZ571176	MZ571185	Zhou et al. (2022)
<i>Geastrum yunnanense</i>	China	CLZhao 24800	PP511307	–	–	–	Yang et al. (2024)
<i>Geastrum yunnanense</i>	China	CLZhao 24893	PP511308	PP511310	–	–	Yang et al. (2024)
<i>Geastrum yunnanense</i>	China	CLZhao 24922	PP511309	PP511311	–	–	Yang et al. (2024)
<i>Schenella pityophila</i>	Spain	Herb. Zamora 530	KF988346	KF988464	KF988599	KF988734	Zamora et al. (2014)

pseudo-replicates (Felsenstein 1985). Descriptive tree statistics - tree length (TL), composite consistency index (CI), composite retention index (RI), composite rescaled consistency index (RC) and composite homoplasy index (HI) were calculated for each maximum parsimonious tree generated. Maximum likelihood (ML) analysis was performed using the CIPRES Science Gateway (Miller et al. 2012) based on the dataset using the RAxML -HPC BlackBox tool, with setting RAxML -HPC BlackBox halt bootstrapping automatically and 0.25 for maximum hours and obtaining the best tree using ML search. Other parameters in ML analysis used default settings, and statistical support values were obtained using nonparametric bootstrapping with 1, 000 replicates.

The best-evolutionary model of each alignment was estimated using jModelTest (Guindon and Gascuel 2003; Posada 2008) under the Akaike information criterion. MrModeltest 2.3 (Nylander 2004) was used to determine the best-fit evolution model for the dataset for Bayesian Inference (BI). Bayesian Inference was performed with MrBayes 3.1.2 with a general time reversible (GTR+I+G) model of DNA substitution and a gamma distribution rate variation across sites (Ronquist et al. 2012). Four Markov chains were run for two runs from random starting trees for 12 million generations combined ITS+nrLSU+rPB1+ATP6 sequences (Fig. 1), with trees and parameters sampled every 100<sup>th</sup> generations. The first quarter of all the generations were discarded as burn-ins. A majority rule consensus tree of all remaining trees and posterior probabilities were calculated. Branches were considered significantly supported if they received a maximum likelihood bootstrap value (BS) of  $\geq 70\%$ , a maximum parsimony bootstrap value (BT) of  $\geq 50\%$ , or Bayesian posterior probabilities (BPP) of  $\geq 0.95$ . The sequence alignments were deposited in figshare (DOI: 10.6084/m9.figshare.28130300).



**Figure 1.** Maximum likelihood strict consensus tree illustrating the phylogeny of four new species and related species in *Gastrum* based on ITS+nrLSU+RPB1+ATP6 sequences. Branches are labeled with maximum likelihood bootstrap values  $\geq 70\%$ , parsimony bootstrap values  $> 50\%$  and Bayesian posterior probabilities  $\geq 0.95$ , respectively. New species accessions are in bold, \* type material

## Results

### Phylogenetic analyses

The combined ITS+nrLSU+RPB1+ATP6 dataset (Fig. 1) included sequences from 128 fungal specimens representing 62 species. The dataset had an aligned length of 3,511 characters, of which 2,389 characters were constant, 193 were variable and parsimony-uninformative and 929 were parsimony-informative. Maximum parsimony analysis yielded one equally parsimonious tree (TL = 5,572, CI = 0.3087, HI = 0.6913, RI = 0.6999 and RC = 0.2161). The best model for the ITS dataset estimated and applied in the Bayesian analysis was GTR+I+G (lset nst = 6; rates = invgamma; prset statefreqpr = dirichlet (1, 1, 1, 1)). The bayesian and ML analyses resulted in similar topology to that of the MP analysis with an average standard deviation of split frequencies = 0.009556 (BI), and the effective sample size (ESS) across the two runs is double the average ESS (avg. ESS) = 2491.5. Branches that received bootstrap support for ML and BI  $\geq$  70%, and 0.95 were considered significantly supported, respectively. The results of BLAST queries in NCBI, based on ITS+nrLSU+RPB1+ATP6 separately, showed the sequences producing significant alignment descriptions.

In ITS BLAST results of *Gastrum artocarpicola*, the top ten taxa were *G. schweinitzii* (Berk. & M.A. Curtis) Zeller, *G. laneum* T. Bau & Xin Wang, *G. pusillipilosum* J.O. Sousa, Alfredo, R.J. Ferreira, M.P. Martín & Baseia, and *G. baculicrystallinum* J.O. Sousa, Accioly, Baseia & M.P. Martín (Maximum record descriptions: Max score 850; Total score 850; Query cover 91%; E value 0; Ident 94.28%). In nrLSU BLAST results, the top ten taxa were *Gastrum flexuosum* (L.S. Domínguez & Castellano) Jeppson & E. Larss., *G. yanshanense*, *G. striatum* DC., *G. lageniforme* Vittad., *G. schweinitzii*, *G. xerophilum* Long ex Desjardin, *G. velutinum* Morgan, and *G. saccatum* Fr. (Maximum record descriptions: Max score 1495; Total score 1495; Query cover 98%; E value 0.0; Ident 98.81%). In RPB1 BLAST results, the top ten taxa were *Gastrum schweinitzii*, *G. pleosporum* Douanla-Meli, *G. yanshanense*, and *G. elegans* Vittad. (Maximum record descriptions: Max score 1256; Total score 1256; Query cover 87%; E value 0.0; Ident 94.82%). In ATP6 BLAST results, the top ten taxa were *Gastrum mirabile* Mont., *G. schweinitzii*, and *G. pleosporum* (Maximum record descriptions: Max score 990; Total score 990; Query cover 96%; E value 0.0; Ident 91.73%).

In ITS BLAST results of *Gastrum fibulatum*, the top ten taxa were *G. triplex* Jungh. (Maximum record descriptions: Max score 957; Total score 957; Query cover 92%; E value 0; Ident 96.43%). In nrLSU BLAST results, the top ten taxa were *Gastrum melanocephalum* (Czern.) V.J. Staněk, *G. triplex*, *G. floriforme* Vittad., *G. smardae* V.J. Staněk, *G. flexuosum*, and *G. saccatum* (Maximum record descriptions: Max score 2501; Total score 2501; Query cover 99%; E value 0.0; Ident 99.35%). In RPB1 BLAST results, the top ten taxa were *Gastrum michelianum* (Sacc.) W.G. Sm., *G. melanocephalum*, *G. fuscoglebum* (Zeller) Jeppson & E. Larss., *G. smardae*, and *G. triplex* (Maximum record descriptions: Max score 1982; Total score 1982; Query cover 85%; E value 0.0; Ident 98.74%).

In ITS BLAST results of *Gastrum sinense*, the top ten taxa were *G. saccatum*, *G. melanorhynchum* T. Bau & Xin Wang, *G. fimbriatum* Fr., and *G. lageniforme* Vittad. (Maximum record descriptions: Max score 791; Total score 791; Query cover 96%; E value 0; Ident 90.72%). In nrLSU BLAST results, the top ten taxa were *Gastrum floriforme*, *G. xerophilum*, *G. saccatum*, *G. fimbriatum*, *G. schmidelii*, *G. striatum*, and *G. gorgonicum* (Maximum record descriptions: Max score 2425; Total score 2425;

Query cover 98%; E value 0.0; Ident 98.48%). In RPB1 BLAST results, the top ten taxa were *Gastrum saccatum*, *G. lageniforme*, *G. corollinum* (Batsch) Hollós, and *G. flexuosum* (Maximum record descriptions: Max score 1487; Total score 1487; Query cover 85%; E value 0.0; Ident 90.30%). In ATP6 BLAST results, the top ten taxa were *Gastrum lageniforme*, and *G. fimbriatum* (Maximum record descriptions: Max score 1035; Total score 1035; Query cover 94%; E value 0.0; Ident 93.30%).

In ITS BLAST results of *Gastrum trachelium*, the top ten taxa were *G. suae* Z.Q. Zhang, C.H. Li & Z.L. Luo, *G. rubellum* P.-A. Moreau & Lécuru, and *G. triplex* (Maximum record descriptions: Max score 946; Total score 946; Query cover 94%; E value 0; Ident 95.19%). In nrLSU BLAST results, the top ten taxa were *Gastrum yanshanense*, *G. floriforme*, *G. schweinitzii*, and *G. saccatum* (Maximum record descriptions: Max score 2431; Total score 2431; Query cover 99%; E value 0.0; Ident 98.55%). In RPB1 BLAST results, the top ten taxa were *Gastrum yanshanense*, *G. schweinitzii*, and *G. velutinum* (Maximum record descriptions: Max score 1230; Total score 1230; Query cover 97%; E value 0.0; Ident 92.56%).

The topology based on ITS+nrLSU+RPB1+ATP6 sequences, showed that all of the four new taxa were clustered into the genus *Gastrum*. *G. fibulatum* was sister to *G. michelianum*, *G. sinense* was closely related to *G. melanorhynchum*, *G. trachelium* was sister to *G. suae*, and *G. artocarpicola* formed a monophyletic lineage.

## Taxonomy

### *Gastrum artocarpicola* X. Yang & C.L. Zhao, sp. nov.

Mycobank No: 856264

Figs 2, 3

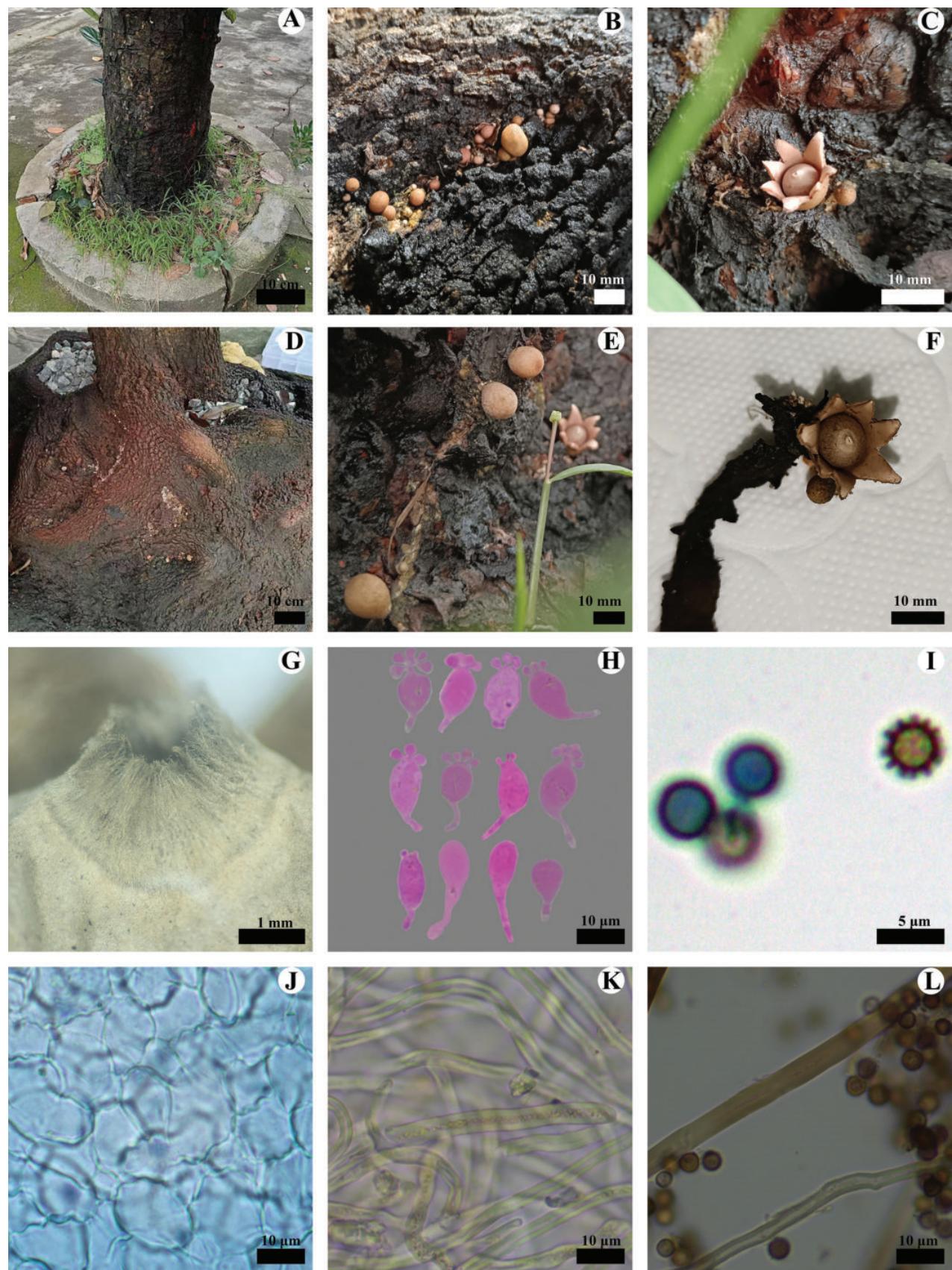
**Holotype.** CHINA • Guangdong Province, Yangjiang, Jiangcheng District, Bitian Lake Park, 21°52'N, 111°58'E, elev. 27 m, on the living tree of *Artocarpus*, 7 June 2024, CLZhao 36079 (SWFC).

**Etymology.** *Artocarpicola* (Lat.): referring to the species growing on *Artocarpus*.

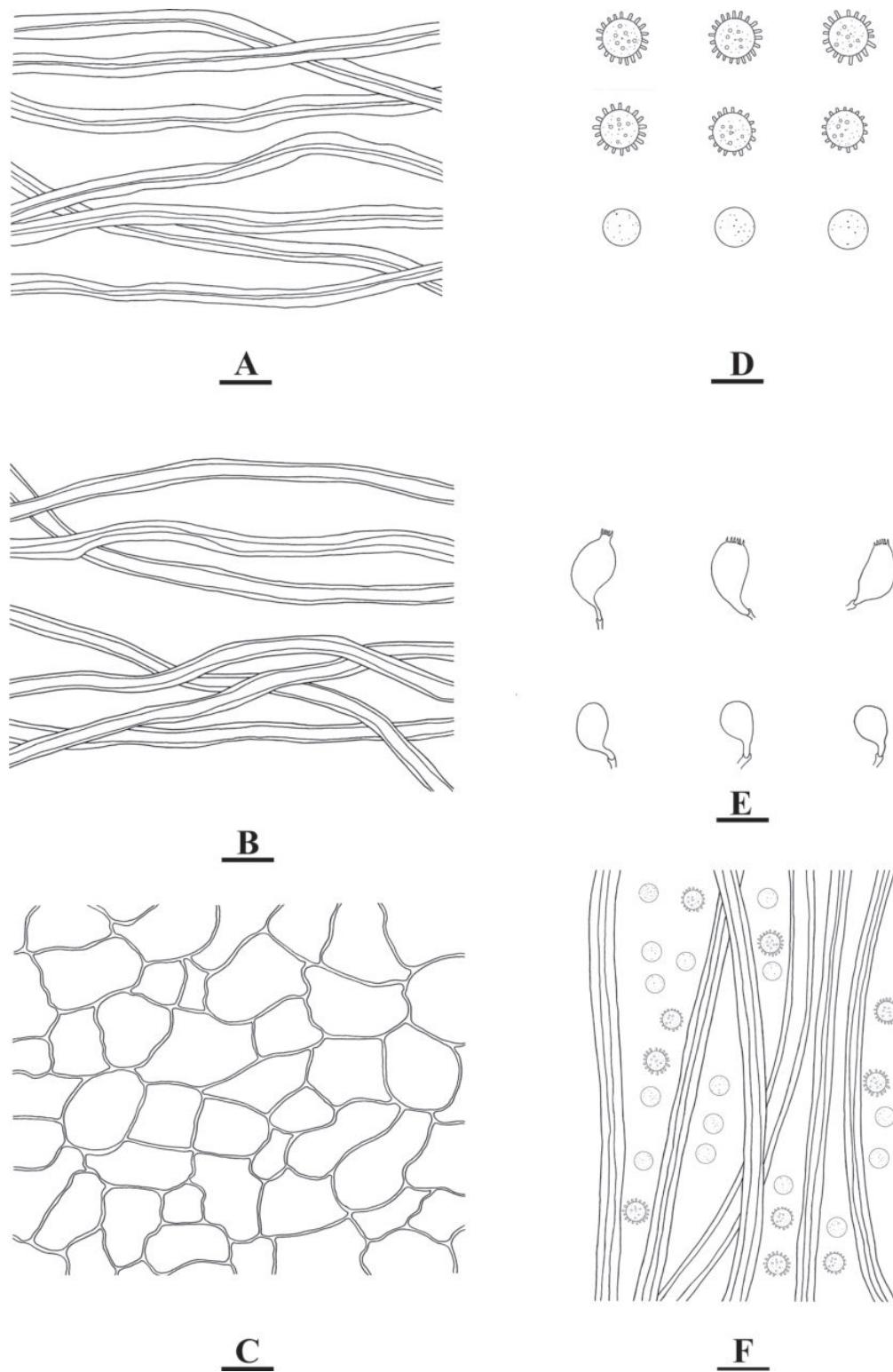
**Description. Fruiting body:** *Unexpanded basidiomata* 6–11 mm in diameter, 5–8 mm in height, pyriform to ellipsoidal, lightly pink to pinkish buff when fresh, pinkish buff to gray pink upon drying. *Expanded basidiomata* small sized, 0.8–2 cm in diameter, 0.7–1 cm in height. **Exoperidium:** shallowly saccate to deep saccate, dehiscence often less than halfway or halfway down, splits into 7–8 lobes at maturity, lobes 2–6 mm wide, tapered at the front end, rolled outwards, soft and thin upon drying. **Mycelial layer** gray pink, without debris, not easily dislodged. **Fibrous layer** white, tightly attached to the mycelial layer. **Pseudoparenchymatous layer** smooth surface, flesh-pink, easily exfoliation, dried thin, aseptic collar.

**Endoperidial body:** Globular, 4–6 mm in diameter, projecting apically, 1–2 mm length, sessile. **Endoperidium** slightly pink when fresh, pale mouse-gray upon drying, with a smooth surface under the dissecting microscope. **Peristome:** wide conical, fibrillose, lighter in color than the endoperidium, distinctly delimited.

**Hyphal structure:** *Capillitrial hyphae:* 2–5.5 µm in diameter, thick-walled, tawny, unbranched, IKI–, CB–, tissues unchanged in KOH. **Exoperidium mycelium layer** outside, mycelium hyphae thick-walled, 3–5 µm in diameter; **fibrous layer** in the middle, formed of the interlacing filament tissue, fibrous hyphae slightly thick-walled, 3–4.5 µm in diameter; **pseudoparenchymatous layer** inside, formed of the angular cell structured, 12–31 × 7–21 µm.



**Figure 2.** *Geastrum artocarpicola* (CLZhao 36079, CLZhao 39927) **A, D** the environment of *G. artocarpicola* **B, E** unexpanded basidiomata **C, F** basidiomata **G** peristome **H** basidia **I** basidiospores **J** pseudoparenchymatous layer **K** mycelium hyphae **L** capillitium hyphae.



**Figure 3.** *Gaeastrum artocarpicola* (CLZhao 36079) **A** mycelium hyphae **B** fibrous hyphae **C** pseudoparenchymatous layer **D** basidiospores **E** basidia **F** capillitium hyphae. Scale bars: 10 µm (**A–C, E, F**); 5 µm (**D**).

**Basidiospores:** Spherical, (3–)3.5–4(–4.6) × (3.1–)3.5–4(–4.6) µm in diameter, yellowish brown to dark brown, IKI–, CB–; spore surface smooth or ornamentation verrucose, 0.4–0.6(–0.8) µm in length,  $Q = 1.01$ ,  $Q_m = 1.01 \pm 0.01$ . **Basidia** 11–20 × 7–8 µm, bubble-shaped to flask shaped, with 6 inconspicuous basidial pedicels.

**Additional specimens examined (paratypes).** CHINA • Guangdong Province, Yangjiang, Jiangcheng District, Bitian Lake Park, 21°52'N, 111°58'E, elev. 27 m, on living tree of *Dimocarpus*, 30 July 2024, CLZhao 39927 (SWFC).

**Notes.** Based on ITS+nLSU+RPB1+ATP6 data (Fig. 1), the species *G. artocarpicola* was closely related to both species *G. mirabile* and *G. laneum*. The taxon *G. artocarpicola* resembles *G. laneum* and *G. mirabile* in sharing sessile of endoperidial body. However, *G. laneum* differs from *G. artocarpicola* due to its mycelial layer visible coarse short villus, and its delicately echinulate basidiospores surface (Wang and Bau 2023); *G. mirabile* can be distinguished from *G. artocarpicola* by its expanded basidiomata which has a mycoderm at the base, and capillitrial hyphae surface debris (Zhou et al. 2007).

***Gastrum fibulatum* X. Yang & C.L. Zhao, sp. nov.**

Mycobank No: 856265

Figs 4, 5

**Holotype.** CHINA • Jiangsu Province, Nanjing, Jiangning County, Diaoyutai Mountain, 31°58'N, 118°55'E, elev. 93.1 m, on the ground, 28 May 2024, CLZhao 36068 (SWFC).

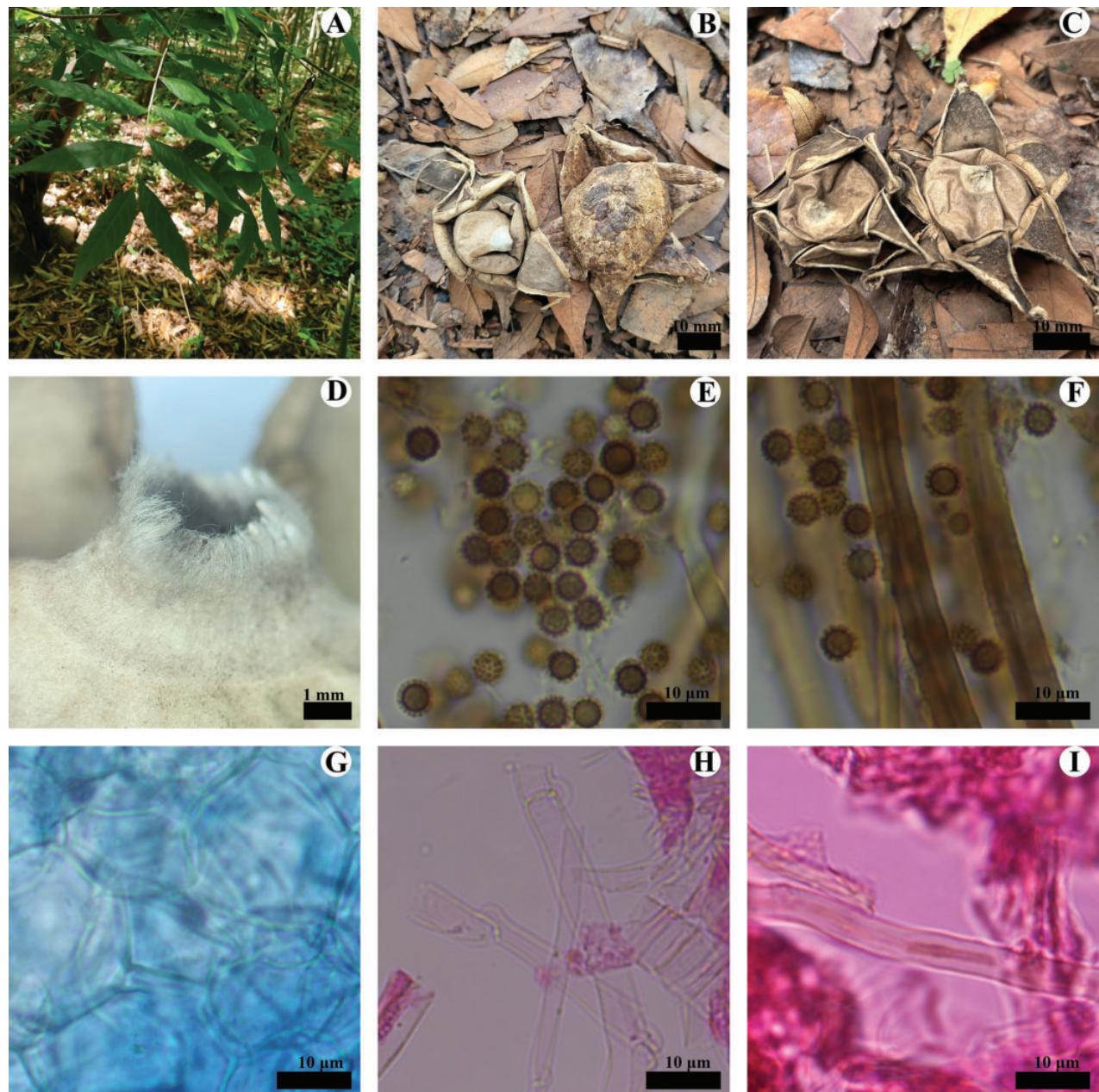
**Etymology.** *Fibulatum* (Lat.): referring to the generative hyphae with clamp connections of type specimen.

**Description. Fruiting body:** *Expanded basidiomata* small to medium-sized, 4–6 cm in diameter, 1–2.5 cm in height. **Exoperidium:** shallowly saccate to deep saccate, dehiscence often greater than halfway, splits into 5–6 lobes at maturity, lobes 8–18 mm wide, lobes long and mostly rolled outward, toughened and thin upon drying. **Mycelial layer** clay buff to grayish brown when dry, without debris, not easily dislodged. **Fibrous layer** white, tightly attached to the pseudoparenchymatous layer. **Pseudoparenchymatous layer** smooth surface, white to cream when fresh, cinnamon-buff to olivaceous to grayish brown to fuscous when dry, not deciduous, aseptic collar, thin when dry.

**Endoperidial body:** Globular, 15–25 mm in diameter, projecting apically, 1–5 mm length, sessile. **Endoperidium** of clay pink to clay buff, with a smooth surface and grayish villus visible under the dissecting microscope. **Peristome** silky fibrillose, color lighter than the endoperidium, distinctly delimited.

**Hyphal structure:** **Capillitrial hyphae** 3–6 µm in diameter, thick-walled, olivaceous buff, unbranched, IKI–, CB–; tissues unchanged in KOH. **Exoperidium mycelium layer** outside, dimitic hyphal system; generative hyphae with clamp connections, thin-walled, colorless, 3.5–6 µm in diameter, skeletal hyphae thick-walled to subsolid, slightly yellowish, 3–5.5 µm in diameter; **fibrous layer** in the middle, formed of the interlacing filament tissue, fibrous hyphae thick-walled to solid, 2.5–6 µm in diameter; **pseudoparenchymatous layer** inside, formed of the angular cell structured, 18–54 × 10–26 µm.

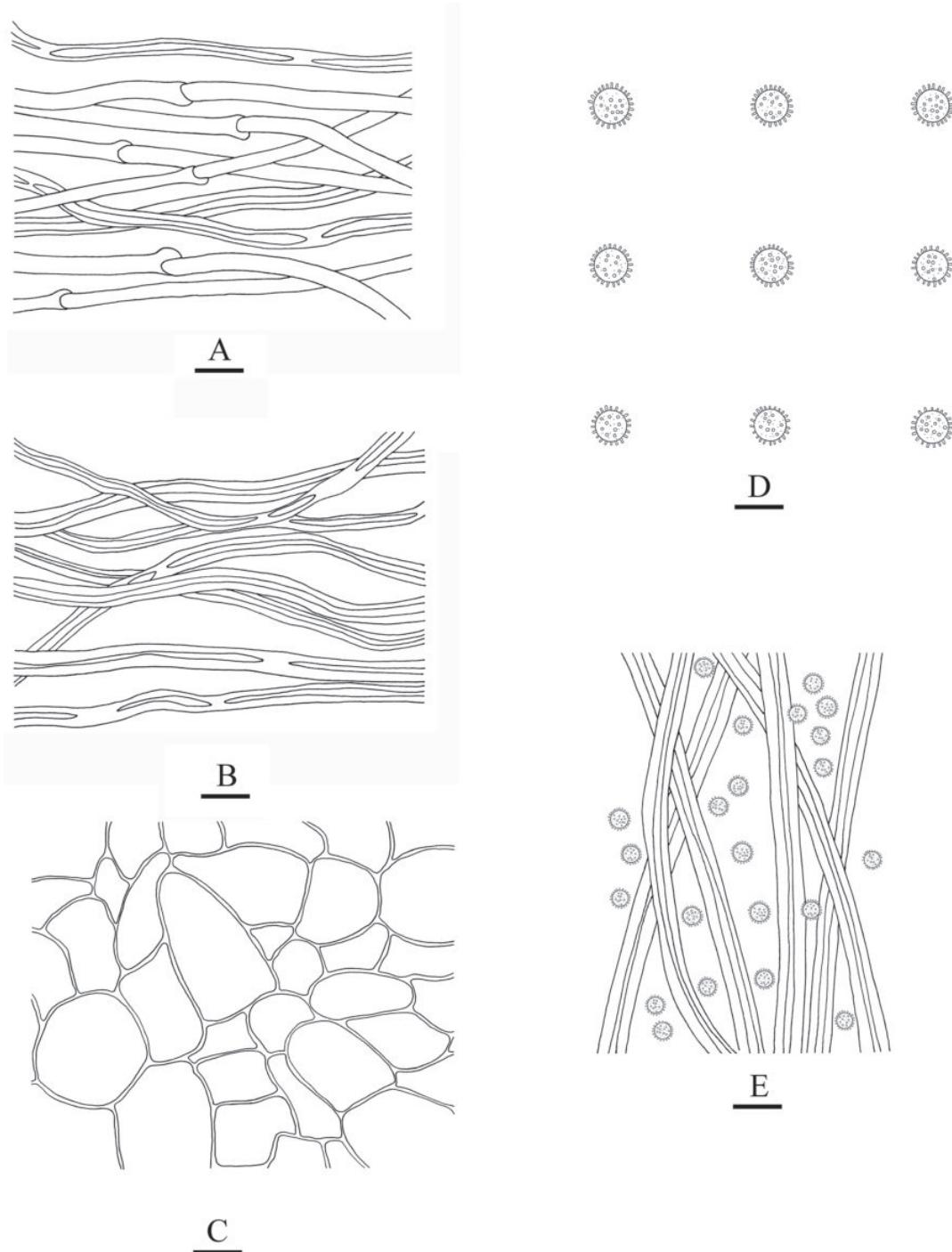
**Basidiospores:** Spherical, 3–3.6(–4) × 3–3.5(–4) µm in diameter, brown, thick-walled, to grayish brown, IKI–, CB–; spore surface with ornamentation verrucose, 0.5–0.8(–1) µm in length, Q = 1.01, Q<sub>m</sub> = 1.01 ± 0.01.



**Figure 4.** *Gaeastrum fibulatum* (CLZhao 36066, CLZhao 36067, CLZhao 36068, CLZhao 36069) **A** the environment of *G. fibulatum* **B, C** basidiomata **D** peristome **E** basidiospores **F** capillitium hyphae **G** pseudoparenchymatous layer **H, I** mycelium hyphae

**Additional specimens examined (paratypes).** CHINA • Jiangsu Province, Nanjing, Jiangning County, Diaoyutai Mountain, 31°58'N, 118°55'E, elev. 93.1 m, on the ground, 28 May 2024, CLZhao 36066; CLZhao 36067; CLZhao 36069(SWFC).

**Notes.** Based on dataset of ITS+nLSU+RPB1+ATP6 data (Fig. 1), *Gaeastrum fibulatum* was sister to *G. michelianum*. The taxon *G. fibulatum* resembles *G. fuscogleba* and *G. melanocephalum* in sharing verrucose basidiospores. However, *G. fuscogleba* differs from *G. fibulatum* by its arched expanded basidiomata, and its larger basidiospores (4.5–7 µm, Domínguez and Castellano 1996); the species *G. melanocephalum* can be distinguished from *G. fibulatum* by its endoperidial body with stalk, and arched expanded basidiomata (Jeppson et al. 2013).



**Figure 5.** *Gastrum fibulatum* (CLZhao 36068) **A** mycelium hyphae **B** fibrous hyphae **C** pseudoparenchymatous layer **D** basidiospores **E** capillital hyphae. Scale bars: 10 µm (A–C, E); 5 µm (D).

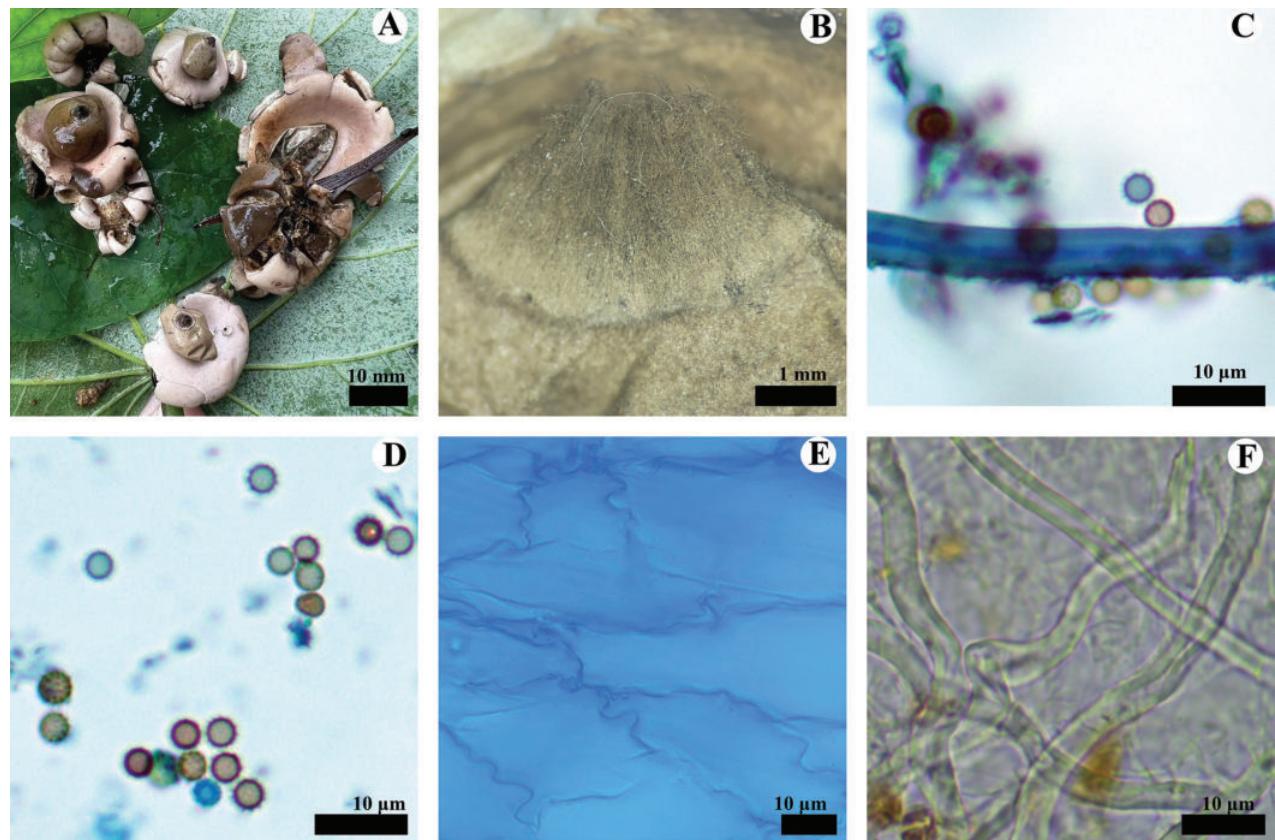
***Gastrum sinense* X. Yang & C.L. Zhao, sp. nov.**

Mycobank No: 856266

Figs 6, 7

**Holotype.** CHINA • Fujian Province, Xiamen, Siming District, Xiamen Botanical Garden, 24°27'N, 118°6'E, elev. 179 m, on the ground, 24 May 2024, CLZhao 36033 (SWFC).

**Etymology.** *Sinense* (Lat.): referring to the species being found in China.



**Figure 6.** *Geastrum sinense* (CLZhao 36029, CLZhao 36031, CLZhao 36032, CLZhao 36033, CLZhao 36038) **A** basidiomata **B** peristome **C** capillitium hyphae **D** basidiospores **E** pseudoparenchymatous layer **F** mycelium hyphae.

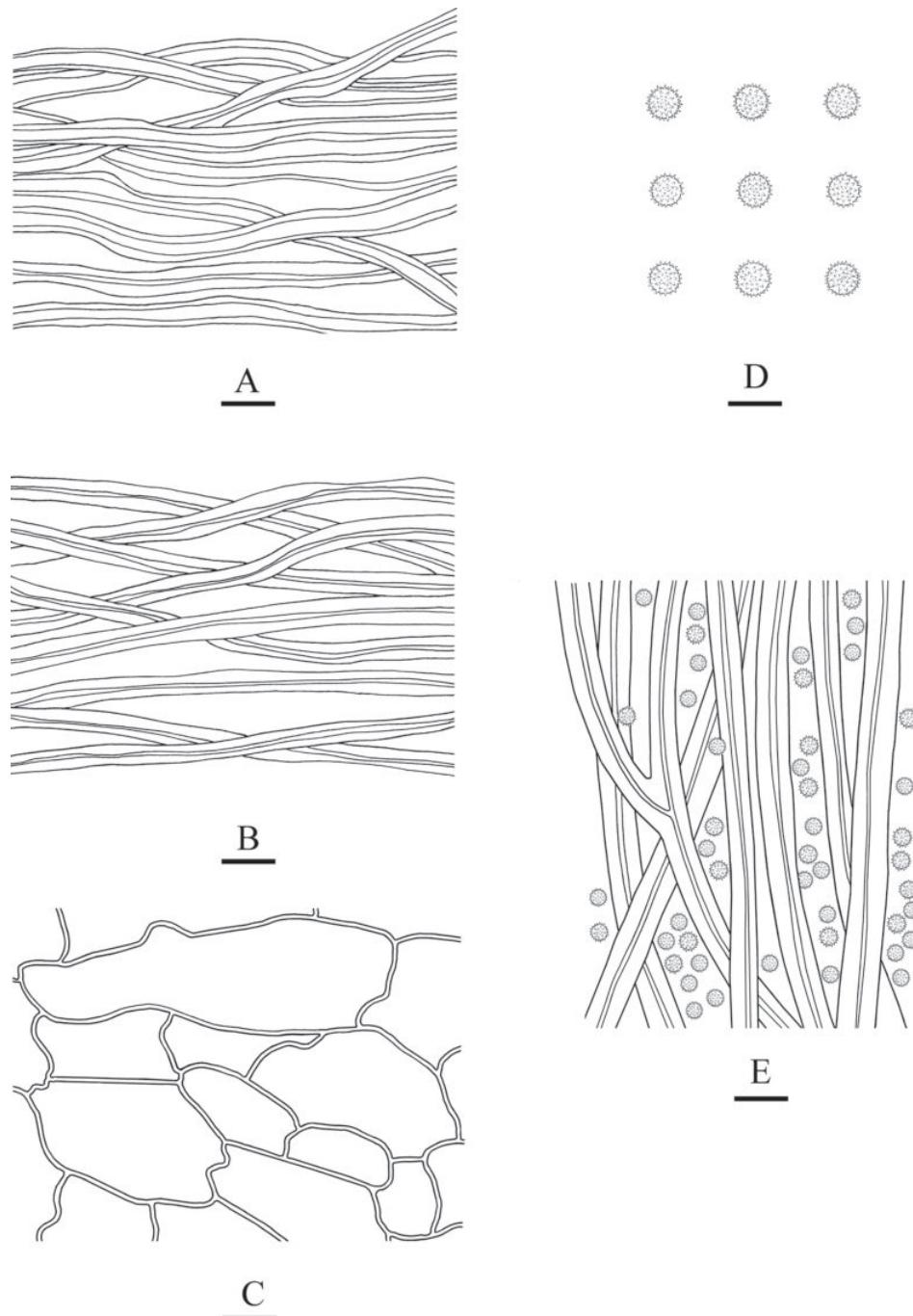
**Description.** **Fruiting body:** *Expanded basidiomata* medium sized, 10–25 mm in height, 15–40 mm in diameter. **Exoperidium:** arched, dehiscence often half-way down, splits into 5–9 lobes at maturity, lobes 3–10 mm wide, mostly rolled outward to under the outer exoperidial disc, tapered at the front end, turn to soft and thin upon drying. **Mycelial layer:** pinkish buff, without debris, not easily dislodged. **Fibrous layer:** white, tightly attached to the mycelial layer. **Pseudoparenchymatous layer:** smooth surface, cream to pink, not deciduous, aseptic collar when fresh, clay-buff to grayish brown, thinner and soft when dry.

**Endoperidial body:** Globular, 8–15 mm in diameter, projecting apically, 1–3 mm length, sessile. **Endoperidium** pale to dark brown. **Peristome** broadly conical, fibrillose, dark brown to gray, distinctly delimited.

**Hyphal structure:** **Capillitium hyphae:** up to 4.5–6.5  $\mu\text{m}$  in diameter, thick-walled to subsolid, brown, occasionally branched, IKI–, CB–; tissues unchanged in KOH. **Exoperidium mycelium layer** outside, mycelium hyphae slightly thick-walled to thick-walled, 4–5  $\mu\text{m}$  in diameter. **fibrous layer** in the middle, formed of the interlacing filament tissue, fibrous hyphae slightly thick-walled, 3–4.5  $\mu\text{m}$  in diameter; **pseudoparenchymatous layer** inside, formed of the angular cell structured, 20–75  $\times$  13–25  $\mu\text{m}$ ;

**Basidiospores:** Spherical, 2.6–3(–3.5)  $\times$  2.5–3(–3.5)  $\mu\text{m}$  in diameter, yellowish brown to dark brown, IKI–, CB–; spore surface with echinulate, 0.4–0.8  $\mu\text{m}$  in length,  $Q = 1.01$ ,  $Q_m = 1.01 \pm 0.01$ .

**Additional specimens examined (paratypes).** CHINA • Fujian Province, Xiamen, Siming, Xiamen Botanical Garden, 24°27'N, 118°6'E, elev. 179 m, on the



**Figure 7.** *Gastrum sinense* (CLZhao 36033) **A** mycelium hyphae **B** fibrous hyphae **C** pseudoparenchymatous layer **D** basidiospores **E** capillitatal hyphae. Scale bars: 10 µm (A–C, E); 5 µm (D).

ground, 24 May 2024, CLZhao 36029; CLZhao 36031; CLZhao 36032 and CLZhao 36038 (SWFC).

**Notes.** Based on ITS+nLSU+RPB1+ATP6 data (Fig. 1), *G. sinense* was sister to *G. melanorhynchum*. The *G. sinense* resembles *G. oxysepalum* and *G. floriforme* in sharing sessile endoperidial body. However, *G. oxysepalum* differs from *G. sinense* by its shallowly saccate expanded basidiomata, and non-constant peristomal ring (Wang and Bau 2023); *G. floriforme* can be distinguished from *G. sinense* by its deep saccate expanded basidiomata, and larger basidiospores (5.5–7 µm, Zhou et al. 2007)

***Gastrum trachelium* X. Yang & C.L. Zhao, sp. nov.**

Mycobank No: 856267

Figs 8, 9

**Holotype.** CHINA • Guangdong Province, Guangzhou, Huangpu District, Shuangchuanshi Mts., 23°11'N, 113°32'E, elev. 69.6 m, on the fallen angiosperm leaves, 14 May 2024, CLZhao 35992 (SWFC).

**Etymology.** *Trachelium* (Lat.): referring to the species having a long stipe.

**Description.** Fruiting body: **Unexpanded basidiomata**, 9–12 mm in diameter, ellipsoidal to fusiform, white to pink. **Expanded basidiomata** small to medium sized, 2–3 cm in diameter, 1.5–2 cm in height, and long stipe (height 0.5–1 cm). **Exoperidium**: deep saccate, dehiscence often halfway down, splits into 5–6 lobes at maturity, lobes 5–10 mm wide, tapered at the front end, exoperidium attached to the rhizomorphs, soft and thin upon drying. **Mycelial layer**: cream to clay-buff when fresh, without debris, not easily dislodged, turning to clay-buff when dry. **Fibrous layer**: white, tightly attached to the mycelial layer. **Pseudoparenchymatous layer**: smooth surface, cream to flesh pink when fresh, not deciduous, aseptic collar, turning to gray brown, thinner when dry.

**Endoperidial body:** Globular, 10–13 mm in diameter, projecting apically, 1–3 mm length, sessile. **Endoperidium** ash-gray, with a smooth surface and buff villus visible under the dissecting microscope. **Peristome** wide conical, fibrillose, ash-gray to dark gray, distinctly delimited.

**Hyphal structure:** **Capillital hyphae**: 3–4.5 µm in diameter, thick-walled, slightly yellowish, unbranched, IKI–, CB–; tissues unchanged in KOH. **Exoperidium mycelium layer** outside, mycelium hyphae slightly thick-walled to solid, 2.2–3.1 µm in diameter; **fibrous layer** in the middle, formed of the interlacing filament tissue, fibrous hyphae slightly thick to thick-walled, 2.5–3.3 µm in diameter; **pseudoparenchymatous layer** inside, formed of the angular cell structured, 22–56 × 14.5–33 µm.

**Basidiospores:** Spherical, (3.5–)3.7–4.3(–4.6) × (3.5–)3.8–4.3(–4.6) µm in diameter, grayish brown to dark brown, IKI–, CB–; spore surface with delicately echinulate, 0.4–0.9 µm in length,  $Q = 1.01$ ,  $Q_m = 1.01 \pm 0.01$ . **Basidia** flask shaped, 13.5–18.5 × 6–9 µm, with 6 inconspicuous basidial pedicels.

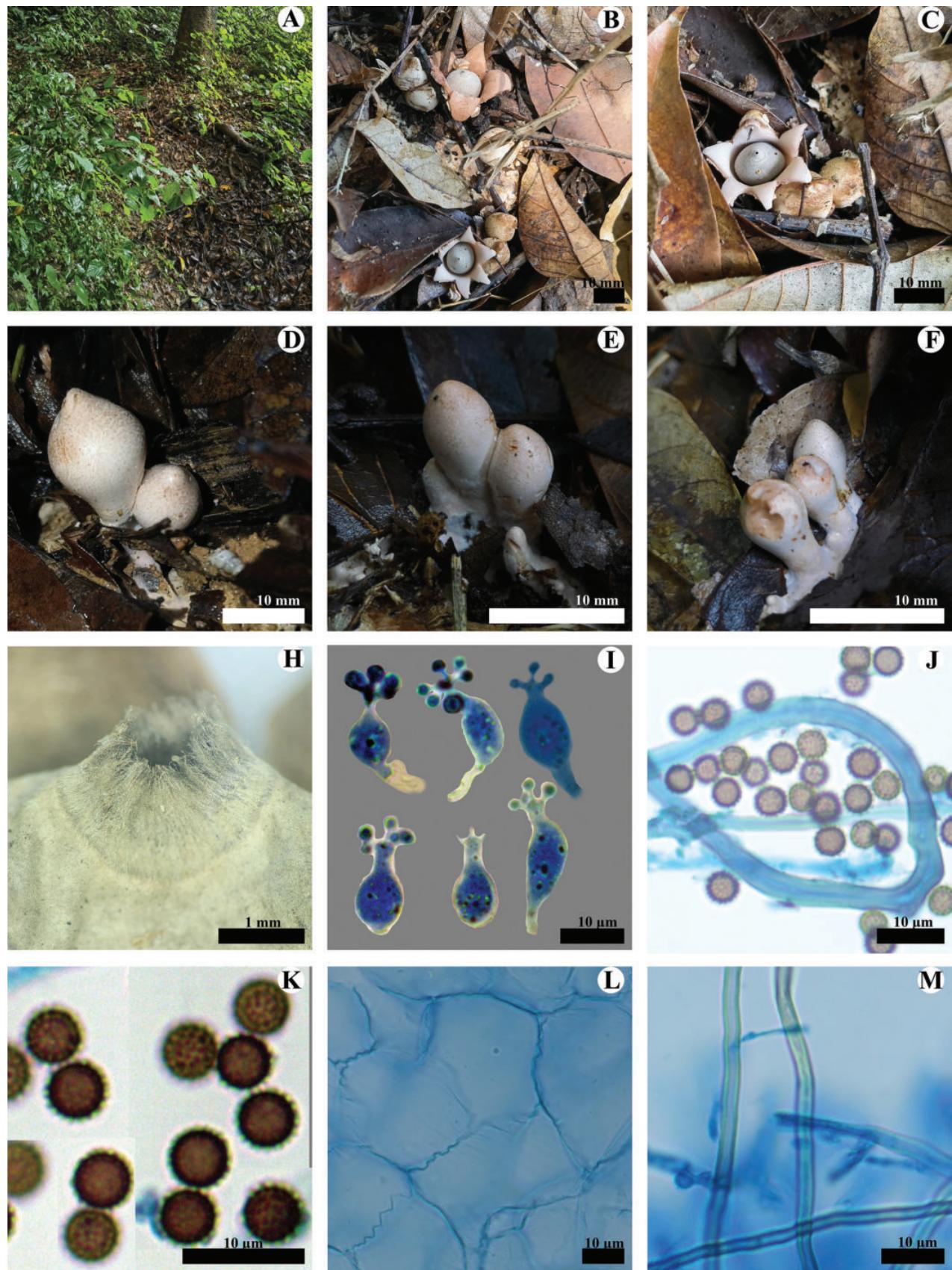
**Additional specimens examined (paratypes).** CHINA • Guangdong Province, Guangzhou, Huangpu, Shuangchuanshi Mts., 23°11'N, 113°32'E, elev. 69.6 m, on the fallen angiosperm leaves, 9 May 2024, CLZhao 35939; 1 June 2024, CLZhao 36055(SWFC).

**Notes.** Based on ITS+nLSU+RPB1+ATP6 data (Fig. 1), *G. trachelium* was sister to *G. suae*. The species *G. trachelium* resembles *G. yanshanense* and *G. hirsutum* in sharing sessile endoperidial body. However, *G. yanshanense* differs from *G. trachelium* by its smaller basidiospores (2.7–3.2 × 2.8–3.3 µm, Zhou et al. 2022); *G. hirsutum* can be distinguished from *G. trachelium* by its subglobose to obovate unexpanded basidiomata, and smaller basidiospores (2.5–3 µm, Baseia and Calonge 2006).

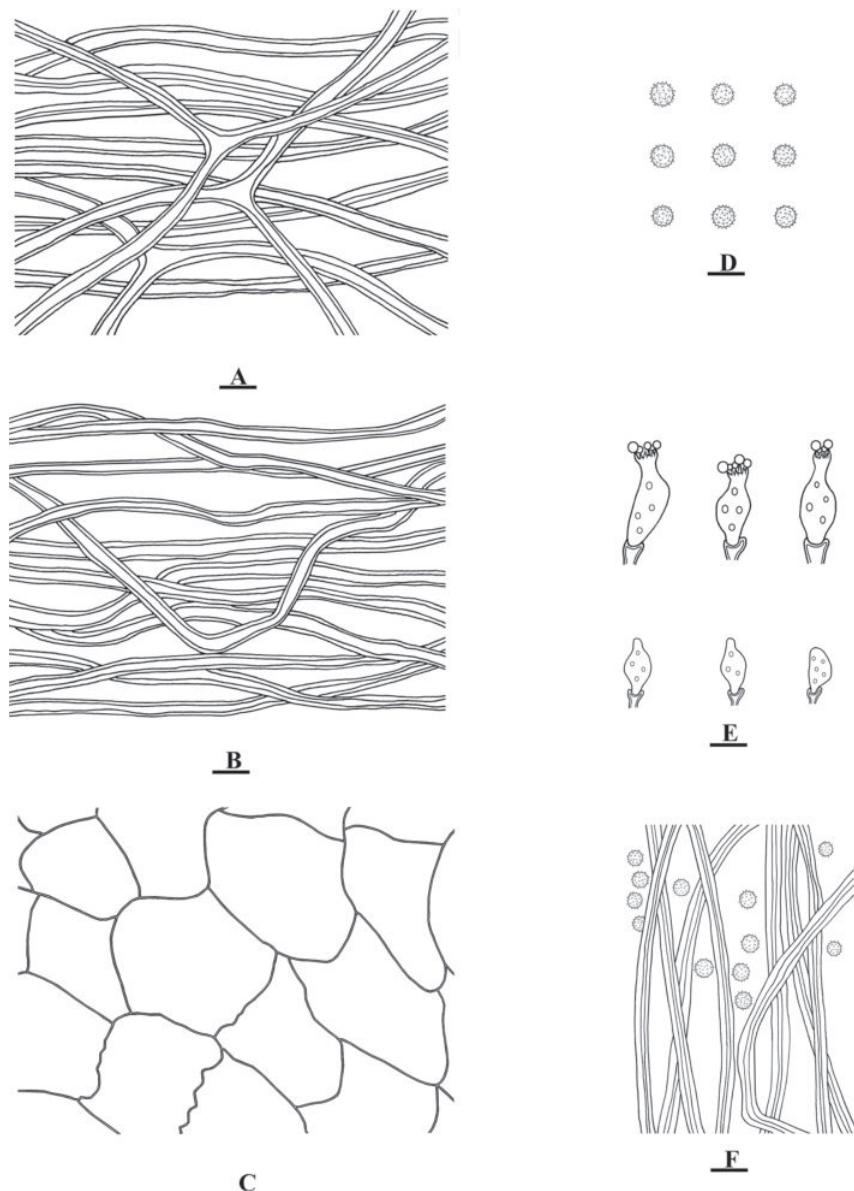
***Gastrum beijingense* C.L Hou, Hao Zhou & Ji Qi Li, Mycosistema 41: 11 (2022)**

Mycobank No: 570850

**Typus.** China, Beijing, Yanqing District, Zhangjiaying Village, BJTC 248 (holotype).



**Figure 8.** *Geastrum trachelium* (CLZhao 35939, CLZhao 35992, CLZhao 36055) **A** the environment of *G. trachelium* **B, C** basidiomata **D, E, F** unexpanded basidiomata **G** peristome **H** basidia **I** capillital hyphae **J** basidiospores **K** pseudoparenchymatous layer **L** mycelium hyphae.



**Figure 9.** *Geastrum trachelium* (CLZhao 35992) **A** mycelium hyphae **B** fibrous hyphae **C** pseudoparenchymatous layer **D** basidiospores **E** basidia **F** capillitium hyphae. Scale bars: 10 µm (**A–C, E, F**); 5 µm (**D**).

***Geastrum sanglinense* Y.Q. Wu & Shu R. Wang, Mycoscience 65: 13 (2024).**

**Synonym**

Mycobank No: 845733

**Typus.** China, Shanxi Province, Yangcheng County, the Manghe Rhesus Monkey National Nature Reserve, located near Huiquan Village, HMSAU 15020 (holotype)

**Note.** Our results show that the types of *Geastrum beijingense* and *G. sanglinense* represent a single species, because *G. beijingense* has the priority (Zhou et al. 2022), and *G. sanglinense* is treated as synonym of *beijingense*. In addition, the type localities of these two taxa are very close with similar forest and climate. In the phylogenetic tree based on ITS+LSU+rpb1+atp6 (Fig. 1), both *G. beijingense* and *G. sanglinense* are nested in a lineage.

## Discussion

In the present study, four new species *Gastrum artocarpicola*, *G. fibulatum*, *G. sinense* and *G. trachelium* are described, based on the phylogenetic analyses and morphological characteristics.

The integrative taxonomy inferred from the ITS and LSU nrDNA, RPB1, and ATP6 revealed an unexpected diversity in the genus *Gastrum* (Gastrales, Basidiomycota). A total of 27 lineages was proposed and a revised taxonomy of the combination of the different sources of taxonomic information was presented (Zamora et al. 2015). Phylogenetically, four new species were nested in the genera *Gastrum* within the order Gastrales, in which *G. fibulatum* was sister to *G. michelianum*, and the *G. sinense* was closely related to *G. melanorhynchum*, and the *G. trachelium* was sister to *G. suae*, and the *G. artocarpicola* formed a monophyletic lineage. However, morphologically, *G. michelianum* differs from *G. fibulatum* by having the arched of the fruitbody, and the collar pseudoparenchymatous layer at the base of endoperideal body (Muñoz Sánchez 2020). The species *G. melanorhynchum* differs from *G. sinense* by having the shallowly saccate fruiting body, and larger basidiospores (3.5–3.9 µm) (Wang and Bau 2023). The taxon *G. suae* differs from *G. trachelium* by having larger unexpanded basidiomata (13–28 mm), and high expanded basidiomata (35–70 mm) in, and larger basidiospores (4.5–6 × 5–6 µm; Zhang et al. 2023b).

Morphologically, our new species resemble 16 similar species in the *Gastrum*, individually as *G. fuscoglebum* (Zeller) Jeppson & E. Larss., *G. michelianum*, *G. melanocephalum*, *G. smardae*, *G. triplex*, *G. melanorhynchum*, *G. oxysepalum* T. Bau & Xin Wang, *G. floriforme*, *G. suae*, *G. yanshanense*, *G. hirsutum* Baseia & Calonge, *G. brunneocapillatum* J.O. Sousa, Accioly, M.P. Martín & Baseia, *G. laneum*, *G. mirabile*, *G. pleosporum* and *G. schweinitzii*. A morphological comparison among our new species and sixteen similar species are presented in Table 2.

Macrofungi are an important part of forest ecosystems, which are mainly composed of most members of Basidiomycota and some members of Ascomycota, and they possess important economic value and ecological functions (Wu et al. 2019, 2022; Dai et al. 2021; Deng and Zhao 2023; Guan et al. 2023; Yuan et al. 2023b; Zhang et al. 2023a, 2024; Dong et al. 2024a, b; Luo et al. 2024). The family Geastraceae is an extensively studied group of Basidiomycota (Zhou et al. 2007; Finy et al. 2021; Wang and Bau 2023). However, the *Gastrum* species diversity in China is still not well known, especially in the subtropical and tropical areas. Therefore, a multidisciplinary approach, combining taxonomists, molecular biologists and field practitioners, is required. Edible and medicinal fungi have huge potential as food and medicines, especially in Asia and their prospects (Dai and Yang 2008; Dai et al. 2010; Cui et al. 2023; Li et al. 2023; Xie et al. 2024). In the present study, four new species are introduced from China, which will further enrich our knowledge of the macrofungal diversity. We anticipate that more undescribed *Gastrum* taxa will be discovered throughout China after extensive collection combined with morphological and molecular analyses.

**Table 2.** A morphological comparison among four new *Gastrum* species and other similar species.

Species name	Unexpanded basidiomata	Expanded basidiomata	Endoperidial body	Basida	Basidiospores	References
<i>Gastrum artocarpicola</i>	5–8 mm high, 6–11 mm diam; pyriform to ellipsoidal	Shallowly saccate to deep saccate; 7–10 mm high, 8–20 mm diam; 7–8 rayes	Sessile; 4–6 mm diam; obvious oral margin ring	bubble-shaped to flask shaped; 11–20 × 7–8 µm.	Spherical; 3.5–4 × 3.5–4 µm diam	Present study
<i>Gastrum brunneocapillatum</i>	6–10 mm high, 7–13 mm diam; obpyriform to oval	Sacate; 4.1–11 mm high, 8–26 mm diam; 5–7 rayes	Sessile; 15–25 mm diam; obvious oral margin ring	Thin-walled; clavate; pyriform to lageniform; 12.2–19.3 × 3.8–6.7 µm.	Globose to subglobose; 2.8–4 × 2.7–4 µm diam	Accioly et al. (2019)
<i>Gastrum fibulatum</i>	—	Shallowly saccate to deep saccate; 10–25 mm high, 40–60 mm diam; 5–6 rayes	Sessile; 15–25 mm diam; obvious oral margin ring	—	Spherical; 3–3.6 × 3–3.5 µm diam	Present study
<i>Gastrum floriforme</i>	6–10 mm diam; subglobose	Deep saccate; 6–14 mm diam; 6–11 rayes	Sessile; 4–11 mm diam; non-constant peristomal ring	—	Globose to subglobose; 5.5–7 µm diam	Zhou et al. (2007)
<i>Gastrum fuscogleba</i>	10–50 mm diam; globose to subglobose	3–4 rayes	12–20 mm diam	Lecythiform to lageniform; 6 sporde; 11 × 7 µm	Globose; 4.5–7 µm diam	Domínguez and Castellano (1996)
<i>Gastrum hirsutum</i>	5–10 mm high, 4–8 mm diam; subglobose to obovate	Saccate; 15–20 mm diam when open; 5–7 rayes	Sessile; 4–6 mm diam; obvious oral margin ring	—	Globose; 2.5–3 µm diam	Baseia and Calonge (2006)
<i>Gastrum laneum</i>	3–10 mm in size	Shallowly saccate; 4.5–9.5 mm high; 5–7 rayes	Sessile; 2–7 mm diam	—	Spherical; 2.5–3.9 µm diam	Wang and Bau (2023)
<i>Gastrum melanocephalum</i>	—	Arched; 40–200 mm diam; 5–8 rayes	Stalk; 25–60 mm diam; peristome lacking	—	Globose; 3.5–4.5 µm diam	Jeppson et al. (2013)
<i>Gastrum melanorhynchum</i>	—	Shallowly saccate, arched; 12–35 mm diam; 7–9 rayes	Sessile; 6–20 mm diam; obvious oral margin ring	Clavate, sublageniform; 4 sporde; 14.4–19.7 × 9.1–11.4 µm	Spherical; 3.5–3.9 µm diam	Wang and Bau (2023)
<i>Gastrum michelianum</i>	—	Arched; up to 120 mm diam; 5–6 rayes	Up to 40 mm diam; obvious oral margin ring	—	Globose; up to 6 µm diam	Muñoz (2020)
<i>Gastrum mirabile</i>	5–10 mm high, 4–8 mm diam; globose, subglobose, obovate	Shallowly saccate to deep saccate; 6–20 mm in high; 5–7 rayes	Sessile; 3–9 mm diam	—	Subglobose to obovate; 2.5–4 µm diam	Zhou et al. (2007)
<i>Gastrum oxysepalum</i>	13–16 mm diam	shallowly saccate; 12–17 mm diam; 5–8 rayes	Sessile; 6–13 mm diam; non-constant peristomal ring	—	Spherical; 2.7–3.9 µm diam	Wang and Bau (2023)
<i>Gastrum pleosporum</i>	12–15 mm diam; globose to depressed globose	5–6 rayes	Sessile; 5–8 mm diam	Clavate, ventricose to flask-shaped with a more or less long collar; 4–8 spored; 15.5–19 × 4–5.5 µm	Globose, cylindrical, elliptic, reniform, club-shaped; 4–6 × 4–5 µm	Douanla-Meli et al. (2005)
<i>Gastrum schweinitzii</i>	oval	Deep saccate; 7–9 mm high × 12–20 mm diam; 5–8 rayes	Sessile; 3–9 mm diam; spherical or oblate spherical; obvious oral margin ring	—	Spherical; 2–3.6 × 2–3.4 µm	Han and Bau (2015)
<i>Gastrum sinense</i>	—	Arched; 10–25 mm high, 15–40 mm diam; 5–9 rayes	Sessile; 8–15 mm diam; obvious oral margin ring	—	Spherical; 2.6–3 × 2.5–3 µm	Present study
<i>Gastrum smardae</i>	—	Pseudofoznicate; 40–60 mm diam; 7–9 rayes	Shortly stalked; 15–20 mm diam; obvious oral margin ring	—	Globose; 3–4.5 µm diam	Jeppson et al. (2013)
<i>Gastrum suae</i>	13–28 mm high; cylindrical to ellipsoidal	Deep saccate; 35–70 mm high, 18–37 mm diam; 6 rayes; exoperidium attached to the rhizomorphs	Sessile; 11–23 mm high; obvious oral margin ring	—	Globose; 4.5–6 × 5–6 µm	Zhang et al. (2022)

Species name	Unexpanded basidiomata	Expanded basidiomata	Endoperidial body	Basida	Basidiospores	References
<i>Gastrum trachelium</i>	9–12 mm diam; ellipsoidal to fusiform	Deep saccate; 15–20 mm high, 20–30 mm diam; 5–6 rays; exoperidium attached to the rhizomorphs	Sessile; 10–13 mm diam; obvious oral margin ring	Flask shaped; 13.5–18.5 × 6–9 µm; 6 sporae	Spherical; 3.7–4.3 × 3.8–4.3 µm	Present study
<i>Gastrum triplex</i>	14–22 mm diam and 15–25 mm high; onion shaped to obpyriform	Saccate; 35–50 mm diam; 5–7 rays	Sessile; 10–18 mm diam; obvious oral margin ring	—	Globose; 3.6–4.2 × 3.3–4.1 µm diam	Vishal et al. (2023)
<i>Gastrum yanshanense</i>	9–15 mm high; spherical to ellipsoidal	Deep saccate; 15–25 mm high; 5–7 rays; exoperidium attached to the rhizomorphs	Sessile; 8–13 mm high; obvious oral margin ring	Bubble-shaped to flask shaped; 8.1–9 × 12.6–13.5 µm; with 4–7 sporae	Globose to ellipsoid; 2.7–3.2 × 2.8–3.3 µm	Zhou et al. (2021)

## Additional information

### Conflict of interest

The authors have declared that no competing interests exist.

### Ethical statement

No ethical statement was reported.

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### Author contributions

Conceptualization, CZ; methodology, CZ and XY; software, CZ, XY and YZ; validation, CZ and XY; formal analysis, CZ and XY; investigation, CZ, XW and XY; resources CZ; writing—original draft preparation, CZ, JD, YZ, SD and XW; writing—review and editing, CZ and XY; visualization, CZ and XY; supervision, CZ and XY; project administration, CZ; funding acquisition, CZ. All authors have read and agreed to the published version of the manuscript.

### Author ORCIDs

Xin Yang  <https://orcid.org/0009-0009-7755-0581>

Yonggao Zhu  <https://orcid.org/0009-0008-5341-3798>

Songjing Duan  <https://orcid.org/0009-0005-3122-0318>

Xingxing Wu  <https://orcid.org/0009-0001-1497-8937>

Changlin Zhao  <https://orcid.org/0000-0002-8668-1075>

### Data availability

All of the data that support the findings of this study are available in the main text.

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